Management of neonates born to women with Graves’ disease: a cohort study

Alix Besançon1, Jacques Beltrand1,2, Isabelle Le Gac1, Dominique Luton3 and Michel Polak1,2

1Endocrinologie Gynécologie Diabétologie Pédiatriques, Hôpital Universitaire Necker Enfants Malades, Assistance Publique-Hôpitaux de Paris, 149 Rue de Sèvres, 75015 Paris France, 2INSERM U1016, IMAGINE Institute, Université Paris Descartes, Sorbonne Paris Cité, Paris, France and 3Département de Maternité Bichat-Beaujon, Assistance Publique-Hôpitaux de Paris, DHU Risque et Grossesse, Université Paris VII, Paris, France

Correspondence should be addressed to M Polak
Email Michel.polak@nck.aphp.fr

Abstract

Objective: Hyperthyroidism in neonates born to mothers with Graves’ disease (GD) can be associated with significant morbidity and mortality, but is still overlooked by clinicians. Management of neonatal hyperthyroidism would be improved by a better understanding of the predictive factors involved. The aim of this study was to evaluate the course of thyroid function and clinical outcomes during the first postnatal month in babies born to mothers with GD.

Design: Prospective observational study.

Methods: Sixty-eight neonates born to mothers with GD were managed from birth and divided into three groups based on thyrotropin receptor antibody (TRAb) and anti-thyroid drug (ATD) status in the mother: TRAb−ve/ATD−ve, n=27; TRAb−ve/ATD+ve, n=8; and TRAb+ve/ATD+ve, n=33. The main outcome measures were clinical examination, thyroid function tests (TSH, free thyroxine (FT4), free triiodothyronine, and TRAb), echocardiography, thyroid ultrasonography, and bone maturation assessment.

Results: None of the infants born to TRAb−ve mothers with GD developed neonatal hyperthyroidism. Of the 33 TRAb+ve/ATD+ve neonates, 24 (72.7%) had positive TRAb on cord blood assays, and seven of these developed neonatal hyperthyroidism. FT4 elevation between days 3 and 7 but not at birth was predictive of the development of hyperthyroidism.

Conclusions: TRAb status should be checked in the third trimester in mothers with GD and on cord blood in their neonates; if positive, it indicates a high risk of neonatal hyperthyroidism. FT4 measurement at birth should be repeated between days 3 and 5 (and by day 7 at the latest); rapid FT4 elevation during the first postnatal week is predictive of hyperthyroidism and warrants ATD therapy.

Introduction

Graves’ disease (GD) is present in about 0.2% of pregnancies. Neonatal autoimmune hyperthyroidism, although seen in only 2% of cases and usually transient (1, 2, 3), is associated with mortality rates of up to 25% and with both immediate and long-term morbidities. Cardiac insufficiency is one of the major risks in these infants. Liver dysfunction (hepatitis and cholestatic jaundice), coagulopathy, pulmonary arterial hypertension, craniosenosis, microcephaly, and psychomotor disabilities may occur in severely affected infants (4, 5, 6, 7). Fetal thyroid dysfunction precedes neonatal hyperthyroidism. Optimal management of GD during pregnancy is crucial to prevent fetal death or permanent neurological impairments (8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19).

Neonatal hyperthyroidism has been evaluated in several studies (2, 3, 7) but recent case reports (20, 21, 22) indicate that this transient disease is still overlooked by clinicians, leading to severe complications that could have been prevented. Better knowledge of predictors of neonatal hyperthyroidism would result in improved preventive and therapeutic management.
The main objective of this observational study was to evaluate the clinical outcomes and course of thyroid function during the first postnatal month in a large prospective cohort of babies born to mothers with GD, with the goal of developing management and follow-up plans for the first month of life.

**Subjects and methods**

**Patients**

This study was approved by the Paris-Saint-Louis Ethics Committee for biomedical studies in humans and was conducted in compliance with French law. Written informed consent was obtained from the pregnant women before study inclusion.

