Bone mineral density, bone markers, and fractures in adult males with congenital adrenal hyperplasia

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Abstract

Objective: The aim of this study was to determine bone mineral density (BMD), markers of bone metabolism, fractures, and steroids reflecting hormonal control in adult males with congenital adrenal hyperplasia (CAH).

Subjects, methods, and design: We compared CAH males with 21-hydroxylase deficiency (n = 30), 19–67 years old, with age- and sex-matched controls (n = 32). Subgroups of CYP21A2 genotypes, age, glucocorticoid preparation, poor control vs overtreatment, and early vs late (≥ 36 months) diagnosis were studied. BMD measured by dual energy X-ray absorptiometry and markers of bone metabolism and androgens/17-hydroxyprogesterone levels were investigated.

Results: All, including older (>30 years), CAH patients had lower BMD in all measured sites compared with control subjects. The null group demonstrated lower BMD in more locations than the other groups. Osteoporosis/osteopenia was present in 81% of CAH patients compared with 32% in controls (≥30 years). Fracture frequency was similar, osteocalcin was lower, and fewer patients than controls had vitamin D insufficiency. IGF1 was elevated in the milder genotypes. In patients, total body BMD was positively correlated to weight, BMI, total lean body mass, and triglycerides, and negatively to prolactin. Patients on prednisolone had lower BMD and osteocalcin levels than those on hydrocortisone/cortisone acetate. Patients with poor control had higher femoral neck BMD. There were no differences in BMD between patients with an early vs late diagnosis.

Conclusions: CAH males have low BMD and bone formation markers. BMD should be monitored, adequate prophylaxis and treatment established, and glucocorticoid doses optimized to minimize the risk of future fractures.

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Introduction

Congenital adrenal hyperplasia (CAH) is characterized by impaired activity of one of the enzymes needed for cortisol synthesis. The most commonly affected enzyme, 21-hydroxylase, results in cortisol and aldosterone deficiency in addition to androgen excess (1, 2). Therapy in these cases consists of lifelong glucocorticoid and often mineralocorticoid supplementation that will compensate the cortisol deficiency and normalize the elevated androgens. However, oral glucocorticoid replacement cannot accomplish normal variations in serum cortisol and the doses needed to suppress adrenal androgens are often somewhat supraphysiological. It is very well established that glucocorticoid excess in endogenous Cushing’s syndrome and pharmacological glucocorticoid therapy can lead to osteoporosis via multiple mechanisms by increasing bone resorption and decreasing bone formation. Intestinal calcium absorption may also be impaired and renal calcium excretion enhanced leading to secondary hyperparathyroidism. IGF1, their binding proteins and the secretion of gonadal steroids can also be affected (3).

However, reported data concerning bone mass and bone metabolism in CAH patients are conflicting. In children and young adults, bone mineral density (BMD) was reported to be increased (4), normal (5, 6), or decreased (7). Other pediatric studies found decreased BMD in males (8), in long-term treated girls (9), and at puberty (10). In adults, BMD has been shown to be normal (11, 12, 13, 14), or decreased (15, 16, 17, 18, 19, 20, 21, 22, 23, 24). Moreover, we have previously described increased fracture frequency compared with age- and sex-matched controls in CAH women (19). Therefore, we wanted to explore bone health in adult CAH males. Furthermore, the secretion...
of gonadal steroids is an important determinant of bone development and is very different in the two sexes.

The aim of this study was to evaluate bone health in adult men with CAH in more detail. We investigated BMD, fracture prevalence, markers of bone metabolism, and the relationship of these parameters to the therapy with adrenocortical hormones. We also studied whether there were differences in these parameters in relation to the CYP21A2 genotypes or between younger and older patients. When applicable, the results were compared with those in age- and sex-matched controls.

