Effects of 6-month vitamin D supplementation on insulin sensitivity and secretion: a randomised, placebo-controlled trial

Patricia Lemieux¹, S John Weisnagel^{1,2}, Annabelle Z Caron¹, Anne-Sophie Julien³, Anne-Sophie Morisset^{2,4,5}, Anne-Marie Carreau¹, Jonathan Poirier¹, André Tchernof^{2,4,5,6}, Julie Robitaille^{2,4,5}, Jean Bergeron^{1,2}, André Marette^{1,2,5,6}, Marie-Claude Vohl^{4,5} and Claudia Gagnon^{1,2,5}

¹Department of Medicine, Université Laval, Québec City, Québec, Canada, ²Endocrinology and Nephrology Unit, CHU de Québec-Université Laval Research Centre, Québec City, Québec, Canada, ³Department of Mathematics and Statistics, Québec City, Québec, Canada, ⁴School of Nutrition, Laval University, Québec City, Québec, Canada, ⁵Institute of Nutrition and Functional Foods, Québec City, Québec, Canada, and ⁶Quebec Heart and Lung Institute Research Centre, Québec City, Québec, Canada

Correspondence should be addressed to C Gagnon

Email
claudia.gagnon@crchudequebec.ulaval.ca

Abstract

Objective: To determine whether vitamin D₃ supplementation improves insulin sensitivity, using the hyperinsulinemic-euglycemic clamp.

Design: This single-centre, double-blind, placebo-controlled trial randomised 96 participants at high risk of diabetes or with newly diagnosed type 2 diabetes to vitamin D₃ 5000 IU daily or placebo for 6 months.

Methods: We assessed at baseline and 6 months: (1) primary aim: peripheral insulin sensitivity (M-value using a 2-h hyperinsulinemic-euglycemic clamp); (2) secondary aims: other insulin sensitivity (HOMA2%S, Matsuda) and insulin secretion (insulinogenic index, C-peptide area under the curve, HOMA2-B) indices using a 2-h oral glucose tolerance test (OGTT); β -cell function (disposition index: M-value $\times$ insulinogenic index); fasting and 2-h glucose post OGTT; HbA_{1c}; anthropometry.

Results: Baseline characteristics were similar between groups (% or mean $\pm$ s.d.): women 38.5%; age 58.7 $\pm$ 9.4 years; BMI 32.2 $\pm$ 4.1 kg/m²; prediabetes 35.8%; diabetes 20.0%; 25-hydroxyvitamin D (25(OH)D) 51.1 $\pm$ 14.2 nmol/L. At 6 months, mean 25(OH)D reached 127.6 $\pm$ 26.3 nmol/L and 51.8 $\pm$ 16.5 nmol/L in the treatment and placebo groups, respectively ($P < 0.001$). A beneficial effect of vitamin D₃ compared with placebo was observed on M-value (mean change (95% CI): 0.92 (0.24–1.59) vs –0.03 (–0.73 to 0.67); $P = 0.009$) and disposition index (mean change (95% CI): 267.0 (–343.4 to 877.4) vs –55.5 (–696.3 to 585.3); $P = 0.039$) after 6 months. No effect was seen on other outcomes.

Conclusions: In individuals at high risk of diabetes or with newly diagnosed type 2 diabetes, vitamin D supplementation for 6 months significantly increased peripheral insulin sensitivity and β -cell function, suggesting that it may slow metabolic deterioration in this population.

European Journal of
Endocrinology
(2019) **181**, 287–299

Introduction

Low serum 25-hydroxyvitamin D (25(OH)D) has been associated with an increased risk of developing insulin resistance and type 2 diabetes in prospective observational studies (1, 2). Since low vitamin D status is highly

prevalent worldwide (3), the potential role of vitamin D supplementation in improving glucose homeostasis generated great enthusiasm among scientists and clinicians. However, randomised controlled trials (RCTs)

of vitamin D supplementation have shown inconstant effects on measures of insulin sensitivity, insulin secretion and β -cell function. A meta-analysis concluded that variable results among RCTs could be explained by heterogeneous study populations in terms of ethnicity, glucose tolerance and vitamin D status, by variations in vitamin D dosage and duration of treatment and by use of surrogate measures of insulin sensitivity (4, 5). Yet, the five studies that used the gold-standard method to assess insulin sensitivity, the hyperinsulinemic-euglycemic clamp, found no effect of vitamin D supplementation on insulin sensitivity (6, 7, 8, 9, 10). However, three of these trials were limited by small sample size ($n=12-18$) (6, 7, 8). Despite adequate sample size ($n=62-65$) and robust methodology, the two other studies failed to show benefits of vitamin D in metabolically healthy overweight or obese subjects (10) and in individuals with long-standing type 2 diabetes (9).

The primary aim of this 6-month trial was to determine whether vitamin D₃ supplementation improves peripheral insulin sensitivity (M-value) in individuals at high risk for type 2 diabetes or with newly diagnosed type 2 diabetes, using the gold-standard hyperinsulinemic-euglycemic clamp. Secondary aims were to evaluate the effects of vitamin D₃ on: fasting- and OGTT-derived indices of insulin sensitivity, insulin secretion, β -cell function, metabolic markers, blood pressure and anthropometric measurements.

Methods

Study design

This randomised, double-blind, placebo-controlled, parallel-group trial was conducted at a single site (Centre Hospitalier Universitaire (CHU) de Québec-Université Laval) located in Québec City, Canada. After screening, eligible participants were randomly assigned, in a 1:1 ratio, to receive vitamin D₃ or placebo for 6 months. Randomisation, computerised based on the Pocock and Simon minimisation method (11), was conducted by a biostatistician to balance treatment arms according to sex, BMI ($<$ or ≥ 30 kg/m²) and age ($<$ or ≥ 50 years old). The biostatistician sent treatment allocation directly to the pharmacist who was in charge of dispensing the study medication. The investigators, research personnel, care providers and participants involved in this study were blinded to the treatment assignment until the last participant completed the study.

The study protocol and all amendments were approved by the CHU de Québec-Université Laval Ethics Committee. The study was registered at clinicaltrials.gov before starting recruitment (NCT01779908) and conduct of the trial followed the 2010 Consort guidelines (12). This study was also conducted according to the guidelines laid down in the Declaration of Helsinki and all procedures involving patients were approved by the CHU de Québec-Université Laval Ethics Committee (Project number B12-12-1095). Written informed consent was obtained from each participant before any trial-related activities.

Participants

Participants were recruited between January 2013 and October 2016 via flyers in hospitals and public places, through newspapers and radio advertisements and via emails sent to employees of the CHU de Québec-Université Laval and Université Laval. Initially, eligible participants included vitamin D-deficient (25(OH)D ≤ 50 nmol/L) Caucasian men and postmenopausal women aged ≥ 50 years, meeting the criteria of the hypertriglyceridemic waist phenotype (waist circumference ≥ 90 cm plus fasting triglycerides ≥ 2.0 mM in men or waist circumference ≥ 85 cm plus fasting triglycerides ≥ 1.5 mM in women). During the trial, inclusion criteria were modified a few times to improve the recruitment rate. Eligible participants finally included Caucasian men and women aged ≥ 25 years with a serum 25(OH)D at screening ≤ 55 nmol/L, an increased waist circumference (≥ 102 cm for men and ≥ 88 cm for women) and at least one metabolic abnormality or risk factor associated with insulin resistance. Those were defined as (1) fasting triglycerides ≥ 1.7 mM or dyslipidemia treated with lipid-lowering drugs, (2) prediabetes or type 2 diabetes not on glucose-lowering drugs: fasting glucose ≥ 5.6 mM or 2-h glucose post OGTT ≥ 7.8 mM or HbA_{1c} $\geq 5.7\%$, (3) first-degree relative with type 2 diabetes and (4) history of gestational diabetes.

