Clinical Study

Umbilical cord levels of sclerostin, placental weight, and birth weight are predictors of total bone mineral content in neonates

Kristin Godang1, Kathrine Frey Frosli2,3, Tore Henriksen4,5, Gunhild A Isaksen1, Nanna Voldner4, Tove Lekva1,6, Thor Ueland6 and Jens Bollerslev1,5

1Section of Specialized Endocrinology, Department of Endocrinology, Oslo University Hospital Rikshospitalet, PO Box 4950 Nydalen, Oslo 0424, Norway, 2Department of Biostatistics, Institute of Basic Medical Sciences, University of Oslo, Oslo, Norway, 3Norwegian Resource Center for Women’s Health, Oslo University Hospital, Oslo, Norway, 4Division of Obstetrics and Gynecology, Oslo University Hospital Rikshospitalet, Oslo, Norway, 5Faculty of Medicine, University of Oslo, Oslo, Norway and 6Faculty of Medicine, Research Institute for Internal Medicine, University of Oslo, Oslo, Norway

(Correspondence should be addressed to K Godang; Email: kgodang@ous-hf.no)

Abstract

Context: During pregnancy, changes occur in the maternal calcium homeostasis to fulfill fetal demand. We hypothesized that the fibroblast growth factor 23 (FGF23) system and Wnt signaling pathway are important for normal skeletal development in the offspring.

Aims: Circulating α-klotho, FGF23, sclerostin, and 25-hydroxyvitamin D (25(OH)D) at the fetal and maternal sides of the placenta were measured to investigate associations with newborn bone mass independent of maternal BMI, calcium and phosphate levels, placental weight, and birth weight.

Methods: In a prospective cohort of healthy pregnant women, the total body bone mineral content (BMC) in 202 newborns was measured by dual-energy X-ray absorptiometry. Maternal circulating levels of the biomarkers were measured at gestational weeks 30–32 and in umbilical cord plasma (UCP) at birth.

Results: Mean α-klotho and sclerostin concentrations in the UCP were significantly higher than maternal levels (3004 vs 1077 pg/ml; *P* < 0.001 and 629 vs 346 pg/ml; *P* < 0.001 respectively), and mean 25(OH)D was lower (31 vs 45 nmol/l; *P* < 0.001). The UCP and maternal FGF23 levels were similar. No significant effects of maternal biomarkers on BMC were found in regression analyses. Among UCP biomarkers, only UCP sclerostin was significantly associated with BMC in univariate analyses, and the effect remained significant after adjustment for birth weight and other confounders.

Conclusions: We found that UCP sclerostin levels, birth weight, and placental weight were significant predictors of neonatal BMC but found no evidence for a main role of maternal levels of α-klotho, FGF23, sclerostin, or 25(OH)D nor of UCP levels of α-klotho, FGF23, or 25(OH)D.

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Introduction

During pregnancy, significant changes occur in the maternal bone metabolism and calcium homeostasis to fulfill the fetal demand for calcium. Adequate mineralization of the skeleton relies on the following three related parameters: maternal food intake, the function of the placenta, and the ability of the fetus to use the nutritional supplies (1).

The regulation of maternal placental–fetal mineral homeostasis and skeletal development has remained largely unknown. Generally, there are increases in both the calcium absorption from the gut and bone resorption in the mother to meet the demand for calcium transfer to the growing child. Maternally regulated cytokines and hormones might influence the placental to fetal calcium transport by influencing the ambient maternal blood calcium levels and by a direct effect on the placental function (2). Calcium is actively transported across the placenta, and the fetal serum calcium and inorganic phosphate (Pi) levels are higher than those on the maternal side (3, 4).