Materials and methods

Subjects

The study group comprised 32 adult males aged 19–67 years (≥ 40 years, n = 10; > 50 years, n = 5) with genetically confirmed CAH. They were recruited from the two participating university hospitals. Male controls, one for each patient and born on the same date as each patient, were recruited from the National Population Registry. Characteristics of this cohort have been reported earlier (25, 26).

In brief, 31 patients had 21-hydroxylase deficiency (CYP21A2 gene mutations) and one had 3β-hydroxysteroid dehydrogenase (3β-HSD) type II deficiency (C75R genotype). The phenotype was classic in 30, and 17 of them had the salt-wasting (SW) form. Two patients had the non-classic (NC) phenotype.

The patient with 3β-HSD type II deficiency and one male with CYP21A2 gene mutation and female karyotype were excluded from the statistical analyses, but are separately described in the Results section. Thus, 30 patients were included in the statistical analysis.

All subjects were divided into two age groups, < 30 years (patients n = 9, controls n = 10) and ≥ 30 years (patients n = 21, controls n = 22). Pediatric endocrinology was introduced in Sweden about 30 years before the inclusion of the present cohort, which could affect the outcome. The patient data were also divided into subgroups according to the three most prevalent CYP21A2 mutations: null, I2 splice, and I172N. Null refers to mutations completely abolishing enzyme activity and is associated with the SW phenotype. I2 splice retains a very low, but measurable, level of activity and is usually associated with SW, whereas I172N is less severe and found most often in the simple virilizing (SV) form. Effects of early (< 36 months) and late diagnosis (≥ 36 months), and of poor treatment control, arbitrarily defined as urinary pregnantriol ≥110 μmol/24 h and/or a 24-h median 17-hydroxyprogesterone (17OHP) value ≥96 nmol/l, and of overtreatment (a 24-h median 17OHP value at the level of detection i.e. ≤ 5 nmol/l) were also studied.

The study was approved by the Ethics Committees of the Karolinska Institutet, Stockholm, and the University of Gothenburg, Göteborg, Sweden. All participants gave their written informed consent.

Study protocol

Patients and controls were examined as outpatients at the Department of Endocrinology, Metabolism and Diabetes, Karolinska University Hospital, Stockholm (patients n = 22, controls n = 22) or the Department of Endocrinology, Sahlgrenska University Hospital, Göteborg (patients n = 10, controls n = 10), Sweden. Medical histories were obtained from both patients and controls. BMD was studied by dual energy X-ray absorptiometry (DXA). Blood samples were collected after an overnight fast. A morning urinary sample was collected for NTX and creatinine. In patients, a 24-h urinary pregnantriol and a diurnal 17OHP curve using dried blood spots were analyzed.

Bone mineral density

Total body, lumbar spine (L1–L4) and femoral neck BMD were estimated by DXA (Lunar Model Prodigy equipment; Lunar Radiation, Madison, WI, USA). The instruments at the two investigation sites were calibrated. Results were calculated both in g/cm², Z-score, and T-score. A T-score between −1 and −2.5 s.d. from the mean of young male adults at any measured site was defined as osteopenia, and values below −2.5 s.d. as osteoporosis.

Fractures

All types of clinical fractures which had been verified by X-ray were recorded. Fractures in vertebrae, wrist, and hip were considered to be associated with osteoporosis. The FRAX tool (www.shef.ac.uk/FRAX/) was used to evaluate fracture risk in patients and controls. This instrument provides fracture risk assessments in individuals aged 40 years or older.

Glucocorticoid supplementation

The current glucocorticoid doses were converted to hydrocortisone equivalents using anti-inflammatory equivalents (30 mg hydrocortisone = 37.5 mg cortisone acetate = 7.5 mg prednisolone = 0.75 mg dexamethasone) (27). Body surface area was calculated as the square root of (height (cm)×weight (kg)/3600 (m²)) and was used to indicate hydrocortisone equivalents in mg/m².

Biochemical assays

All assays were performed on serum unless otherwise stated. Testosterone, estradiol and dried blood spot 17OHP (measured at 0800, 1400, 1900, 0100, and
0600 h), and 24-h urinary pregnanetriol were measured as described previously (25, 26).