Participants were excluded if they had any of the following conditions: type 2 diabetes under drug therapy, HbA_{1c} $> 7\%$, BMI ≥ 40 kg/m², medication influencing vitamin D or glucose metabolism in the last 3 months, regular consumption of supplements containing > 400 IU/day of vitamin D₃ over the last 2 months, renal insufficiency (estimated glomerular filtration rate < 60 mL/min), pregnancy, cirrhosis, intestinal malabsorption (e.g. bypass surgery, celiac disease) or osteoporosis, history of nephrolithiasis, hypercalcaemia (> 2.6 mM), hypercalciuria (> 0.6 fasting urine

Clinical Study	P Lemieux and others	Vitamin D, insulin sensitivity and secretion	181:3	289
----------------	----------------------	--	-------	-----

calcium/creatinine ratio), >5% change in weight in the last 3 months, diseases affecting glucose metabolism (e.g. hyperthyroidism), pacemaker (for bioimpedance only); inability to provide informed consent and complete questionnaires due to physical or mental problems.

Intervention

The treatment group received 5,000 IU of vitamin D₃ once a day (half a tablet of D-tabs 10,000 IU, Riva Laboratory, Québec, Canada), aimed to reach serum 25(OH)D ≥ 75 nmol/L (13), whereas the control group received placebo daily, for 6 months. Placebo pills were produced by the hospital pharmacy using lactose monohydrate powder. To ensure blinding, both the vitamin D and placebo pills were enrobed in capsules of identical shape and colour. The pharmacy department dispensed study pills and asked participants to take the pill with the morning meal. The pharmacist assessed compliance with treatment at 3 and 6 months using pill count. Participants were questioned about adverse effects or change in their health condition during the intervention. All participants were instructed to maintain their usual diet and exercise habits throughout the study. Medication was assessed at baseline and any further change was documented during follow-up. An independent blinded clinician performed safety monitoring of serum calcium at 3 months.

Outcome measures

Hyperinsulinemic-euglycemic clamp

The primary endpoint was change between baseline and 6 months in peripheral insulin sensitivity, measured by M-value, using a 2-h hyperinsulinemic-euglycemic clamp. Participants were instructed to eat the same dinner the night before the clamp at 0 and 6 months and to avoid physical activity (except walking), alcohol and caffeine for 24 h prior to the procedure. After a 12-h overnight fast, an intravenous catheter was placed in an antecubital vein for the administration of regular insulin (Humulin® R) and dextrose 20%. A second catheter was placed in the other arm for blood sampling. Insulin was infused at a fixed rate of 40 mU/m²/min. Dextrose infusion rate was adjusted every 5 min to maintain capillary glucose measurement (using Xceed Pro and Accu-Chek Inform II glucose metres) between 5.0 and 5.5 mM. Serum insulin was measured every 5 min during the last 30 min of the clamp. M-value, an index of stimulated glucose disposal rate, was obtained by dividing exogenous glucose

infusion rate by kilogram of body weight during the last 30 minutes (mg/kg×min). Another index of peripheral insulin sensitivity (M/I), defined as M-value divided by the mean serum insulin concentration during the last 30 min of the clamp, was calculated.

Oral glucose tolerance test

After a 12-h fast, a 75 g 2-h OGTT was performed in the morning at baseline and at 6 months. The OGTT was scheduled in a period of 1 to 7 days prior to or after the hyperinsulinemic-euglycemic clamp. Blood samples were collected through a venous catheter in serum separating tubes (SST) at 0, 15, 30, 60, 90 and 120 min for measurement of insulin, glucose and C-peptide concentrations. Hepatic insulin sensitivity was assessed by HOMA2%S using fasting glucose and insulin concentrations (HOMA2 calculator – www.dtu.ox.ac.uk/homa) and whole-body insulin sensitivity was assessed by Matsuda index (10,000 divided by the squared root of ((fasting glucose×fasting insulin)×(mean glucose^{30–120 min}×mean insulin^{30–120 min})). Matsuda index was added *a posteriori* for comparison with previously published studies. Insulin secretion was assessed by HOMA2-B using fasting C-peptide and glucose (HOMA2 calculator), the insulinogenic index (delta C-peptide^{0–30 min} divided by delta glucose^{0–30 min}) and the area under the curve (AUC) for C-peptide. Disposition index, calculated by multiplying M-value by insulinogenic index, was used to estimate β -cell function.

Anthropometry and blood pressure

Blood pressure and anthropometric measurements including weight, height, waist and hip circumference were assessed at baseline and every 3 months. Participants were weighed without shoes in light clothing on a calibrated scale to the nearest 0.1 kg. Height was measured at the nearest millimetre, using a wall-mounted stadiometer. BMI was calculated in kg/m². Blood pressure was measured three times in a sitting position after a 15-min rest; the mean of the last two measurements was calculated. Waist circumference was measured twice, at the nearest 0.5 cm, using a tape placed on top of the upper iliac crest, at the end of a normal expiration. Hip circumference was measured in duplicate, at the nearest millimetre, using a tape placed at the maximum width of the buttocks. Percent body fat was calculated by bioimpedance, using the InBody520 device.

Serum 25(OH)D, parathyroid hormone and metabolic markers

Serum was stored at -80°C until the end of the study, where samples were analysed in a single batch for all biomarkers. For screening of the participants, serum 25(OH)D was analysed by an electrochemiluminescence immunoassay (Roche, Cobas 6000, CV 10%), which was more readily available. However, for all statistical analyses, serum 25(OH)D was analysed in a single batch at the study end for assessment of baseline, 3-month and 6-month frozen samples by ultra-performance liquid-chromatography tandem mass spectrometry (Waters ACQUITY UPLC System, CV 7.3%). The department of biochemistry measuring serum 25(OH)D met the performance target set by the DEQAS Advisory Panel. PTH was assessed by a chemiluminescence immunoassay (Beckman Coulter, Access 2, CV 4%), while HbA_{1c} was measured by a second-generation colorimetric method (Roche, Integra 800, CV 4%). Plasma glucose was measured enzymatically by the hexokinase method (Roche, Modular E170, CV 1.8%), whereas insulin and C-peptide were measured with an electrochemiluminescence immunoassay (Roche, Modular E170, CV 3 and 2.5%, respectively). Calcium was measured via the o-cresolphthalein complexone method (Roche, Modular E170, CV 3.6%) every 3 months for safety purposes.

Diet, physical activity and sun exposure

Weekly sun exposure was calculated (h/week) via a non-validated questionnaire, based on the time that participants reported spending outdoors between 11:00h and 14:00h over the last 3 months (14), excluding days between November 1st and March 31th, as there is no skin production of vitamin D₃ during that period in Québec City. Participants were also asked about their sunscreen use (sun protection factor, frequency), clothes worn (hat, shirt, pants, long sleeves, etc.) and localisation (shadow, sun) while performing outdoor activities. Physical activity habits during the last week before the baseline and 6-month study visits were estimated using the short International Physical Activity Questionnaire (IPAQ, 2005). The volume of activity per week (walking, moderate and vigorous physical activity) was computed by weighting each type of activity by its energy requirements (METs) and by multiplying by the number of corresponding days per week. Results were summed to yield a score in MET-minutes per week. Finally, a Web-based, food frequency questionnaire, validated in the Québec City

metropolitan area, was administered to evaluate daily energy, vitamin D and calcium intakes both at the beginning and at the end of the study (15).

Statistical methods

As there was no available similar study to estimate the mean M-value of our study population when the study was planned, our sample size was based on the mean M-value (5.49 ± 2.33) of 65 vitamin D-deficient postmenopausal women aged 50–68 years with a waist circumference measurement of $>88\text{cm}$ who had taken part in a cross-sectional study conducted in the Québec City area (unpublished result). We hypothesised that the M-value would remain stable between baseline and 6 months in the control group, while it would increase by 22% in the treatment group. This was based on the study by von Hurst *et al.*, who showed that vitamin D-deficient South Asian women with insulin resistance who reached serum 25(OH)D concentrations $>80\text{nmol/L}$ with vitamin D supplementation experienced an increase in insulin sensitivity (HOMA-IS) by 22% versus placebo at 6 months (16). This difference corresponds to a medium effect with a Cohen's d of 0.52 (17). The trial was designed to recruit a total of 120 participants, providing a statistical power of 80% to detect this clinically significant difference between groups at 6 months with a Student's *t*-test and with an alpha error of 0.05. To account for a dropout rate of 10%, sample size was increased at 130 participants.