We studied the cohort of pregnant women with GD and their babies from a previous study by our group (17). The women were included prospectively and managed over a 3-year period (between 1999 and 2002) at the Robert Debré Teaching Hospital (Paris, France). The newborns were also managed in the same center. The inclusion criterion was a past or current history of GD diagnosed by an endocrinologist based on clinical and laboratory test evidence of hyperthyroidism with goiter, Graves’ ophthalmopathy, or dermopathy, plus at least one positive test for thyrotropin receptor antibodies (TRAbs) (23, 24). Patients were not selected based on thyroid function, disease activity, or history of thyroidectomy or anti-thyroid drug (ATD) therapy. No exclusion criteria were used. The women were included as early as possible in the pregnancy. We chose a single-center design to ensure that all fetal sonograms were performed by the same person (É Vuillard, see ‘Acknowledgements’ section). In each woman, thyroid function tests and TRAb assays were carried out monthly from study inclusion to delivery. The ATDs and other treatments were adjusted by the endocrinologist to keep the serum free thyroxine (FT4) level at the upper limit of normal (ULN) for pregnant women.

The initial cohort comprised 72 mothers and neonates, of whom four neonates (all born to TRAb-ve/ATD-ve mothers) did not receive follow-up at our institution, which left 68 women for our study. Mean age of the women was 33 years (range, 26–43 years) and gestational age at inclusion was 17 weeks (range, 10–28 weeks). We divided the women into three groups based on the risk of fetal thyroid dysfunction (25); consistently negative TRAb assays in the third trimester of pregnancy but need for ATD therapy (TRAb-ve/ATD+ve group); and at least one positive TRAb assay in the third trimester and need for ATD therapy (TRAb+ve/ATD+ve group). In this third group, fetuses and neonates were considered at risk for thyroid dysfunction. Maternal thyroid function did not differ across these three groups at any time during the pregnancies, indicating appropriate adjustment of ATD and/or levothyroxine (l-T4) treatment. In that study, during the third trimester, none of the pregnant women was TRAb+ve/ATD-ve.

**Methods**

At delivery, cord blood was retrieved for thyroid function tests (TSH, FT4, free triiodothyronine (FT3), and TRAb) and the neonates were examined by a pediatric endocrinologist who recorded predefined criteria (heart rate, congestive heart failure, goiter, hyperexcitability, fever, sweating, voracity, vomiting, weight gain, diarrhea, orbitopathy, hepatosplenomegaly, and jaundice). Cord blood was taken from the clamp of the umbilical cord during delivery and not through amniotic fluid or cord puncture. In neonates born after at least 36 weeks of gestation (WG), normal cord blood values were defined within the 2.5th and the 97.5th percentiles, as follows: FT4 10.4–16.4 pmol/l and TSH 2.6–11.8 mU/l (26). Hypothyroidism was defined as FT4 below the 2.5th percentile and TSH above the 97.5th percentile. Hyperthyroidism was defined as FT4 above the 97.5th percentile and TSH below the 2.5th percentile. Subclinical hypothyroidism was defined as normal serum FT4 and FT3 with TSH elevation above the 97.5th percentile and subclinical hyperthyroidism as normal serum FT4 and FT3 with TSH below the 2.5th percentile. These values were also compared with the values obtained in our cohort (see Table 1 below).

Each neonate was examined again on days 7, 15, and 30. Serum FT4, FT3, and TSH were assayed at the same time points. TRAb was assayed on days 7, 15, and 30 in babies with positive TRAb on cord blood and weekly in those requiring ATD therapy. Echocardiography and thyroid ultrasonography (US) were performed within a few days after birth, by the same ultrasonographer (É Vuillard) (27). Goiter was defined as a thyroid gland volume equal to or greater than the mean +2 S.D. or as a thyroid gland diameter equal to or greater than the 95th percentile (28). Echocardiography was done in 60 of the 68 patients. Bone maturation was assessed by a routine knee radiograph at birth and classified as advanced, delayed, or...
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Table 1 Serum FT₄, FT₃, and TSH values during the first postnatal month in the control group (23 neonates born to TRAb⁻/ATD⁻ mothers). Data are presented as mean (interquartile range) min–max.

<table>
<thead>
<tr>
<th></th>
<th>FT₃ (pmol/l)</th>
<th>FT₄ (pmol/l)</th>
<th>TSH (mU/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td>D1</td>
<td>2.3 (1.9–2.6)</td>
<td>13.8 (13.1–14.6)</td>
<td>8.4 (4.9–10.1)</td>
</tr>
<tr>
<td>D7</td>
<td>6.6 (5.7–7.3)</td>
<td>24.5 (21.5–27.8)</td>
<td>2.9 (1.7–3.5)</td>
</tr>
<tr>
<td>D15</td>
<td>6.6 (5.9–7.2)</td>
<td>18.1 (16.9–20.2)</td>
<td>3 (1.6–3.9)</td>
</tr>
<tr>
<td>D30</td>
<td>7 (6.4–7.7)</td>
<td>16.9 (14.8–18.9)</td>
<td>2.8 (2–3.2)</td>
</tr>
</tbody>
</table>

D, day; FT₃, free triiodothyronine; FT₄, free thyroxine; TSH, thyroid-stimulating hormone.