Biochemical markers of the maternal bone turnover indicate that bone remodeling is low during the first half of the pregnancy and may increase in the third trimester (5, 6, 7). Fibroblast growth factor 23 (FGF23) is expressed predominantly in osteoblasts and osteocytes (8). Recent observations suggested that FGF23 is a potent hormonal factor that regulates inorganic Pi and 1,25-dihydroxyvitamin D3 (9). However, the role of FGF23 protein in neonatal mineral homeostasis is unknown. A previous study suggested that FGF23 might significantly influence the Pi homeostasis in healthy term infants (10). FGF23 binds to its receptor together with the obligatory co-receptor, α-klotho, which is expressed in other tissues including the placenta (11). Notably, α-klotho appears to be predominantly expressed in syncytiotrophoblasts with
some expression in the endothelium of fetal vessels (12). If the syncytiotrophoblasts can secrete and activate fetal α-klotho, it could contribute to elevated levels of α-klotho which would be found in the umbilical cord plasma (UCP).

Multiple studies have demonstrated a pivotal role of the Wnt signaling pathway for bone development, mass, and structure (13, 14). Recently, the importance of the osteocyte product sclerostin for regulation of the Wnt pathway was established based on genetic studies of patients with high bone mass (15). Sclerostin is a glycoprotein that inhibits osteoblast differentiation and bone formation (16). Although the underlying mechanisms are unclear, it was hypothesized that sclerostin has an inhibitory effect on bone formation by directly blocking the Wnt signaling pathway (17). Thus far, the sclerostin levels have not been investigated in the placenta or in human newborns.

We have recently established a cohort of well-described normal newborn babies (18) who were recruited from a prospective study of normal pregnancies, the STORK study (19). We hypothesized that the FGF23 system and the Wnt signaling pathway are important for the development of a normal skeleton in newborn babies. The aims of this study were to determine the circulating levels of α-klotho, FGF23, sclerostin, and 25-hydroxyvitamin D$_3$ (25(OH)D) at the fetal and maternal side of the placenta and to test whether these variables were related to the newborns’ bone mass independent of the mothers’ early pregnancy BMI, maternal circulating levels of calcium and Pi, the placental weight, and birth weight.

Materials and methods

The present work was performed in a subsample of the STORK study (19). STORK is a prospective cohort study of healthy women with Scandinavian heritage who registered for obstetric care at Oslo University Hospital Rikshospitalet, from 2001 to 2008 (n = 1031). The exclusion criteria were multiple pregnancies, known pre-gestational diabetes, and severe chronic diseases (lung, cardiac, gastrointestinal, or renal). The women were scheduled for four antenatal visits at gestational weeks 14–16, 22–24, 30–32, and 36–38. The maternal height (self-reported) was obtained at the first visit, and the weight was measured at each visit. The BMI was calculated as weight (in kilograms) divided by height (in meters) squared. In the analyses, only the BMI from the first visit was used as this was the best estimate for the women’s normal or pre-gestational body composition. At birth, measurements of birth weight and placental weight were recorded.

We invited 234 women from the cohort and their babies (2005–2008) to participate in a substudy of the newborns’ total bone mineral content (BMC) as measured by dual-energy X-ray absorptiometry (DXA) (18). Women with a preterm birth, defined as < 37 weeks of gestation, were excluded before the analyses (20). This resulted in a study sample of n = 202 (Fig. 1).

For the DXA analyses, a narrow fan-beam GE Lunar Prodigy Densitometer (GE Medical Systems, Lunar Corp., Madison, WI, USA; software version 12.10) was used. The total body BMC and total body bone mineral density (BMD) were measured. The scanning procedure has been described in detail previously (18). According to the manufacturer, the coefficients of variation (CV%) for the Lunar DPX-L instrument (regarded by the manufacturers to be similar to the Lunar Prodigy) is 1.1% for the total body BMC (21).

Ethics

The study was approved by the Regional Ethics Committee and performed according to the Declaration of Helsinki. All women provided written informed consent.

Biochemical variables

The maternal circulating levels of α-klotho, FGF23, sclerostin, and 25(OH)D were measured in blood samples at gestational weeks 30–32. These measurements were also performed in the UCP. Maternal levels of total calcium, Pi, and fasting glucose were analyzed in blood samples at gestational weeks 30–32.