Statistical analyses were performed by an independent statistician using SAS version 9.4 (SAS Institute Inc.). Group comparability at baseline was assessed using Student's *t*-test and chi-squared tests for continuous and categorical variables, respectively. Baseline comparison between participants who completed the study and those who dropped out was performed using the same tests. Changes from baseline to endpoint measures of primary and secondary outcomes between and within groups were assessed using repeated-measures two-way analyses of variance (rmANOVA). Treatment and time effects and their interaction on outcome measures were tested. The interaction term (time-by-treatment effect) is the term of interest as it tests whether the temporal change differs between groups. The treatment term tests if a difference exists between groups, for all visits combined, while the time term tests if a difference exists between visits, for all groups. rmANOVA were estimated using a linear mixed model. These models were adjusted for variables that differed at baseline between groups. Squared-root transformation was used when needed to meet model

assumptions, such as normality and homoscedasticity of the residuals. According to the intention-to-treat principle, data were analysed for all randomised participants regardless of compliance to treatment and dropout. Linear mixed model enables to use all the available data, even if no values are present after baseline for an individual; thus, no participant was excluded from the analysis. A sensitivity analysis was also conducted for the primary outcome (M-value by clamp), excluding participants with hypoglycaemia (capillary blood glucose <4.0 mmol/L) at any timepoint during the clamp and/or with a difference of ≥ 0.5 mmol/L between mean capillary blood glucose in the last 30 min of the clamp at 0 and 6 months. A pre-specified subgroup analysis was performed per-protocol, including participants who took $\geq 80\%$ of their study medication. Moreover, *post hoc* exploratory subgroup analyses were done according to baseline glucose tolerance status (normal glucose tolerance versus prediabetes versus diabetes) and baseline 25(OH)D (<50 nmol/L).

Results

Due to difficulties enrolling participants in the trial, recruitment was stopped after randomising the 96th participant. Out of the 796 screened participants, 96 met the inclusion criteria and were randomised, 48 in each group (Fig. 1). Main reasons for exclusion were absence of serum 25(OH)D criterion ($n=185$), declined participation ($n=154$), waist circumference below defined inclusion criteria ($n=104$), consumption of vitamin D supplements ($n=65$) or diabetes medication ($n=42$) and absence of risk factors associated with insulin resistance ($n=38$). One participant in the treatment group withdrew after the first part of the baseline study visit (did not undergo the OGTT nor the clamp) for personal reasons and did not receive the allocated intervention. Four participants in the placebo group withdrew during the trial and did not complete the follow-up visits. Reasons for withdrawal were personal reasons ($n=2$) or experiencing side effects ($n=2$).

Figure 1

Flowchart showing participant enrolment, allocation and analysis.

Moreover, three more participants stopped taking the study supplementation during the trial because of adverse effects (treatment $n=2$, placebo $n=1$) but completed the follow-up visits. Finally, one participant in the placebo group had no data for the primary outcome due to impossibility to insert an intravenous catheter to perform the hyperinsulinemic-euglycemic clamp. Compared to the participants who did not complete the study, those who completed had a worse insulin sensitivity profile, as determined by significantly lower HOMA2%S (40.6 vs 62.4, $P=0.008$) and Matsuda (2.14 vs 3.51, $P=0.015$) indices. There was also a trend for the inclusion of more men (63.7 vs 20.0%, $P=0.071$) with higher fasting glucose (6.0 vs 5.2 mmol/L, $P=0.079$) and insulin (130 vs 81 pmol/L, $P=0.091$) concentrations.

Baseline characteristics of the 96 randomised participants are shown in Table 1. Participants were a majority of men in their 50s with obesity. Over half of the participants had either prediabetes (35.8%) or newly diagnosed type 2 diabetes (20%). Mean baseline serum 25(OH)D concentration by mass spectrometry was 51.1 nmol/L, with 45.8% of the participants having a 25(OH)D concentration ≤ 50 nmol/L (min–max: treatment, 19–81 nmol/L; placebo, 25–97 nmol/L). Baseline characteristics were similar for both groups, except for significantly lower serum triglycerides in the treatment group, a trend for a higher dietary vitamin D intake and for a lower serum 25(OH)D concentration in the treatment group. Difference in 25(OH)D between groups was thought to be potentially clinically significant and therefore, results were adjusted for baseline 25(OH)D. As further adjustment for serum triglycerides did not change results, these data are not presented.

Change in serum 25(OH)D concentrations and compliance with study medication

Serum 25(OH)D was significantly higher in the treatment group at 3 months (122.9 vs 52.2 nmol/L, $P<0.001$) and 6 months (127.6 vs 51.8 nmol/L, $P<0.001$) compared with the control group. After 6 months, serum 25(OH)D increased by a mean of 79.1 nmol/L (95% CI 73.1, 85.2) in the treatment group and did not change in the placebo group (1.87 nmol/L (95% CI –4.33, 8.07)) (Fig. 2). Moreover, 95.7% of the participants in the treatment group reached a serum 25(OH)D value of >75 nmol/L at 6 months. Compliance was similar in both groups, with 93.2 and 88.6% of the participants in the treatment and control groups who took $\geq 80\%$ of their study medication,

respectively ($P=0.713$). Compliance data were missing for eight participants.

Change in primary outcome (M-value)

At baseline, 80.9% ($n=38/47$) of participants in both groups reached the target capillary blood glucose during the last 30 min of the clamp while at 6 months, 85.1% ($n=40/47$) and 83.7% ($n=36/43$) of participants in treatment and placebo groups, respectively met this criterion. There was a significant time-by-treatment interaction for insulin sensitivity, as assessed by M-value and M/I ratio, in favour of the vitamin D group (Table 2). Indeed, M-value increased by a mean of 22.9% in the treatment group, while it remained stable in the placebo group. Results were similar in the sensitivity analysis including participants who had no hypoglycaemia and/or a difference <0.5 mmol/L between mean capillary blood glucose in the last 30 min of the clamp at 0 and 6 months ($n=72$). A *post hoc* exploratory subgroup analysis revealed that between-group differences in M-value appeared to be more important in participants with diabetes or prediabetes at baseline (Table 3). A second *post hoc* subgroup analysis restricted to participants with baseline serum 25(OH)D <50 nmol/L ($n=44$) no longer demonstrated a significant between-group difference for change in M-value.

Changes in secondary outcomes (insulin sensitivity, insulin secretion, β -cell function, metabolic markers, blood pressure and anthropometric measures)

No between-group differences were observed for changes in insulin sensitivity and insulin secretion indices derived from fasting values or the OGTT (Table 4). However, there was a significant time-by-treatment interaction ($P=0.039$) for disposition index, calculated as M-value $\times$ insulinogenic index, with stability in the control group and improvement in the treatment group at 6 months. Changes in fasting glucose, 2-h glucose post OGTT and HbA_{1c} were similar between groups. Moreover, changes in weight, BMI, waist and hip circumference, body fat mass and blood pressure did not differ between the intervention and placebo groups (Table 5). Subgroup analyses restricted to participants who took $\geq 80\%$ of their study medication or to those with abnormal baseline glucose tolerance status (prediabetes or diabetes) gave similar results.

Clinical Study	P Lemieux and others	Vitamin D, insulin sensitivity and secretion	181:3	293
----------------	----------------------	--	-------	-----

Table 1 Baseline characteristics of the participants by treatment group^a.