Table 2 Clinical presentation of the 68 neonates at birth according to the three groups: born to mothers TRAb⁻/ATD⁻ (n = 27), mothers TRAb⁻/ATD⁺ (n = 8), or mothers TRAb⁺/ATD⁻ (n = 33).

<table>
<thead>
<tr>
<th></th>
<th>TRAb⁻/ATD⁻ (n = 27)</th>
<th>TRAb⁻/ATD⁺ (n = 8)</th>
<th>TRAb⁺/ATD⁻ (n = 33)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intrauterine fetal death</td>
<td>0</td>
<td>0</td>
<td>1 (35 WG, due to fetal hyperthyroidism)</td>
<td>a³</td>
</tr>
<tr>
<td>Preterm birth (&lt;37 WG)</td>
<td>3</td>
<td>1 (after fetal blood sampling)</td>
<td>5</td>
<td>1 (NS)</td>
</tr>
<tr>
<td>Very premature birth (&lt;32 WG)</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>a³</td>
</tr>
<tr>
<td>Low BW (&lt;10th percentile)</td>
<td>2</td>
<td>0</td>
<td>6</td>
<td>0.27 (NS)</td>
</tr>
<tr>
<td>Birth length (&lt;10th percentile)</td>
<td>1</td>
<td>0</td>
<td>4</td>
<td>0.02</td>
</tr>
<tr>
<td>Abnormal symptoms</td>
<td>2³</td>
<td>2</td>
<td>4</td>
<td>0.005</td>
</tr>
<tr>
<td>Clinical goiter</td>
<td>0</td>
<td>0</td>
<td>13</td>
<td>a³</td>
</tr>
<tr>
<td>Goiter (ultrasonography)</td>
<td>3</td>
<td>4</td>
<td>8 (including one with tetralogy of Fallot, one with left ventricular hypertrophy and pericardial detachment, and one with ventricular dysfunction)</td>
<td>a³</td>
</tr>
<tr>
<td>Congenital heart defects (echocardiography)</td>
<td>2 (PFO in premature infants)</td>
<td>3 (two atrial septal defects and one ventricular septal defect)</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td>Other congenital birth defects</td>
<td>1 congenital pulmonary airway malformation</td>
<td>0</td>
<td>0</td>
<td>a³</td>
</tr>
<tr>
<td>Bone maturation⁴</td>
<td>A = 0/N = 23/D = 1/NA = 3</td>
<td>N = 8</td>
<td>A = 1/N = 24/D = 2/NA = 6</td>
<td>1 (NS)</td>
</tr>
<tr>
<td>Craniosynostosis</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td></td>
</tr>
</tbody>
</table>

WG, weeks of gestation; BW, birth weight; ATD, anti-thyroid drug; TRAb, TSH receptor antibody; PFO, patent foramen ovale; A, advanced bone maturation; N, normal bone maturation; D, delayed bone maturation; NA, not available; NS, nonsignificant.

*aToo few events to allow statistical tests.
*bVomiting and hypotonia (n = 2).
*cHypotonia (n = 2), bradycardia (n = 1), hyperexcitability (n = 1), and aplasia cutis congenital of the scalp (n = 1).
*dAssessed by routine knee radiography at birth.

Statistical analyses were carried out using JMP Software, version 10.0.0 (SAS corporation, Tulsa, OK, USA). The data were described as mean ± S.D. and 95% CI or interquartile range. We verified that data distribution was normal. We compared the three groups (TRAb⁻/ATD⁻, TRAb⁻/ATD⁺, and TRAb⁺/ATD⁺) using χ² tests or Fisher’s exact test when needed. Spearman’s rank correlation test was performed to assess correlations between variables. The values of P < 0.05 were considered statistically significant.

Results

Clinical description of the newborns at birth

There were 27 TRAb⁻/ATD⁻, 8 TRAb⁻/ATD⁺, and 33 TRAb⁺/ATD⁺ newborns and their clinical presentation is shown in Table 2. For each group mean (range), birth weight (BW) was 3250 (1850–4830), 3102 – 4470, and 3175 – 4800 g, respectively. The data were described as mean ± standard deviation (S.D.) and 95% confidence interval (CI).

As an antibody titer > 2 IU/l. The results are reported as multiples of the ULN (X·ULN).

normal (29). Cranial radiographs were obtained routinely to look for craniosynostosis.