DHEAS and PTH were measured on an Advantage automatic immune analyzer (Nichols Institute Diagnostics, San Clemente, CA, USA); the reference limits of PTH were 12–55 ng/l. RIA methods were used for the determination of androstenedione (DiaSorin S.p.A., Saluggia, Italy) and IGF1 (28). IGF1 levels expressed as SDS were calculated from values in 448 healthy subjects (29). Carboxy-terminal cross-linked telopeptide of type I collagen (CTX) was measured on a Roche Elecsys 1010/2010 immunoassay analyzer (Roche Diagnostics Ltd.); reference limits were for males < 50 years < 580 ng/l and 50–70 years < 700 ng/l. Bone-specific ALP (BALP) was measured with the Access Ostase assay with reference limit in males < 20 μg/l, and for prolactin a chemiluminescent immunoassay (Beckman Coulter, Fullerton, CA, USA) was used. Osteocalcin was determined by an IRMA (CIS Bio International, Gif-sur-Yvette, France); reference limits were for males 18–29 years < 70 μg/l, 30–49 years < 45 μg/l, and ≥ 50 years < 50 μg/l. A chemiluminescent immunoassay (DiaSorin S.p.A.) was used for analyzing 25-hydroxy-vitamin D (25OHD); levels < 25 nmol/l indicated deficiency, 25–74 nmol/l insufficiency, and optimal levels 75–250 nmol/l (30).

Urinary amino-terminal collagen crosslinks (NTX) was measured with enzyme immunoassay Osteomark (Inverness Medical, Princeton, NJ, USA); reference limits were 3–51 nmol/l bone collagen equivalents/ mmol creatinine. Ionized calcium, phosphate, albumin, creatinine, and thyroid function tests were determined by standardized and certified procedures. The within and between assay coefficients of variation were PTH, 6.7 and 8.7%; DHEAS, 4.4 and 8.7%; IGF1, 4 and 11%; CTX, 2 and 17.9%; BALP, 4.2 and 10%; osteocalcin, 3.5 and 4.2%; 25OHD, 2.9 and 7.9%; and NTX, 4 and 10%.

**Statistical analysis**

Results are presented as the mean ± S.D. unless otherwise stated. Comparisons between two groups were made using the unpaired t-test when values were normally distributed. Otherwise, the Mann–Whitney rank-sum test was used and in these cases the median and range are reported. When continuous variables were compared in three groups, one-way ANOVA was used for normal distributions, otherwise the Kruskal–Wallis test was performed. χ² was used in frequency table calculations or, when the expected frequency was small (< 5) Fisher’s exact test was used. All proportions were calculated discounting missing values. Correlations between variables were assessed using linear and multiple regression analysis. Statistical significance was set at *P* < 0.05 and tendency at 0.05–0.10. Data were analyzed using SigmaStat for Windows (Systat Software, Inc., San Jose, CA, USA).

**Results**

**Bone mineral density**

All patients (including the older cohort) with CAH had lower BMD at all measured sites compared with controls (Table 1 and Fig. 1). Older patients had lower BMD in femoral neck compared with younger patients, whereas there were no differences between younger and older control subjects. Osteoporosis or osteopenia was present in 67% of all CAH patients compared with 39% of all controls (*P* = 0.054), and in males ≥ 30 years the corresponding figures were 81% compared with 32% (*P* = 0.003). Compared with controls, total body BMD was significantly lowered in the null and I2 splice genotype groups and showed a similar trend in the I172N group (Table 1). In the null genotype group femoral neck BMD was also decreased (Table 1). Nine patients were on calcium and vitamin D supplementation for osteoporosis or osteopenia compared with none among the controls (*P* < 0.001). Two of these patients, aged 41 and 67 years, had in addition bisphosphonate therapy with alendronate.