	Treatment, <i>n</i> = 48	Placebo, <i>n</i> = 48	<i>P</i> value ^b
Age, years	58.3 (8.8)	59.1 (9.5)	0.674
Women	41.7	35.4	0.675
Menopausal women	85.0	82.4	1.000
Season of recruitment			0.782
Winter	29.2	29.2	
Spring	35.4	29.2	
Summer	16.7	25.0	
Fall	18.8	16.7	
Education			0.127
Primary school	0.0	4.2	
High school	16.7	22.9	
College	14.6	25.0	
University	68.8	47.9	
Family income			0.489
0–19,000\$	10.4	2.1	
20,000–39,000\$	10.4	14.6	
40,000–59,000\$	16.7	22.9	
60,000–79,000\$	14.6	16.7	
80,000–99,000\$	14.6	16.7	
>100,000\$	29.2	27.1	
Declined to answer	4.2	0.0	
Current smoking	8.3	4.2	0.677
First-degree relative with diabetes	59.6	47.9	0.306
BMI, kg/m ²	32.2 (4.3)	32.1 (3.9)	0.889
Waist circumference, cm	107.9 (9.0)	108.5 (9.5)	0.760
Systolic blood pressure, mmHg	129.2 (13.6)	131.2 (17.3)	0.532
Diastolic blood pressure, mmHg	76.2 (9.3)	76.5 (9.7)	0.862
HbA _{1c} , %	5.7 (0.3)	5.8 (0.3)	0.678
Fasting glucose, mmol/L	5.9 (0.8)	6.1 (1.1)	0.588
Fasting insulin, pmol/L	130.7 (60.0)	125.4 (54.0)	0.649
2-h glucose post OGTT, mmol/L	8.2 (3.4)	8.4 (3.1)	0.745
Glucose status			0.895
Prediabetes ^c	34.0	37.5	
Diabetes ^d	19.1	20.8	
25-Hydroxyvitamin D ^e , nmol/L	48.5 (13.0)	53.7 (15.0)	0.073
Parathyroid hormone, ng/L	57.3 (21.2)	57.3 (20.3)	0.992
Corrected calcium, mmol/L	2.25 (0.08)	2.27 (0.07)	0.251
Serum creatinine, µmol/L	74.6 (14.1)	73.9 (13.1)	0.800
Triglycerides, mmol/L	1.66 (0.65)	2.33 (1.90)	0.023
ALT, U/L	26.5 (14.0)	29.6 (14.5)	0.302
Urinary calcium/creatinine ratio, mmol/mmolcr	0.19 (0.15)	0.17 (0.11)	0.623
Time spent outdoor between 11 h and 14 h, h/week	4.0 (4.9)	3.6 (4.8)	0.738
Physical activity, MET, min/week	1709 (1928)	1371 (1798)	0.377
Percent body fat ^f , %	38.5 (8.3)	37.4 (7.8)	0.581
Daily energy intake, kJ	11740 (3879)	11033 (3075)	0.328
Dietary vitamin D intake, IU/day	380 (228)	312 (168)	0.086
Dietary calcium intake, mg/day	1528 (678)	1352 (512)	0.153

^aData are presented as mean (s.d.) or %. ^bStudent's *t*-test or exact chi-squared test, as appropriate. ^cFasting glucose 6.1–6.9 mmol/L or 2-h glucose post OGTT 7.8–11.0 mmol/L. ^dFasting glucose ≥7.0 mmol/L or 2-h glucose post OGTT ≥11.1 mmol/L. ^eBy liquid-chromatography tandem mass spectrometry.

^fBy bioimpedance.

MET, metabolic equivalent of task.

Safety and adverse events

Five participants discontinued the study because of side effects, two in the treatment group (gastrointestinal complaints and dizziness) and three in the placebo group

(gastrointestinal complaints, anxiety and hypoglycaemia). Moreover, participants who completed the study reported gastrointestinal issues (treatment, *n*=6, placebo, *n*=5), dizziness (placebo, *n*=1), polyuria (treatment, *n*=1) and musculoskeletal symptoms (placebo, *n*=1). None of the

Figure 2

Mean (SD) serum 25-hydroxyvitamin D concentrations at baseline and after 3 and 6 months in the vitamin D and placebo groups. * $P < 0.001$ for the difference between groups.

participants developed hypercalcaemia, hypercalciuria or nephrolithiasis during the trial.

Discussion

In this randomised, placebo-controlled trial conducted in individuals at high risk of type 2 diabetes or with newly diagnosed type 2 diabetes, vitamin D₃ supplementation at a dosage of 5000 IU once daily for 6 months significantly increased peripheral insulin sensitivity, as measured by the gold-standard hyperinsulinemic-euglycemic clamp. A subgroup analysis revealed that this effect appeared to be more important in subjects with new diabetes or newly diagnosed diabetes at baseline. Moreover, although there were no between-group differences in indices of

insulin secretion, there was a significant beneficial effect of vitamin D on the disposition index, suggesting that vitamin D may improve β -cell function. However, there was no effect of vitamin D supplementation on any measures of insulin sensitivity derived from fasting values and from the OGTT, anthropometric measures, blood pressure, fasting and 2-h glucose and HbA_{1c}.

All five RCTs that used the hyperinsulinemic-euglycemic clamp failed to demonstrate a beneficial effect of vitamin D on insulin sensitivity. However, three of them were limited by small sample size and short duration of treatment. Indeed, one study treated 18 healthy subjects with 1.5 μ g of calcitriol daily for 7 days, while in another study, 12 healthy subjects received 50,000 IU of vitamin D₂ weekly for 8 weeks (6, 7). The third trial included 16 patients with type 2 diabetes who were supplemented with vitamin D₃ 11,200 IU daily for 2 weeks and then 5600 IU daily for 10 weeks (8). While insulin sensitivity did not differ between groups, borderline improvements in insulin secretion were noted in the treatment group. In the fourth trial, the effect of a bolus dose of 100,000 IU of vitamin D₃ followed by 4,000 IU daily for 16 weeks in 65 overweight and vitamin D-deficient participants was evaluated (10). Compared with our trial, participants in this study were healthier and had a better insulin sensitivity profile, with a higher baseline M-value (6.7 ± 2.9 vs 4.1 ± 2.3 mg/kg $\times$ min). Results of this study are concordant with our subgroup analysis showing that vitamin D supplementation did not change M-value in participants with normal glucose tolerance. Finally, in a 6-month trial, 62 participants of Nordic or South Asian ethnicity with type 2 diabetes and vitamin D deficiency were treated with a single high dose of vitamin D₃ 400,000 IU, with an additional dose of 200,000 IU at 4 weeks if serum 25(OH)D was below 100 nmol/L (9). Divergence in results with our study may

Table 2 Mean baseline and 6-month values and the mean absolute changes in M-value and M/I ratio in the treatment and placebo groups.

	Mean (S.E.M.)		Mean change (95% CI)	P value ^a		
	Baseline ^b	6 months		Time	Treatment	Interaction
M-value ^c , mg/kg $\times$ min						
Treatment (n = 47)	3.97 (0.41)	4.88 (0.41)	0.92 (0.24, 1.59)	0.234	0.628	0.009
Placebo (n = 47/43 ^d)	4.15 (0.41)	4.12 (0.42)	-0.03 (-0.73, 0.67)			
M/I ratio ^c , mg/kg $\times$ min/pmol/L						
Treatment (n = 47)	0.0062 (0.0007)	0.0077 (0.0007)	0.0015 (0.0003, 0.0027)	0.050	0.705	0.031
Placebo (n = 45/43)	0.0064 (0.0007)	0.0069 (0.0007)	0.0005 (-0.0007, 0.0017)			

^aTwo-way ANOVA with repeated measures was used to assess time and treatment effects as well as their interactions, after adjustment for baseline serum 25-hydroxyvitamin D. The P value for the interaction (time-by-treatment effect) is of interest, as it tests whether the temporal change differs between groups. ^bBaseline and 6-month data are presented as mean (S.E.M.). ^cVariables were squared-root-transformed for analysis but original values are presented. ^dFor each measure, if the number of participants with available data was different between baseline and 6 months, n is presented as baseline/6 months.

S.E.M., standard error of the mean.

Clinical Study	P Lemieux and others	Vitamin D, insulin sensitivity and secretion	181:3	295
----------------	----------------------	--	-------	-----

Table 3 Mean baseline and 6-month values and the mean absolute changes in M-value in the subgroups of participants with normal glucose tolerance, prediabetes or diabetes at baseline.