The maternal blood samples were drawn between 0730 and 0830 h after an overnight fast. The samples were obtained by a venipuncture into vacutainer tubes. Vacutainer tubes containing EDTA were kept on ice before centrifugation (2500 g and 4 °C for 25 min) and the plasma was frozen at −80 °C in aliquots within 1 h of collection until analysis. The umbilical cord blood was collected by the midwife into EDTA tubes, centrifuged for plasma separation, and placed at −20 °C for less than a month and stored long term at −80 °C.

The plasma levels of α-klotho and intact FGF23 were measured by two-site ELISA using commercially available kits from Immuno-Biological Laboratories (IBL, Gunma, Japan) and Kainos Laboratories (Tokyo, Japan) respectively (22, 23). The plasma levels of

![Flow chart showing the selection of the study sample.](https://www.eje-online.org)
sclerostin were analyzed by a 2nd Generation High Sensitive ELISA reagent (Quidel Corporation, San Diego, CA, USA). For this assay, the sensitivity was defined as the lowest standard value. 33 pg/ml. All the samples were analyzed in duplicate from a given mother and her baby in the same microtiter plate to minimize run-to-run variability. All samples had detectable levels of sclerostin. Serum 25(OH)D was measured using a RIA from DiaSorin (Stillwater, MN, USA).

All assays were performed according to the manufacturer’s instructions. The intra- and interassay CV were <10% for all assays. The serum levels of calcium and Pi were analyzed using standard automatic analyzer techniques at the accredited laboratory at Oslo University Hospital Rikshospitalet, Norway.

The placental weight was measured before cutting the umbilical cord. The tissue samples were snap frozen in liquid nitrogen within 30 min and stored at −80 °C.

RNA isolation from placentas

The samples were pulverized with a mortar in liquid nitrogen, and TRIzol reagent (Invitrogen) was added. The samples were homogenized, and the RNA was purified according to the manufacturer’s instructions using a RNaseasy micro kit (Qiagen). The integrity was assessed using an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA), and the concentrations were determined by OD readings on a Nanodrop ND-1000 spectrophotometer (Nanodrop Technologies, Wilmington, DE, USA).

Real-time RT-PCR

RT was performed using a High-Capacity cDNA Archive Kit (Applied Biosystems) with 1 μg total RNA. For real-time RT-PCR, sequence-specific oligonucleotide primers were designed using Primer Express software version 2.0 (Applied Biosystems). Primers are available on request.

Gene expression

The gene expression of FGF23, sclerostin-SOST, α-klotho, and β-actin was measured in tissue samples from two placenta. The gene products of the primers for FGF23 and SOST were validated with other cell types.

Statistical analysis

Descriptive statistics are presented as the means and S.D. according to the distributional properties of the data. Comparisons of the study sample and the remaining participants in the STORK study with term births were performed by independent sample t-tests or Pearson’s χ²-tests, as appropriate. Comparisons of the maternal circulating levels of α-klotho, FGF23, sclerostin and 25(OH)D at gestational weeks 30–32 and of the same values from the UCP at birth were performed by paired sample t-tests.

Estimation of the effects of α-klotho, FGF23, sclerostin, and 25(OH)D maternal circulating levels at gestational weeks 30–32 or UCP levels at birth on BMC was performed by linear regression analyses, with standardized regression coefficients as the effect measures. The unadjusted and adjusted effect estimates were obtained from univariate and multiple regression analyses respectively.

Multiple linear regression analyses were used to estimate the adjusted effects of α-klotho, FGF23, sclerostin, and 25(OH)D on the BMC after adjusting for potential confounding variables. The analyses were checked for violations of the assumptions in linear regression models.

A P value <0.05 was considered statistically significant. All analyses were performed with SPSS version 18.0 (SPSS, Inc., Chicago, IL, USA).

Results

Cohort characteristics

Characteristics of the study sample are given in Table 1. The characteristics of the neonates in the study sample were not significantly different from the other neonates born at term in the total STORK cohort, but the pregnant women in this study had a significantly lower BMI at gestational weeks 14–16 (data not shown).