**Fractures**

The numbers of individuals with fractures (total or associated with osteoporosis) were similar in the entire group and in all subgroups compared with their respective controls (Table 1). No difference between the younger and older groups was found (data not shown). Only one patient > 50 years had fractures (clavicle and finger). The I172N group had more individuals with a fracture than the null group (Table 1). When the total number of fractures was compared the result was similar between patients and controls (data not shown). The fractures had often occurred during sport activities, but the underlying trauma was not recorded systematically. Calculation of fracture risk indicated that the 10 years, probability of a major osteoporotic fracture was 8.1 ± 4.0% in CAH males compared with 4.9 ± 3.0% in controls (*P* = 0.058), and a hip fracture was 2.2 ± 2.1% in CAH males compared with 0.8 ± 0.7% in controls (*P* = 0.050). However, if the sole CAH male patient who did not use glucocorticoids was excluded, FRAX evaluation of risk factor increased slightly to 8.6 ± 3.8 and 2.4 ± 2.1% (*P* = 0.025 and 0.029 respectively compared with controls).

A 23-year-old patient (I2 splice, lumbar spine T-score − 1.0 s.d. otherwise normal BMD, 5 mg prednisolone daily, poor control) had 21 individual fractures all obtained during sports activities such as boxing and football. A 41-year-old patient (I2 splice, T-score − 3.5 s.d. in total body, 6.25 mg prednisolone daily, good control, serum testosterone 18 nmol/l, lactose intolerant) had a spontaneous vertebral fracture, and used alendronate in addition to calcium and vitamin D supplementation.
Table 1  BMD, frequency of osteopenia/osteoporosis and fractures and biochemical tests in adult males with 21-hydroxylase deficiency, also divided into the three most common CYP21A2 genotype groups, and male controls (mean± s.d. or median and range).