	Mean (S.E.M.)		Mean change (95% CI) Δ 6 months	P value ^a		
	Baseline ^c	6 months		Time	Treatment	Interaction
M-value ^b , mg/kg × min						
Normal glucose tolerance						
Treatment (n = 22)	5.49 (0.67)	5.94 (0.67)	0.45 (−0.74, 1.64)	0.641	0.788	0.677
Placebo (n = 19/17 ^d)	5.31 (0.72)	5.75 (0.75)	0.43 (−0.91, 1.77)			
Prediabetes						
Treatment (n = 16)	3.13 (0.56)	4.38 (0.56)	1.25 (0.12, 2.38)	0.591	0.952	0.015
Placebo (n = 18/16)	3.88 (0.52)	3.32 (0.55)	−0.56 (−1.68, 0.56)			
Diabetes						
Treatment (n = 9)	1.50 (0.49)	2.96 (0.49)	1.46 (0.58, 2.34)	0.080	0.562	0.030
Placebo (n = 10)	2.62 (0.46)	2.70 (0.46)	0.08 (−0.76, 0.91)			

^aTwo-way ANOVA with repeated measures was used to assess time and treatment effects as well as their interactions, after adjustment for baseline serum 25-hydroxyvitamin D. The P value for the interaction (time-by-treatment effect) is of interest, as it tests whether the temporal change differs between groups.

^bVariables were squared-root-transformed for analysis but original values are presented. ^cBaseline and 6-month data are presented as mean (S.E.M.). ^dFor each measure, if the number of participants with available data was different between baseline and 6 months, n is presented as baseline/6 months.

S.E.M., standard error of the mean.

be explained by the lack of sustained concentrations of serum 25(OH)D in the treatment group, with a mean of 53.7 ± 9.2 nmol/L at 6 months and by the inclusion of participants with long standing type 2 diabetes (mean diabetes duration of 11.0 ± 6.6 years and 7.9 ± 5.7 years in the vitamin D and placebo group, respectively), compared to our trial, which included participants at high risk for diabetes or with newly diagnosed type 2 diabetes. However, it remains unclear how the diabetes duration could have affected the results. Furthermore, no effect of vitamin D was found in two RCTs that used the hyperglycemic clamp to evaluate insulin sensitivity. One was performed in 104 healthy subjects who received 20,000 IU of vitamin D₃ twice weekly for 6 months and the other one in 44 participants with prediabetes or type 2 diabetes who were treated with 30,000 IU of vitamin D₃ once weekly for 8 weeks (18, 19). Nevertheless, the hyperglycemic clamp is less accurate than the gold-standard hyperinsulinemic-euglycemic clamp to estimate insulin sensitivity (20).

In concordance with three recent meta-analyses, one including 23 RCTs in patients with type 2 diabetes, another including 35 RCTs performed in various populations and the third one including 10 RCTs in prediabetes patients (4, 5, 21), we did not find any effect of vitamin D supplementation on measures of insulin sensitivity and insulin secretion based on fasting indices and the OGTT. The vast majority of RCTs, and therefore those included in the meta-analyses, used indirect insulin sensitivity markers such as HOMA and Matsuda instead of the hyperinsulinemic-euglycemic clamp. Just like the population included in our study, many previous trials were performed in insulin-resistant participants, and most of them have shown negative results with the use of surrogate markers of insulin sensitivity

(13, 22, 23, 24, 25, 26, 27). One trial demonstrated a significant improvement in insulin sensitivity assessed by HOMA2%S in 81 vitamin D-deficient South Asian women with insulin resistance treated with vitamin D₃ 4,000 IU daily for 6 months, and a second one showed a significant decrease in HOMA-IR in Iranian women with polycystic ovary syndrome (16, 28). However, these results may not be generalisable to Caucasian populations.

The discordant effects of vitamin D supplementation that we observed on insulin sensitivity measures, with a significant beneficial effect on M-value and no difference on HOMA2%S and Matsuda indices, suggests that vitamin D acts mainly on peripheral insulin sensitivity. Indeed, HOMA2%S primarily reflects hepatic insulin sensitivity, and Matsuda whole-body insulin sensitivity with a major contribution of hepatic insulin sensitivity (29), whereas the hyperinsulinemic-euglycemic clamp (M-value), at the insulin dose that we used, evaluates predominantly muscle insulin sensitivity (30, 31). Although the exact mechanisms by which vitamin D influences muscle insulin sensitivity in humans remain to be elucidated, preclinical evidence suggests that vitamin D may improve diabetes-induced muscle dysfunction and atrophy (32) as well as muscle fat infiltration (33). Also, vitamin D could increase glucose uptake in adipose tissue and muscle by stimulating GLUT-4 expression (34, 35) and reduce low-grade chronic systemic inflammation (36, 37).

Another notable finding of our study is the significant beneficial effect of vitamin D supplementation on disposition index, a measure of β -cell compensatory capacity, with stability in the control group and improvement in the treatment group. Preservation of β -cell function is important, as it was shown to predict reduced

Clinical Study	P Lemieux and others	Vitamin D, insulin sensitivity and secretion	181:3	296
----------------	----------------------	--	-------	-----

Table 4 Mean baseline and 6-month values and the mean absolute changes in insulin sensitivity, insulin secretion, β -cell function and metabolic markers in the treatment and placebo groups.

	Mean (S.E.M.)		Mean change (95% CI)	P value ^a		
	Baseline ^b	6 months	Δ 6 months	Time	Treatment	Interaction
Insulin sensitivity						
HOMA2%S						
Treatment (<i>n</i> = 47)	40.6 (2.4)	41.9 (2.4)	1.3 (−2.6, 5.2)	0.723	0.979	0.201
Placebo (<i>n</i> = 48/44 ^c)	42.4 (2.4)	40.1 (2.4)	−2.3 (−6.3, 1.7)			
Matsuda index						
Treatment (<i>n</i> = 47)	2.09 (0.17)	2.19 (0.17)	0.10 (−0.13, 0.33)	0.640	0.496	0.481
Placebo (<i>n</i> = 46/44)	2.30 (0.17)	2.28 (0.17)	−0.02 (−0.26, 0.22)			
Insulin secretion						
HOMA2-B						
Treatment (<i>n</i> = 47)	132.7 (5.1)	127.5 (5.1)	−5.2 (−13.7, 3.2)	0.044	0.800	0.751
Placebo (<i>n</i> = 48/44)	135.4 (5.1)	128.2 (5.2)	−7.2 (−15.8, 1.5)			
Insulinogenic index, pmol/L/mmol/L						
Treatment (<i>n</i> = 47)	487.5 (45.3)	496.4 (45.3)	8.8 (−44.6, 62.2)	0.855	0.293	0.525
Placebo (<i>n</i> = 47/44)	434.9 (44.9)	419.0 (45.4)	−15.9 (−71.5, 39.6)			
AUC for C-peptide, pmol/L × 120 min						
Treatment (<i>n</i> = 47)	410.2 (16.4)	403.3 (16.4)	−6.9 (−25.6, 11.8)	0.968	0.256	0.296
Placebo (<i>n</i> = 47/43)	377.4 (16.3)	384.9 (16.5)	7.5 (−12.2, 27.1)			
β -cell function						
Disposition index ^d						
Treatment (<i>n</i> = 47)	2425.5 (421.9)	2692.5 (421.9)	267.0 (−343.4, 877.4)	0.497	0.237	0.039
Placebo (<i>n</i> = 47/43)	1768.1 (423.6)	1712.6 (430.5)	−55.5 (−696.3, 585.3)			
Metabolic markers						
HbA _{1c} , %						
Treatment (<i>n</i> = 48/47)	5.7 (0.1)	5.6 (0.1)	−0.10 (−0.19, −0.01)	0.017	0.604	0.694
Placebo (<i>n</i> = 48/43)	5.8 (0.1)	5.7 (0.1)	−0.07 (−0.17, 0.03)			
Fasting glucose, mmol/L						
Treatment (<i>n</i> = 47)	5.9 (0.1)	6.1 (0.1)	0.14 (−0.07, 0.35)	0.520	0.817	0.242
Placebo (<i>n</i> = 48/44)	6.1 (0.1)	6.0 (0.2)	−0.04 (−0.26, 0.18)			
2-h glucose post OGTT, mmol/L						
Treatment (<i>n</i> = 47)	8.1 (0.5)	8.1 (0.5)	−0.00 (−0.53, 0.53)	0.594	0.552	0.586
Placebo (<i>n</i> = 48/44)	8.4 (0.5)	8.6 (0.5)	0.21 (−0.34, 0.75)			

^aTwo-way ANOVA with repeated measures was used to assess time and treatment effects as well as their interactions, after adjustment for baseline serum 25-hydroxyvitamin D. The *P* value for the interaction (time-by-treatment effect) is the one of interest as it tests whether the temporal change differs between groups. ^bBaseline and 6-month data are presented as mean (S.E.M.). ^cFor each measure, if the number of participants with available data was different between baseline and 6 months, *n* is presented as baseline/6 months. ^dCalculated by multiplying M-value by insulinogenic index. Variable was squared-root-transformed for analysis but original values are presented. Mean change for the squared-root values are 4.03 (0.05, 8.01) in the treatment group and −2.05 (−6.23, 2.14) in the control group.