Serum TSH, FT₃, and FT₄ were measured using a chemiluminescence immunoassay with the ACS-180SE system (Bayer Diagnostics) (26). TRAb was measured using a RIA with second-generation antibodies (RIA-2 Dynotest TRAK human, BRAHMS Diagnostica GmbH, Berlin, Germany) (23). A positive result (TRAb⁺) was defined as an antibody titer ≥ 2 IU/l. The results are reported as multiples of the ULN (X·ULN).
(2910–4040), and 2921 (1110–4070) grams; mean gestation 38.5, 38.5, and 37.7 weeks respectively. Premature birth, low-BW, and bone maturation did not differ across the three groups. The single very preterm baby (29 WG) was in the TRAb+ve/ATD+ve group. One fetus in the TRAb+ve/ATD+ve group died in utero at 35.5 WG from congestive heart failure due to hyperthyroidism. A small area of congenital aplasia cutis on the scalp was found in one TRAb+ve/ATD+ve neonate.

Goiter was only detected by clinical examination in the TRAb−ve/ATD+ve and TRAb+ve/ATD+ve groups; but in all three groups and with a much higher frequency on neonatal US (Table 2). In the TRAb+ve/ATD+ve group, three patients had severe cardiac abnormalities (30, 31, 32) namely, tetralogy of Fallot (mother given L-T4 after surgery in one; and FT4 8.2 pmol/l, FT3 2.6 pmol/l, and TSH 6.65 mU/l in the other – when compared with our laboratory reference ranges and with the values of the TRAb−ve/ATD−ve group presented in Table 1. All the remaining neonates (n = 65) had normal thyroid values in cord blood at delivery.

Cord blood samples at delivery from the 35 neonates born to TRAb−ve mothers were consistently negative for TRAb (n = 27 and n = 8). Of the 33 neonates in the TRAb+ve/ATD+ve group, 24 (72.7%) had TRAb+ve cord blood assays. Neonatal hyperthyroidism developed in seven of these 24 neonates (Fig. 1) and in none of the nine cord blood cases who were negative for TRAb (see next paragraph). We found a perfect correlation between TRAb assays in cord blood in the newborns and TRAb assays in their mothers at the third trimester of pregnancy when available (r² = 0.99; P < 0.01) and we also found the same correlation if we consider only the third group of TRAb+ve/ATD+ve mothers and their newborns (r² = 0.99; P < 0.01). We found a good positive correlation between the TRAb titers in cord blood and the development of neonatal hyperthyroidism (r² = 0.61; P < 0.01). Figure 2 shows the TRAb titers in cord blood in the three groups. The positive predictive value of cord blood TRAb is 29% (7/24), the sensitivity is 100% and the specificity is 35%. Of the 61 neonates with normal thyroid function, 17 had positive TRAb cord blood samples with a mean titer of 2.4 ×-ULN (from 1.1 to 6.4 ×-ULN). The group of 27 neonates born to TRAb−ve/ATD−ve mothers did not develop thyroid disease during neonatal follow-up.

Thyroid function in the neonates over the first postnatal month

Between days 2 and 15 after birth, hyperthyroidism was diagnosed in seven neonates, based on laboratory tests in the absence of clinical symptoms, all of whom were in the TRAb+ve/ATD+ve group. Treatment was given when FT4 levels were > 35 pmol/l (19). None of the neonates in the

Figure 1
Evolution of thyroid function of the 33 neonates born to mothers positive for TRAb, in cord blood assays (at day 1) and during the neonatal period.
other two groups had hyperthyroidism or hypothyroidism, and none was treated.

The characteristics of the seven neonates with hyperthyroidism are presented in Table 3. Cord blood tests (TSH, FT3, and FT4) showed subclinical hyperthyroidism in only three of these seven neonates and hypothyroidism (or low FT4) in two of these. Thus, cord blood test results failed to predict subsequent thyroid function. Mean age at diagnosis was 5 days. Of the seven neonates with hyperthyroidism, three had received prenatal treatment for hypothyroidism (two maternal treatment and one intraamniotic \( I-T_4 \) injection) and two for hyperthyroidism (maternal treatment). Mean FT4 and TSH levels were 46.5 \( \pm \) 13.8 pmol/l and 0.22 \( \pm \) 0.13 mU/l, respectively, at treatment initiation, and all seven neonates had positive cord blood tests for TRAb with a mean titer of 13.1 \( \times \)-ULN (2–39.6 \( \times \)-ULN). Carbimazole (the methimazole pro-drug) was given in a daily dosage of 1 mg/kg for a mean duration of 5 weeks. In addition, two neonates whose FT4 levels fell below the normal range for age received concomitant L-T4.