<table>
<thead>
<tr>
<th></th>
<th>Patients (n=30)</th>
<th>Controls (n=32)</th>
<th>P value vs controls</th>
<th>Null (n=7)</th>
<th>P value vs controls</th>
<th>I2 splice (n=11)</th>
<th>P value vs controls</th>
<th>I172N (n=9)</th>
<th>P value vs controls</th>
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</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>35.7±11.4</td>
<td>36.5±11.9</td>
<td>NS</td>
<td>34.5±9.5</td>
<td>NS</td>
<td>31.6±9.4</td>
<td>NS</td>
<td>41.0±14.9</td>
<td>NS</td>
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<td>Total body BMD (g/cm²)</td>
<td>1.17±0.11</td>
<td>1.27±0.10</td>
<td>&lt;0.001</td>
<td>1.16±0.11</td>
<td>0.011</td>
<td>1.17±0.14</td>
<td>0.018</td>
<td>1.20±0.07</td>
<td>NS (0.067)</td>
</tr>
<tr>
<td>Total BMC (g)</td>
<td>2.69±0.42</td>
<td>3.39±0.50</td>
<td>&lt;0.001</td>
<td>2.59±0.51</td>
<td>&lt;0.001</td>
<td>2.75±0.52</td>
<td>&lt;0.001</td>
<td>2.70±0.19</td>
<td>&lt;0.001</td>
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<td>Lumbar BMD (g/cm²)</td>
<td>1.16±0.20</td>
<td>1.23±0.16</td>
<td>0.041</td>
<td>1.19±0.17</td>
<td>NS</td>
<td>1.13±0.23</td>
<td>NS</td>
<td>1.20±0.21</td>
<td>NS</td>
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<td>Femoral neck BMD (g/cm²)</td>
<td>0.96±0.13</td>
<td>1.05±0.14</td>
<td>0.013</td>
<td>0.93±0.10</td>
<td>0.040</td>
<td>0.99±0.18</td>
<td>NS</td>
<td>0.99±0.07</td>
<td>NS</td>
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<td>Osteoporosis, n</td>
<td>4/30</td>
<td>1/31</td>
<td>NS</td>
<td>7/32</td>
<td>NS</td>
<td>2/11</td>
<td>NS</td>
<td>0/9</td>
<td>NS</td>
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<td>Osteoporosis/osteopenia, n</td>
<td>20/30</td>
<td>12/31</td>
<td>NS</td>
<td>47/49</td>
<td>NS</td>
<td>8/11</td>
<td>NS</td>
<td>69/69</td>
<td>NS</td>
</tr>
<tr>
<td>Individuals with fractures, n</td>
<td>16/30</td>
<td>14/32</td>
<td>NS</td>
<td>17/32</td>
<td>NS</td>
<td>6/11</td>
<td>NS</td>
<td>3/11</td>
<td>NS</td>
</tr>
<tr>
<td>Osteoporotic, n</td>
<td>6/30</td>
<td>5/32</td>
<td>NS</td>
<td>7/32</td>
<td>NS</td>
<td>3/11</td>
<td>NS</td>
<td>3/9</td>
<td>NS</td>
</tr>
<tr>
<td>CaD supplementation</td>
<td>9/30</td>
<td>0/30</td>
<td>&lt;0.001</td>
<td>2/7</td>
<td>0.021</td>
<td>5/11</td>
<td>&lt;0.001</td>
<td>2/9</td>
<td>0.044</td>
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<tr>
<td>S-25OH-vit D (nmol/l)*</td>
<td>49±22</td>
<td>45±24</td>
<td>NS</td>
<td>46±16</td>
<td>NS</td>
<td>45±24</td>
<td>NS</td>
<td>56±34</td>
<td>NS</td>
</tr>
<tr>
<td>S-crea (µmol/l)</td>
<td>75.1±11.0</td>
<td>82.8±10.9</td>
<td>0.008</td>
<td>72.6±10.6</td>
<td>0.031</td>
<td>76.2±11.8</td>
<td>NS</td>
<td>73.3±9.5</td>
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<td>S-cystatin C (mg/l)</td>
<td>0.84±0.14</td>
<td>0.85±0.17</td>
<td>NS</td>
<td>0.85±0.17</td>
<td>NS</td>
<td>0.87±0.16</td>
<td>NS</td>
<td>0.79±0.08</td>
<td>NS</td>
</tr>
<tr>
<td>S-BALP (µg/l)</td>
<td>10.3±3.5</td>
<td>12.7±6.9</td>
<td>NS (0.088)</td>
<td>9.2±3.3</td>
<td>NS (0.070)</td>
<td>9.9±2.9</td>
<td>NS</td>
<td>10.6±4.8</td>
<td>NS</td>
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<tr>
<td>S-osteocalcin (µg/l)</td>
<td>18.6±7.5</td>
<td>25.6±8.5</td>
<td>&lt;0.001</td>
<td>14.3±4.1</td>
<td>&lt;0.001</td>
<td>20.3±4.5</td>
<td>0.012</td>
<td>18.5±10.3</td>
<td>0.027</td>
</tr>
<tr>
<td>S-CTX (ng/l)</td>
<td>445±144</td>
<td>455±247</td>
<td>NS</td>
<td>396±117</td>
<td>NS</td>
<td>490±134</td>
<td>NS</td>
<td>371±178</td>
<td>NS</td>
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<tr>
<td>U-NTX/cr (nmol/mmol)</td>
<td>53±21</td>
<td>51±34</td>
<td>NS</td>
<td>47±23</td>
<td>NS</td>
<td>61±18</td>
<td>NS</td>
<td>43±15</td>
<td>NS</td>
</tr>
<tr>
<td>S-prolactin (µg/l)</td>
<td>11.4±5.4</td>
<td>9.9±3.1</td>
<td>NS</td>
<td>10.1±4.4</td>
<td>NS</td>
<td>13.1±6.8</td>
<td>0.044</td>
<td>9.8±4.5</td>
<td>NS</td>
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<tr>
<td>S-IGF1 (µg/l)</td>
<td>272±101</td>
<td>237±45</td>
<td>NS (0.084)</td>
<td>269±98</td>
<td>NS</td>
<td>293±132</td>
<td>0.045</td>
<td>262±58</td>
<td>NS</td>
</tr>
<tr>
<td>S-IGF1 (SDS)</td>
<td>0.80 (−3.60)</td>
<td>0.40 (−0.8)</td>
<td>NS (0.060)</td>
<td>1.1 (−3.6)</td>
<td>NS (0.089)</td>
<td>0.59±1.08</td>
<td>NS</td>
<td>1.00±0.96</td>
<td>0.012</td>
</tr>
</tbody>
</table>