AUC, area under the curve; HOMA2%S, Homeostasis Model Assessment 2 index of insulin sensitivity; HOMA2-B, Homeostasis Model Assessment 2 index of β -cell function; OGTT, oral glucose tolerance test; S.E.M., standard error of the mean.

diabetes risk (38). Indeed, DeFronzo *et al.* demonstrated that deterioration of β -cell function, assessed by the disposition index, is the strongest predictor of conversion to diabetes in subjects with impaired glucose tolerance (39). Even though we observed no differences in indices of insulin secretion following OGTT between groups, a beneficial effect of vitamin D on insulin secretion relative to insulin sensitivity was seen, reflected by the disposition index. Moreover, we could have missed positive effects of vitamin D on insulin secretion *per se*, as we did not use the hyperglycemic clamp, the gold-standard method to assess insulin secretion (20). Mechanisms by which vitamin D plays a role on preservation of β -cell function remain largely unknown. However, previous studies have shown

that pancreatic β -cells contain high levels of vitamin D receptor and that vitamin D enhances β -cell intracellular calcium concentration and influx (40, 41).

Our study has notable strengths including the randomised, double-blind design and the use of the gold-standard hyperinsulinemic-euglycemic clamp to evaluate insulin sensitivity. Furthermore, we selected participants at high risk for type 2 diabetes or with newly diagnosed type 2 diabetes, a group that had not been specifically studied using the clamp. Another strength is the use of high-dose vitamin D supplementation for 6 months and the high rate of compliance, with a large difference in serum 25(OH)D during the study between the treatment and control groups. However, limitations include

Clinical Study	P Lemieux and others	Vitamin D, insulin sensitivity and secretion	181:3	297
----------------	----------------------	--	-------	-----

Table 5 Mean baseline and 6-month values and the mean absolute changes in anthropometry and blood pressure in the treatment and placebo groups.

	Mean (S.E.M.)	Mean change from baseline (95% CI)		P value ^a		
	Baseline ^b	Δ 3 months	Δ 6 months	Time	Treatment	Interaction
Weight, kg						
Treatment (n = 48/47 ^c)	91.0 (2.1)	0.01 (−0.63, 0.65)	−0.59 (−1.49, 0.31)	0.004	0.911	0.696
Placebo (n = 48/44)	90.7 (2.1)	0.14 (−0.51, 0.80)	−0.84 (−1.76, 0.09)			
BMI, kg/m ²						
Treatment (n = 48/47)	32.2 (0.6)	0.02 (−0.20, 0.24)	−0.18 (−0.49, 0.13)	0.004	0.868	0.674
Placebo (n = 48/44)	32.1 (0.6)	0.06 (−0.17, 0.28)	−0.28 (−0.60, 0.03)			
Fat mass by bioimpedance, %						
Treatment (n = 36)	38.4 (1.3)	ND	−0.32 (−1.00, 0.37)	0.026	0.303	0.305
Placebo (n = 30)	36.8 (1.3)	ND	−0.84 (−1.59, −0.09)			
Waist circumference, cm						
Treatment (n = 48/47)	107.9 (1.4)	0.75 (−0.02, 1.53)	0.18 (−0.90, 1.26)	0.020	0.826	0.814
Placebo (n = 48/44)	108.6 (1.4)	0.39 (−0.40, 1.19)	−0.19 (−1.30, 0.92)			
Hip circumference, cm						
Treatment (n = 46)	112.5 (1.2)	ND	0.41 (−0.52, 1.35)	0.912	0.205	0.179
Placebo (n = 47/44)	112.9 (1.2)	ND	−0.49 (−1.43, 0.45)			
SBP, mmHg						
Treatment (n = 48/47)	129.4 (2.2)	1.52 (−1.83, 4.88)	−1.61 (−5.94, 2.72)	0.001	0.756	0.526
Placebo (n = 47/44)	131.0 (2.2)	1.85 (−1.57, 5.27)	−4.06 (−8.49, 0.37)			
DBP, mmHg						
Treatment (n = 48/47)	76.5 (1.4)	0.66 (−1.33, 2.65)	−2.45 (−5.04, 0.15)	<0.001	0.576	0.487
Placebo (n = 48/44)	76.3 (1.4)	2.22 (0.20, 4.25)	−0.37 (−3.03, 2.28)			

^aTwo-way ANOVA with repeated measures was used to assess time and treatment effects as well as their interaction, after adjustment for baseline serum 25-hydroxyvitamin D. The P value for the interaction (time-by-treatment effect) is the one of interest as it tests whether the temporal change differs between groups. ^bBaseline data are presented as mean (S.E.M.). ^cFor each measure, if the number of participants with available data was different between baseline and 6 months, n is presented as baseline/6 months.

DBP, diastolic blood pressure; ND, not documented; SBP, systolic blood pressure; S.E.M., standard error of the mean.

the selection of a Caucasian population, restricting generalizability to other ethnic groups. Moreover, we did not reach our target sample size. However, our sample of 96 participants still had the power to detect a medium to large effect. Indeed, using the same criteria as mentioned in the methods section, it was still possible to detect an effect size of $d=0.58$ with a sample size of 96 participants (where d of 0.5 is considered a medium effect and 0.80 a large effect), compared to our planned sample of 120 participants that could detect a medium effect ($d=0.52$) (17). Since effect sizes in the present study were mostly small, it is probable that reaching the target sample size would not have changed the conclusions. Furthermore, the sample size calculation was based on postmenopausal women, even though the present study included men and women, due to lack of available data at the time of the study. Another limitation is that the mean baseline serum 25(OH)D was higher than expected and only about half of the participants had vitamin D deficiency at study entry due to different methods for serum 25(OH)D measurement and to time difference of up to 3 months between screening and baseline visits. Although subgroup analysis restricted to participants with baseline 25(OH)D <50 nmol/L did not show any significant differences between groups, these

results should be interpreted with caution due to reduced study power and the *post hoc* nature of the analysis. Additionally, although both groups were comparable in terms of confounding factors such as sun exposure, dietary intake of calcium and vitamin D and physical activity, the assessment methods employed, such as the use of a non-validated questionnaire for sun exposure, may have failed to capture differences. Furthermore, although a minority of patients dropped out of the study, excluded participants had a better insulin sensitivity profile. These selection and attrition bias could have overestimated the effect in the mixed models. Finally, as we did not use tracer infusion to estimate glucose rate of appearance and disappearance and as we used an insulin dose that may not have suppressed completely endogenous glucose production in the most highly insulin-resistant participants, we may have underestimated peripheral insulin sensitivity as assessed by M-value or M/I.

In conclusion, this study showed that high-dose vitamin D supplementation for 6 months significantly improved peripheral insulin sensitivity, as assessed by the hyperinsulinemic-euglycemic clamp, and β -cell function in individuals at high risk of diabetes or with newly diagnosed type 2 diabetes. Of interest, two large

RCTs evaluating the effect of high-dose vitamin D₃ (20 000 IU/week for 5 years and 4000 IU/day for a median of 2.5 years) in patients with prediabetes who were mostly vitamin D sufficient at baseline did not show any effect on progression to type 2 diabetes (42, 43). Larger and longer-term RCTs are required to evaluate whether subgroups of patients including those with low vitamin D status may benefit from vitamin D supplementation.

Declaration of interest

A T receives research funding from Johnson & Johnson Medical Companies, Pfizer and Medtronic for studies unrelated to this project. C G, S J W, M C V and A M received research funding from the Food & Health Programmatic grant program from the Canadian Institutes of Health Research (CIHR). A M holds a Canadian Institutes of Health Research (CIHR)/Pfizer research Chair in the pathogenesis of insulin resistance and cardiovascular diseases. M C V is tier 1 Canada Research Chair in Genomics Applied to Nutrition and Metabolic Health. C G received research funding from Technologies Khlôros. She is also a Scholar of the Fonds de Recherche du Québec-Santé and a recipient of a New Investigator award from Diabetes Canada. The other authors have nothing to disclose.