Table 1 Sample characteristics of the pregnant women and their infants.

<table>
<thead>
<tr>
<th></th>
<th>Women (n=202)</th>
<th>Infants (n=202)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (S.D.)</td>
<td>Mean (S.D.)</td>
</tr>
<tr>
<td>BMI (weeks 14–16; kg/m²; self-reported)</td>
<td>23.9 (3.8)</td>
<td>40.2 (1.2)</td>
</tr>
<tr>
<td>Gestational age at birth (weeks)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Placental weight (g)</td>
<td>705 (162)</td>
<td>3612 (480)</td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Scan day (after birth)</td>
<td>1.8 (1.0)</td>
<td>3408 (474)</td>
</tr>
<tr>
<td>Scan day weight (g)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total body BMC (g)</td>
<td>93 (12)</td>
<td>0.345 (0.042)</td>
</tr>
<tr>
<td>Total body BMD (g/cm²)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total calcium (mmol/l)&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.10 (0.09)</td>
<td>1.2 (0.2)</td>
</tr>
<tr>
<td>Phosphate (mmol/l)&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1077 (586)</td>
<td>3004 (1535)</td>
</tr>
<tr>
<td>α-Klotho (pg/ml)&lt;sup&gt;c&lt;/sup&gt;</td>
<td>263 (115)</td>
<td>255 (125)</td>
</tr>
<tr>
<td>Sclerostin (pg/ml)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>346 (153)</td>
<td>629 (295)</td>
</tr>
<tr>
<td>25(OH)D (nmol/l)&lt;sup&gt;e&lt;/sup&gt;</td>
<td>45 (18)</td>
<td>31 (18)</td>
</tr>
</tbody>
</table>

BMC, bone mineral content; BMD, bone mineral density; FGF23, fibroblast growth factor 23; 25(OH)D, 25-hydroxyvitamin D<sub>3</sub>.

<sup>a</sup>n may vary due to missing values.

<sup>b</sup>Born at term, defined as birth at gestational week 37 or later.

<sup>c</sup>Comparison of maternal and neonatal values.

<sup>d</sup>MATernal values are obtained from blood samples at gestational weeks 30–32.
Circulating levels of α-klotho, FGF23, sclerostin, and 25(OH)D

The mean α-klotho and sclerostin concentrations in the UCP were significantly higher than the mean maternal circulating levels (3004 vs 1077 pg/ml; P < 0.001 and 629 vs 346 pg/ml; P < 0.001 respectively), and the mean 25(OH)D concentration in UCP was significantly lower than in maternal circulation (31 vs 45 nmol/l; P < 0.001; Table 1 and Fig. 2). The mean FGF23 concentration in the UCP and the maternal circulating levels were not significantly different (Table 1 and Fig. 2).

Gene expression

The gene expression of FGF23, sclerostin, and α-klotho was analyzed in two placental tissue samples, and we found α-klotho to be highly expressed, whereas no gene expression of FGF23 and sclerostin was detected (data not shown).

Analyses of the effect of the maternal circulating levels of α-klotho, FGF23, sclerostin, and 25(OH)D on BMC

Univariate analyses showed no statistically significant associations between the maternal circulating levels of α-klotho, FGF23, sclerostin, or 25(OH)D on the neonatal BMC (Table 2). We used directed acyclic graphs as a regression modeling tool to evaluate the need for adjustments for other variables in the subsequent multiple regression analyses (24). As mentioned below, several variables were considered in the graphs, but variables that were assumed to be predictive for only one of the variables in the above paragraph were left out of the analysis. When in doubt, we included the variables in supplementary analyses.

Based on the literature, we hypothesized that the early pregnancy BMI might influence both the circulating fetal levels of α-klotho, FGF23, and sclerostin in late gestation (25, 26) and the neonatal BMC (26, 27). Thus, BMI was included as a potential confounding variable (28). Maternal circulating levels of calcium, Pi, and 25(OH)D were included as potential confounders in the analyses, by similar argumentation.