According to our study protocol, the first thyroid function evaluation after birth was to occur on day 7. However, in two neonates, FT4 and TSH were also assayed on days 3 and 5 because cord blood data had shown hypothyroidism and led the physicians to control FT4 and TSH earlier than the protocol. As shown in Fig. 3, FT4 in both neonates was normal at birth (in the lowest values of normal) then increased markedly between days 3 and 5 compared with values in the TRAb\(^{-}\)ve/ATD\(^{-}\)ve group. Thus, rapid FT4 elevation during the first postnatal week may predict hyperthyroidism.

Clinical course of the neonates over the first month

Weight gain during the 1st week did not differ between neonates with and without hyperthyroidism. None of the seven neonates with hyperthyroidism developed clinical symptoms of hyperthyroidism.

Discussion

The main findings of our study were as follows: i) positive TRAb tests in the mother during pregnancy indicates a risk of hyperthyroidism in the neonate; ii) persistently negative TRAb testing during pregnancy, with or without

Table 3 Characteristics of the seven neonates with transient hyperthyroidism.

<table>
<thead>
<tr>
<th>Patient</th>
<th>TRAb IU/l (( \times )-ULN)</th>
<th>Goiter during third trimester</th>
<th>Goiter on D1 clinical/US</th>
<th>FT4 (pmol/l) on D1</th>
<th>FT3 (pmol/l) on D1</th>
<th>TSH (mU/l) on D1</th>
<th>CMZ treatment duration</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>23.5 (14.7) 90th percentile</td>
<td>No/yes</td>
<td>9.7</td>
<td>1.2</td>
<td>5.03</td>
<td>D3–D30</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>24 (2.7) Yes</td>
<td>Yes/yes</td>
<td>12.8</td>
<td>2</td>
<td>10.5</td>
<td>D7–D30</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>46 (5.1) No</td>
<td>Yes/yes</td>
<td>9.6</td>
<td>2.3</td>
<td>20.2</td>
<td>D5–D30</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>63 (39.6) Yes</td>
<td>Yes/yes</td>
<td>8.2</td>
<td>2.6</td>
<td>6.65</td>
<td>D2–D40</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>40 (25) Yes</td>
<td>No/yes</td>
<td>12.9</td>
<td>3.1</td>
<td>0.05</td>
<td>D4–M2</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>5.2 (3.3) 90th percentile</td>
<td>No/yes</td>
<td>16.4</td>
<td>2.1</td>
<td>1.95</td>
<td>D5–D30</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>28 (2) No</td>
<td>Yes/yes</td>
<td>20.5</td>
<td>NA</td>
<td>48</td>
<td>D15–D30</td>
<td></td>
</tr>
</tbody>
</table>

TRAb, TSH receptor antibody; ULN, upper limit of normal; D, day; US, ultrasonography; CMZ, carbimazole (methimazole pro-drug); FT4, serum free thyroxine concentration; TSH, serum thyroid-stimulating hormone concentration; NA, not available; FT3, serum free triiodothyronine.
ATD treatment, is not followed by neonatal hyperthyroidism; iii) a positive TRAb test on cord blood indicates a significant risk of neonatal hyperthyroidism (29%); iv) cord blood FT4 and TSH levels mirrored the fetal thyroid function but not the post-natal risk of neonatal hyperthyroidism; and v) rapid FT4 elevation during the first postnatal week may predict hyperthyroidism and warrants consideration of ATD therapy.

All seven neonates who developed hyperthyroidism had positive cord blood TRAb tests with values >2 × ULN, in keeping with earlier studies (33, 34, 35). It has been reported that TRAb levels above ~40 IU/l using the BRAHMS method were predictive of neonatal hyperthyroidism (33). However, in our study, some neonates who developed hyperthyroidism had cord blood TRAb titers of 20 IU/l, leading us to question this threshold. The TRAb values were also reported as multiples of the ULN in order to be comparable if the study was replicated with other assays. Of the seven neonates with hyperthyroidism, four had low TRAb titers (<6 × ULN) but we found a positive correlation \( r^2 = 0.61 \) between the TRAb titers in cord blood and the development of neonatal hyperthyroidism. Thus, a positive TRAb assay may be a better predictor of neonatal hyperthyroidism than a high TRAb titer. We found a good correlation between TRAb assays in cord blood at delivery in neonates of TRAb+ve/ATD+ve mothers and TRAb assays in their mothers at the third trimester of pregnancy. Thus, if cord blood is not available, the positiveness of TRAb in mothers may identify the subgroup of neonates at risk of neonatal hyperthyroidism and thus can be used as a substitute. The test (cord blood TRAb) as it is, with a very high sensitivity (100%) and a low specificity (35%), is still useful for clinicians. Indeed we do not wish to miss any of those newborns with positive TRAb.