Funding

This work was supported by Diabetes Canada (Grant # OG-3-12-3681) and by the CHU de Québec-Université Laval Research Center.

Author contribution statement

C G formulated the research question, C G and S J W designed research; C G conducted research; A S J analysed data; P L wrote the first draft of the paper; C G had primary responsibility for final content; and all authors critically revised the protocol, read and approved the final version of the manuscript prior to submission.

Acknowledgements

The authors would like to thank all the participants for their commitment as well as the research assistant Sarah Chouinard-Castonguay, all the nurses and research trainees (Claudia Savard, Antoine Leduc, Mehdi Bihya, Véronique Thibeault) for their support in running the trial. The authors also express gratitude to Dr Caroline Albert for vitamin D analysis.

References

- Kayanilyl S, Vieth R, Retnakaran R, Knight JA, Qi Y, Gerstein HC, Perkins BA, Harris SB, Zinman B & Hanley AJ. Association of vitamin D with insulin resistance and beta-cell dysfunction in subjects at risk for type 2 diabetes. *Diabetes Care* 2010 **33** 1379–1381. (<https://doi.org/10.2337/dc09-2321>)
- Gagnon C, Lu ZX, Magliano DJ, Dunstan DW, Shaw JE, Zimmet PZ, Sikaris K, Grantham N, Ebeling PR & Daly RM. Serum 25-hydroxyvitamin D, calcium intake, and risk of type 2 diabetes after 5 years: results from a national, population-based prospective study (the Australian Diabetes, Obesity and Lifestyle study). *Diabetes Care* 2011 **34** 1133–1138. (<https://doi.org/10.2337/dc10-2167>)
- Holick MF. Vitamin D deficiency. *New England Journal of Medicine* 2007 **357** 266–281. (<https://doi.org/10.1056/NEJMr070553>)
- Krul-Poel YH, Ter Wee MM, Lips P & Simsek S. MANAGEMENT of ENDOCRINE DISEASE: The effect of vitamin D supplementation on glycaemic control in patients with type 2 diabetes mellitus: a systematic review and meta-analysis. *European Journal of Endocrinology* 2017 **176** R1–R14. (<https://doi.org/10.1530/EJE-16-0391>)
- Seida JC, Mitri J, Colmers IN, Majumdar SR, Davidson MB, Edwards AL, Hanley DA, Pittas AG, Tjosvold L & Johnson JA. Clinical review: effect of vitamin D3 supplementation on improving glucose homeostasis and preventing diabetes: a systematic review and meta-analysis. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** 3551–3560. (<https://doi.org/10.1210/jc.2014-2136>)
- Fliser D, Stefanski A, Franek E, Fode P, Gudarzi A & Ritz E. No effect of calcitriol on insulin-mediated glucose uptake in healthy subjects. *European Journal of Clinical Investigation* 1997 **27** 629–633. (<https://doi.org/10.1046/j.1365-2362.1997.1520699.x>)
- Simha V, Mahmood M, Ansari M, Spellman CW & Shah P. Effect of vitamin D replacement on insulin sensitivity in subjects with vitamin D deficiency. *Journal of Investigative Medicine* 2012 **60** 1214–1218. (<https://doi.org/10.2310/JIM.0b013e3182747c06>)
- Kampmann U, Mosekilde L, Juhl C, Møller N, Christensen B, Rejnmark L, Wamberg L & Orskov L. Effects of 12 weeks high dose vitamin D3 treatment on insulin sensitivity, beta cell function, and metabolic markers in patients with type 2 diabetes and vitamin D insufficiency – a double-blind, randomized, placebo-controlled trial. *Metabolism: Clinical and Experimental* 2014 **63** 1115–1124. (<https://doi.org/10.1016/j.metabol.2014.06.008>)
- Gulseth HL, Wium C, Angel K, Eriksen EF & Birkeland KI. Effects of vitamin D supplementation on insulin sensitivity and insulin secretion in subjects with type 2 diabetes and vitamin D deficiency: a randomized controlled trial. *Diabetes Care* 2017 **40** 872–878. (<https://doi.org/10.2337/dc16-2302>)
- Mousa A, Naderpoor N, de Courten MP, Teede H, Kellow N, Walker K, Scragg R & de Courten B. Vitamin D supplementation has no effect on insulin sensitivity or secretion in vitamin D-deficient, overweight or obese adults: a randomized placebo-controlled trial. *American Journal of Clinical Nutrition* 2017 **105** 1372–1381. (<https://doi.org/10.3945/ajcn.117.152736>)
- Pocock SJ & Simon R. Sequential treatment assignment with balancing for prognostic factors in the controlled clinical trial. *Biometrics* 1975 **31** 103–115. (<https://doi.org/10.2307/2529712>)
- Moher D, Hopewell S, Schulz KF, Montori V, Gotzsche PC, Devereaux PJ, Elbourne D, Egger M & Altman DG. CONSORT 2010 explanation and elaboration: updated guidelines for reporting parallel group randomised trials. *BMJ* 2010 **340** c869. (<https://doi.org/10.1136/bmj.c869>)
- Gagnon C, Daly RM, Carpentier A, Lu ZX, Shore-Lorenti C, Sikaris K, Jean S & Ebeling PR. Effects of combined calcium and vitamin D supplementation on insulin secretion, insulin sensitivity and beta-cell function in multi-ethnic vitamin D-deficient adults at risk for type 2 diabetes: a pilot randomized, placebo-controlled trial. *PLoS ONE* 2014 **9** e109607. (<https://doi.org/10.1371/journal.pone.0109607>)
- Wacker M & Holick MF. Sunlight and vitamin D: a global perspective for health. *Dermato-Endocrinology* 2013 **5** 51–108. (<https://doi.org/10.4161/derm.24494>)
- Labonte MÈ, Cyr A, Baril-Gravel L, Royer MM & Lamarche B. Validity and reproducibility of a web-based, self-administered food frequency questionnaire. *European Journal of Clinical Nutrition* 2012 **66** 166–173. (<https://doi.org/10.1038/ejcn.2011.163>)
- von Hurst PR, Stonehouse W & Coad J. Vitamin D supplementation reduces insulin resistance in South Asian women living in New Zealand who are insulin resistant and vitamin D deficient – a randomised, placebo-controlled trial. *British Journal of Nutrition* 2010 **103** 549–555. (<https://doi.org/10.1017/S0007114509992017>)
- Sawilowsky SS. New effect size rules of thumb. *Journal of Modern Applied Statistical Methods* 2009 **8** 597–599. (<https://doi.org/10.22237/jmasm/1257035100>)
- Grimmes G, Figenschau Y, Almas B & Jorde R. Vitamin D, insulin secretion, sensitivity, and lipids: results from a case-control study and