In supplementary analyses, adjustments were also performed for the placental weight by assuming that the placental size at birth reflects the size of the placenta at weeks 30–32 and that the placenta may potentially influence the maternal circulating levels of α-klotho, FGF23, sclerostin, or 25(OH)D and the neonatal BMC. Multiple adjustments did not alter the conclusion of the lack of statistical significance for independent direct effects of the maternal circulating levels of α-klotho, FGF23, sclerostin, or 25(OH)D on the neonatal BMC (Table 2).

Analyses of the effect of the UCP levels of α-klotho, FGF23, sclerostin, and 25(OH)D on the BMC

Univariate analyses revealed no significant associations between the UCP levels of α-klotho, FGF23, or 25(OH)D on neonatal BMC, but a significant association between sclerostin and BMC was found (Table 2).
Based on the assumptions that the early pregnancy BMI, maternal circulating levels of calcium and Pi, maternal or UCP levels of 25(OH)D, and birth weight or placental weight might influence both the neonatal BMC and the UCP levels of α-klotho, FGF23, and sclerostin, we chose to include these variables in the analyses as potential confounders. The effect estimates for α-klotho, FGF23, or 25(OH)D remained nonsignificant after adjustments for these confounding factors (Table 2).

<table>
<thead>
<tr>
<th>Maternal variables</th>
<th>Univariate</th>
<th>Multiple</th>
</tr>
</thead>
<tbody>
<tr>
<td>α-Klotho (weeks 30–32)</td>
<td>0.12</td>
<td>0.07</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.08</td>
<td>0.37</td>
</tr>
<tr>
<td>25(OH)D</td>
<td>0.28</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Calcium</td>
<td>−0.11</td>
<td>0.13</td>
</tr>
<tr>
<td>Phosphate</td>
<td>−0.04</td>
<td>0.57</td>
</tr>
<tr>
<td>FGF23 (weeks 30–32)</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.86</td>
<td>0.75</td>
</tr>
<tr>
<td>25(OH)D</td>
<td>0.29</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Calcium</td>
<td>−0.11</td>
<td>0.12</td>
</tr>
<tr>
<td>Phosphate</td>
<td>−0.04</td>
<td>0.56</td>
</tr>
<tr>
<td>Sclerostin (weeks 30–32)</td>
<td>0.01</td>
<td>0.03</td>
</tr>
<tr>
<td>BMI (weeks 14–16; kg/m²; self-reported)</td>
<td>0.91</td>
<td>0.70</td>
</tr>
<tr>
<td>25(OH)D</td>
<td>−0.12</td>
<td>0.11</td>
</tr>
<tr>
<td>Calcium</td>
<td>−0.04</td>
<td>0.60</td>
</tr>
<tr>
<td>Phosphate</td>
<td>−0.01</td>
<td>0.95</td>
</tr>
<tr>
<td>25(OH)D (weeks 30–32)</td>
<td>−0.11</td>
<td>0.12</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.14</td>
<td>0.29</td>
</tr>
<tr>
<td>Calcium</td>
<td>−0.04</td>
<td>0.58</td>
</tr>
<tr>
<td>Phosphate</td>
<td>−0.01</td>
<td>0.96</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Neonatal variables</th>
<th>Univariate</th>
<th>Multiple</th>
</tr>
</thead>
<tbody>
<tr>
<td>α-Klotho (UCP)</td>
<td>0.09</td>
<td>0.06</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.30</td>
<td>0.46</td>
</tr>
<tr>
<td>25(OH)D (UCP)</td>
<td>0.12</td>
<td>0.10</td>
</tr>
<tr>
<td>25(OH)D (weeks 30–32)</td>
<td>0.04</td>
<td>0.59</td>
</tr>
<tr>
<td>Calcium</td>
<td>0.01</td>
<td>0.87</td>
</tr>
<tr>
<td>Phosphate</td>
<td>−0.01</td>
<td>0.95</td>
</tr>
<tr>
<td>Placental weight (g)</td>
<td>0.51</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>FGF23 (UCP)</td>
<td>−0.07</td>
<td>0.21</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.42</td>
<td>0.11</td>
</tr>
<tr>
<td>25(OH)D (UCP)</td>
<td>0.13</td>
<td>0.11</td>
</tr>
<tr>
<td>25(OH)D (weeks 30–32)</td>
<td>0.09</td>
<td>0.23</td>
</tr>
<tr>
<td>Calcium</td>
<td>−0.02</td>
<td>0.85</td>
</tr>
<tr>
<td>Phosphate</td>
<td>0.00</td>
<td>0.96</td>
</tr>
<tr>
<td>Placental weight (g)</td>
<td>0.49</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Sclerostin (UCP)</td>
<td>0.26</td>
<td>0.24</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.02</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>25(OH)D (UCP)</td>
<td>0.24</td>
<td>0.25</td>
</tr>
<tr>
<td>25(OH)D (weeks 30–32)</td>
<td>−0.12</td>
<td>0.06</td>
</tr>
<tr>
<td>Calcium</td>
<td>0.05</td>
<td>0.39</td>
</tr>
<tr>
<td>Phosphate</td>
<td>0.02</td>
<td>0.77</td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td>0.72</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>25(OH)D (UCP)</td>
<td>0.13</td>
<td>0.20</td>
</tr>
<tr>
<td>BMI (weeks 14–16)</td>
<td>0.13</td>
<td>0.04</td>
</tr>
<tr>
<td>Calcium</td>
<td>−0.02</td>
<td>0.83</td>
</tr>
<tr>
<td>Phosphate</td>
<td>−0.01</td>
<td>0.90</td>
</tr>
<tr>
<td>25(OH)D (weeks 30–32)</td>
<td>−0.20</td>
<td>0.04</td>
</tr>
</tbody>
</table>