ATDs are cleared from the body within a few days, whereas TSH-receptor antibodies persist and may result in hyperthyroidism until the TRAb test becomes negative (mean duration of treatment of 5 weeks) (34). Therefore, even TRAb+ve neonates without biochemical or clinical evidence of thyroid dysfunction at birth should have weekly clinical and laboratory follow-up until the TRAb test becomes negative.

Weight gain in the first few postnatal days did not predict hyperthyroidism. The prevalence of congenital heart defects in the two groups of neonates born to TRAb–ve mothers was similar to that in the general population (30, 31, 32). Among the neonates born to TRAb+ve mothers, two had unusual cardiac abnormalities, both mothers having required ATD therapy during pregnancy. A possible link between ATD therapy and the cardiac malformations is unclear and needs to be ascertained in larger databases. Bone maturation mirrored fetal thyroid function but did not predict neonatal thyroid dysfunction.

Antenatal US can detect goiter in fetuses that will finally be diagnosed as TRAb+ve. Anyway, most of the time this exam can be useful to detect fetuses that will develop neonatal hyperthyroidism. We found that neonatal US is far superior to clinical examination at birth and that goiter from TRAb+ve mothers was similar to that in the general population (30, 31, 32). Among the neonates born to TRAb+ve mothers, two had unusual cardiac abnormalities, both mothers having required ATD therapy during pregnancy. A possible link between ATD therapy and the cardiac malformations is unclear and needs to be ascertained in larger databases. Bone maturation mirrored fetal thyroid function but did not predict neonatal thyroid dysfunction.

None of the neonates with negative TRAb cord blood samples developed hyperthyroidism, even in the group born to TRAb+ve/ATD+ve mothers. Consequently, cord blood TRAb–ve neonates can receive standard follow-up without special thyroid function monitoring.

In contrast to TRAb status at birth, thyroid function tests carried out on cord blood did not predict neonatal hyperthyroidism and only mirrored fetal thyroid function. Thus, FT4 and FT3 on day 1 were not correlated with FT4 and FT3 at the time of diagnosis of hyperthyroidism, consistent with previous studies (16). A new finding from our study is that FT4 elevation on days 3–5 in two neonates from TRAb+ve mothers, monitored for suspected hypothyroidism at birth, predicted neonatal hyperthyroidism (Fig. 3). The FT4 increase and TSH decrease developed gradually, after day 3 but before day 15 in neonates who subsequently developed hyperthyroidism.
Unfortunately, we did not anticipate this early FT₄ elevation and had scheduled thyroid function to be measured only on day 7 of life in our protocol. None of the seven neonates with hyperthyroidism had clinical symptoms suggests that our threshold for treatment (FT₄ > 35 pmol/l) was effective in preventing the development of clinical hyperthyroidism and its potentially serious complications. Thus, we recommend that all women with GD (active and inactive) should have TRAb tested during the third trimester to inform neonatal management; that TRAb status should be checked on cord blood; and that all neonates born to TRAb-positive mothers should have thyroid function tests (FT₄, FT₃, and TSH) carried out at birth and repeated between days 3 and 5 (in any case before day 7) or sooner if there are symptoms. FT₄ elevation above 35 pmol/l on one or both assays may warrant ATD therapy (3, 16).

Last, the FT₄, FT₃, and TSH values in the full-term neonates born to TRAb⁻/ve/ATD⁻/ve mothers (Table 1) are consistent with the previously described time-course of FT₄, FT₃, and TSH values in neonates, once gestational age at birth is taken into account (25, 36, 37). When interpreting thyroid function data in neonates, the values must be compared with age-specific normal ranges.

Conclusion

In neonates born to mothers with GD with positive TRAb and having positive cord blood TRAb assays, thyroid function tests should be carried out on cord blood and then in serum between days 3 and 5, and the results should be compared with age-specific normal values. FT₄ elevation detected by these early tests should prompt ATD therapy. Further work is needed to validate our approach, thereby laying the groundwork for developing practice guidelines.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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