- a randomized controlled trial using hyperglycemic clamp technique. *Diabetes* 2011 **60** 2748–2757. (<https://doi.org/10.2337/db11-0650>)
- 19 Wagner H, Alvarsson M, Mannheimer B, Degerblad M & Ostenson CG. No effect of high-dose vitamin D treatment on beta-cell function, insulin sensitivity, or glucose homeostasis in subjects with abnormal glucose tolerance: a randomized clinical trial. *Diabetes Care* 2016 **39** 345–352. (<https://doi.org/10.2337/dc15-1057>)
 - 20 DeFronzo RA, Tobin JD & Andres R. Glucose clamp technique: a method for quantifying insulin secretion and resistance. *American Journal of Physiology* 1979 **237** E214–E223. (<https://doi.org/10.1152/ajpendo.1979.237.3.E214>)
 - 21 Poolsup N, Suksomboon N & Plordplong N. Effect of vitamin D supplementation on insulin resistance and glycaemic control in prediabetes: a systematic review and meta-analysis. *Diabetic Medicine* 2016 **33** 290–299. (<https://doi.org/10.1111/dme.12893>)
 - 22 Davidson MB, Duran P, Lee ML & Friedman TC. High-dose vitamin D supplementation in people with prediabetes and hypovitaminosis D. *Diabetes Care* 2013 **36** 260–266. (<https://doi.org/10.2337/dc12-1204>)
 - 23 Moreira-Lucas TS, Duncan AM, Rabasa-Lhoret R, Vieth R, Gibbs AL, Badawi A & Wolever TM. Effect of vitamin D supplementation on oral glucose tolerance in individuals with low vitamin D status and increased risk for developing type 2 diabetes (EVIDENCE): a double-blind, randomized, placebo-controlled clinical trial. *Diabetes, Obesity and Metabolism* 2017 **19** 133–141. (<https://doi.org/10.1111/dom.12794>)
 - 24 Oosterwerff MM, Eekhoff EM, Van Schoor NM, Boeke AJ, Nanayakkara P, Meijnen R, Knol DL, Kramer MH & Lips P. Effect of moderate-dose vitamin D supplementation on insulin sensitivity in vitamin D-deficient non-Western immigrants in the Netherlands: a randomized placebo-controlled trial. *American Journal of Clinical Nutrition* 2014 **100** 152–160. (<https://doi.org/10.3945/ajcn.113.069260>)
 - 25 Sollid ST, Hutchinson MYS, Fuskevag OM, Figenschau Y, Joakimsen RM, Schirmer H, Njolstad I, Svartberg J, Kamycheva E & Jorde R. No effect of high-dose vitamin D supplementation on glycemic status or cardiovascular risk factors in subjects with prediabetes. *Diabetes Care* 2014 **37** 2123–2131. (<https://doi.org/10.2337/dc14-0218>)
 - 26 Hoseini SA, Aminorroaya A, Iraj B & Amini M. The effects of oral vitamin D on insulin resistance in pre-diabetic patients. *Journal of Research in Medical Sciences* 2013 **18** 47–51.
 - 27 Tuomainen TP, Virtanen JK, Voutilainen S, Nurmi T, Mursu J, de Mello VD, Schwab U, Hakumaki M, Pulkki K & Uusitupa M. Glucose metabolism effects of vitamin D in prediabetes: the VitDmet randomized placebo-controlled supplementation study. *Journal of Diabetes Research* 2015 **2015** 672653. (<https://doi.org/10.1155/2015/672653>)
 - 28 Foroozanfar F, Talebi M, Samimi M, Mehrabi S, Badehnoosh B, Jamilian M, Maktabi M & Asemi Z. Effect of two different doses of vitamin D supplementation on metabolic profiles of insulin-resistant patients with polycystic ovary syndrome: a randomized, double-blind, placebo-controlled trial. *Hormone and Metabolic Research* 2017 **49** 612–617. (<https://doi.org/10.1055/s-0043-112346>)
 - 29 Matsuda M & DeFronzo RA. Insulin sensitivity indices obtained from oral glucose tolerance testing: comparison with the euglycemic insulin clamp. *Diabetes Care* 1999 **22** 1462–1470. (<https://doi.org/10.2337/diacare.22.9.1462>)
 - 30 Rabasa-Lhoret R & Laville M. How to measure insulin sensitivity in clinical practice? *Diabetes and Metabolism* 2001 **27** 201–208.
 - 31 DeFronzo RA & Tripathy D. Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care* 2009 **32** (Supplement 2) S157–S163. (<https://doi.org/10.2337/dc09-S302>)
 - 32 Amin SN, Hussein UK, Yassa HD, Hassan SS & Rashed LA. Synergistic actions of vitamin D and metformin on skeletal muscles and insulin resistance of type 2 diabetic rats. *Journal of Cellular Physiology* 2018 **233** 5768–5779. (<https://doi.org/10.1002/jcp.26300>)
 - 33 Gilsanz V, Kremer A, Mo AO, Wren TA & Kremer R. Vitamin D status and its relation to muscle mass and muscle fat in young women. *Journal of Clinical Endocrinology and Metabolism* 2010 **95** 1595–1601. (<https://doi.org/10.1210/jc.2009-2309>)
 - 34 Manna P, Achari AE & Jain SK. Vitamin D supplementation inhibits oxidative stress and upregulate SIRT1/AMPK/GLUT4 cascade in high glucose-treated 3T3L1 adipocytes and in adipose tissue of high fat diet-fed diabetic mice. *Archives of Biochemistry and Biophysics* 2017 **615** 22–34. (<https://doi.org/10.1016/j.abb.2017.01.002>)
 - 35 Castro AJ, Frederico MJ, Cazarolli LH, Bretanha LC, Tavares Lde C, Buss Zda S, Dutra MF, de Souza AZ, Pizzolatti MG & Silva FR. Betulinic acid and 1,25(OH)(2) vitamin D(3) share intracellular signal transduction in glucose homeostasis in soleus muscle. *International Journal of Biochemistry and Cell Biology* 2014 **48** 18–27. (<https://doi.org/10.1016/j.biocel.2013.11.020>)
 - 36 Shab-Bidar S, Neyestani TR, Djazayeri A, Eshraghian MR, Houshiarrad A, Kalayi A, Shariatzadeh N, Khalaji N & Gharavi A. Improvement of vitamin D status resulted in amelioration of biomarkers of systemic inflammation in the subjects with type 2 diabetes. *Diabetes/Metabolism Research and Reviews* 2012 **28** 424–430. (<https://doi.org/10.1002/dmrr.2290>)
 - 37 Roy P, Nadeau M, Valle M, Bellmann K, Marette A, Tchernof A & Gagnon C. Vitamin D reduces LPS-induced cytokine release in omental adipose tissue of women but not men. *Steroids* 2015 **104** 65–71. (<https://doi.org/10.1016/j.steroids.2015.08.014>)
 - 38 Utzschneider KM, Pigeon RL, Faulenbach MV, Tong J, Carr DB, Boyko EJ, Leonetti DL, McNeely MJ, Fujimoto WY & Kahn SE. Oral disposition index predicts the development of future diabetes above and beyond fasting and 2-h glucose levels. *Diabetes Care* 2009 **32** 335–341. (<https://doi.org/10.2337/dc08-1478>)
 - 39 DeFronzo RA, Tripathy D, Schwenke DC, Banerji M, Bray GA, Buchanan TA, Clement SC, Henry RR, Kitabchi AE, Mudaliar Set al. Prediction of diabetes based on baseline metabolic characteristics in individuals at high risk. *Diabetes Care* 2013 **36** 3607–3612. (<https://doi.org/10.2337/dc13-0520>)
 - 40 Neelankal John A & Jiang FX. An overview of type 2 diabetes and importance of vitamin D3-vitamin D receptor interaction in pancreatic beta-cells. *Journal of Diabetes and its Complications* 2018 **32** 429–443. (<https://doi.org/10.1016/j.jdiacomp.2017.12.002>)
 - 41 Kajikawa M, Ishida H, Fujimoto S, Mukai E, Nishimura M, Fujita J, Tsuura Y, Okamoto Y, Norman AW & Seino Y. An insulinotropic effect of vitamin D analog with increasing intracellular Ca2+ concentration in pancreatic beta-cells through nongenomic signal transduction. *Endocrinology* 1999 **140** 4706–4712. (<https://doi.org/10.1210/endo.140.10.7025>)
 - 42 Jorde R, Sollid ST, Svartberg J, Schirmer H, Joakimsen RM, Njolstad I, Fuskevag OM, Figenschau Y & Hutchinson MY. Vitamin D 20,000 IU per week for five years does not prevent progression from prediabetes to diabetes. *Journal of Clinical Endocrinology and Metabolism* 2016 **101** 1647–1655. (<https://doi.org/10.1210/jc.2015-4013>)
 - 43 Pittas AG, Dawson-Hughes B, Sheehan P, Ware JH, Knowler WC, Aroda VR, Brodsky I, Ceglia L, Chadha C, Chatterjee Ret al. Vitamin D supplementation and prevention of Type 2 diabetes. *New England Journal of Medicine* 2019 [epub]. (<https://doi.org/10.1056/NEJMoa1900906>)

Received 3 March 2019

Revised version received 13 June 2019

Accepted 3 July 2019