BMC, bone mineral content; UCP, umbilical cord plasma; FGF23, fibroblast growth factor 23; 25(OH)D, 25-hydroxyvitamin D.

In contrast, the effect of sclerostin on BMC was highly significant, also after adjustment.

In supplementary analyses, additional adjustments were performed for gestational age due to the assumption that the gestational age influenced both the BMC and the calcium metabolism. In these analyses, the independent direct effects of the UCP levels of α-klotho, FGF23, and 25(OH)D on the BMC remained nonsignificant, whereas the effect of sclerostin was still significant (data not shown).
Discussion

In this study, we demonstrated a significantly higher level of α-klotho and sclerostin and significantly lower 25(OH)D levels in the neonatal UCP compared with the maternal levels at weeks 32–34, whereas the mean FGF23 concentration was not significantly different between the babies and their mothers. Secondly, we showed that the neonatal total body BMC was significantly associated with the UCP levels of sclerostin, birth weight of the newborn, and the placental weight. However, the total body BMC could not be predicted by the maternal sclerostin values or circulating α-klotho, FGF23, or 25(OH)D levels on either the maternal or the fetal side of the placenta.

A recent study suggested that α-klotho was a biomarker for mineral metabolism in the fetus (12). The study showed that the soluble α-klotho level in cord blood was significantly elevated, in accordance with our present study, and indicated that it may have originated from syncytiotrophoblasts in the placenta. Our finding that α-klotho gene was expressed in placental tissue supports the notion of placental origin. However, it has not been determined whether the α-klotho in the fetoplacental circulation could originate directly from the fetal tissues. The lower maternal levels of α-klotho, which had been demonstrated previously (12), indicate a low production at the maternal site. However, it may also be due to a low release from the placenta to the maternal circulation or a rapid removal of the released α-klotho.

Concerning FGF23, we could not demonstrate a difference between the neonatal UCP and maternal late gestational levels. Moreover, the FGF23 gene expression was not demonstrated in the placentental tissue. Thus, our present data do not support a linkage between the plasma FGF23 concentrations and bone formation in neonates, as suggested by a recent study (10). They found that the concentration of total circulating FGF23 was high in healthy term infants, suggesting a more complex role of the FGF23 cleavage process in neonatal mineral homeostasis.

We found a significantly higher level of sclerostin in the UCP compared with the maternal circulation, and a significant effect of UCP sclerostin on neonatal total body BMC. We were not able to demonstrate sclerostin gene expression in placentental tissue. Our data indicate that sclerostin is produced in higher amounts in the newborn infant and positively related to bone mass. It could be speculated that the high sclerostin level was related to the delivery process by itself; however, the association to BMC indicates a more profound mechanism. Sclerostin is solely produced by osteocytes, the mechanoreceptors in the skeleton. Emerging evidence indicates that osteocytes regulate osteoblast function through the inhibitory protein sclerostin (13, 16, 17); however, osteocytes may also be linked to bone resorption and osteoclastogenesis (29). Sclerostin levels are increased during immobilization and decreases following physical activation (30, 31). Our findings of high UCP sclerostin levels are in accordance with the weightless status of the fetus in utero with little strain on the bones despite movements. The association of sclerostin to BMC also indicates to other mechanisms to be explored. For example, unexpected high sclerostin levels were demonstrated in patients with high-bone mass phenotype due to a constitutive activation of the Wnt signaling pathway (32), thus indicating potential regulatory interactions.

The multiple regression models presented in Table 2 were restricted to the relationships between variables that are known from the literature and also to the variables from our study. Under these prerequisites, the multiple models represented the most likely models of variables that were assumed to influence both the levels of α-klotho, FGF23, sclerostin, or 25(OH)D in the maternal circulation during late gestation and in the UCP at birth, and the infants’ BMCs. However, the presented models have shortcomings in terms of unknown or unmeasured confounders and the lack of unknown indirect mechanisms.

Fetal growth is a result of multiple factors including the genetic potential for growth, maternal nutrition, maternal metabolism, endocrine factors, and placental perfusion and function (1, 33). In addition, the ability of the fetus to respond to nutrients and other growth regulatory factors may play a role. We identified birth weight as an independent predictor of the total body BMC in newborns. Birth weight has been associated with long-term effects on health and disease in adult life (34), and several epidemiological studies have shown relationships between growth in early life and the adult bone mass. Similar work in Bath (35) and Hertfordshire, UK (36), showed relationships between the weight at 1 year of age and the adult BMC, and recent studies have found evidence for an association between growth in early life and adult bone health (37, 38).

This study demonstrated that the placental weight was strongly associated with the neonatal BMC. Placentental function is another potential major determinant of fetal growth in addition to glucose and other BMI-related factors. The placentental function includes both transport capacity and endocrine and metabolic properties (39). In principle, maternal factors may affect fetal growth through two main pathways. One pathway may be independent of the placenta, i.e. maternal nutrients (and other factors) that enter the fetal circulation directly without any interference from the placental tissues. The second pathway affects fetal growth indirectly by modifying the placental nutritional transport and metabolism. Accordingly, the placental weight has been shown to be closely correlated with the birth weight in large studies (40).

Our study has limitations. The UCP is a mixture of venous and arterial blood, which makes any conclusion about the direct sources of α-klotho. FGF23, sclerostin,
or 25(OH)D at the fetal site difficult. The time from sampling of the UCP to freezing varied due to practical limitations in a busy delivery unit. However, according to the available information, none of these proteins exhibit an instability that requires immediate cooling or freezing. Whether the blood levels of α-klotho, FGF23, sclerostin, or 25(OH)D may change as a consequence of maternal or fetal stress during delivery is unknown. Further, maternal samples obtained in weeks 30–32 might have changed during the following weeks of the last trimester.

In conclusion, we show that UCP sclerostin, placental weight, and birth weight are predictors of total bone mass in neonates. However, this study does not provide direct support for the hypothesis that maternal levels of sclerostin or the levels of α-klotho, FGF23, or 25(OH)D in the fetal or maternal circulation play a main role for bone mass in the developing skeleton.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References


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