NS, non significant; Lumbar, lumbar spine; F, free; CaD, calcium and vitamin D. *P<0.05 compared with the other genotypes; †post hoc null vs I172N, P=0.041. *Did not differ significantly if those on CaD supplementation were excluded.
Early vs late diagnosis

There were no differences between the early (<36 months, n=21) and late diagnosis cohorts (≥36 months, n=9) in BMD (total body, lumbar spine, and femoral neck), frequency of osteoporosis/osteopenia, and fracture frequency, even though all osteoporotic fractures were in the early-diagnosed group (data not shown).

Markers for bone metabolism and other biochemical tests

Results of biochemical tests are shown in Table 1.

Bone formation markers

Serum osteocalcin levels were lower in all cohorts of patients compared with the control group, whereas serum BALP levels tended to be decreased in all patients and in the null genotype group. All values were within normal range except for two subjects in the control group having elevated BALP levels.

Bone resorption markers

Urinary NTX tended to be increased in the I2 splice group. Eleven patients and five subjects in the control group had values above the upper reference limit. Serum CTX was not different in all comparisons between patients and control subjects, but was increased in both younger patients and younger control subjects compared with older patients and older control subjects. Seven patients and eight control subjects had concentrations above the upper reference range.

Calcium homeostasis and renal function

Ionized calcium and albumin levels tended (P=0.088 and 0.060 respectively) to be lower in patients compared with control subjects (data not shown). Three patients and one control subject had modestly elevated PTH levels. Only three patients and three control subjects had optimal vitamin D values (>75 nmol/l). One of these patients was on calcium and vitamin D supple- mentation. Fewer CAH males had vitamin D deficiency (<25 nmol/l) compared with control subjects (3/30 vs 10/28, P=0.042). There was no obvious difference between patient and control groups regarding the time of year for blood sample collection. Creatinine was lower in all groups of patients compared with controls (except in younger CAH males and in I2 splice it was only a tendency), but serum cystatin C was similar between all groups. PTH and phosphate did not differ between the groups (data not shown).

Nonsteroid hormone determinations

IGF1 was elevated in the I2 splice group (μg/l) and in the I172N group (SDS) compared with controls. Prolactin was elevated in the I2 splice group compared with the control group. No differences were found in thyroid function tests except for a slight increase in free T3 in the null group (data not shown).

Androgen levels

Although in many patients androstenedione levels were either higher or lower than in the control subjects, the average level was not different between the two groups. In contrast, in almost all patients DHEAS levels were very low compared with control subjects.
Overall, serum testosterone levels in the patients tended to be lower than in the control subjects (14.6 ± 5.4 vs 17.0 ± 4.9 nmol/l, \( P = 0.068 \)). The difference was significant in the older group (13.1 ± 5.1 vs 16.2 ± 4.8 nmol/l, \( P = 0.045 \)) but not in the younger group (16.8 ± 3.9 vs 18.7 ± 5.0 nmol/l, \( P = \text{NS} \)) (partly presented in reference (25)).

**Corticosteroid therapy and control of treatment**

Glucocorticoids were taken by 93% (28/30) of the patients (25, 26). The most frequently used preparations were prednisolone (62%, 18/29) and hydrocortisone (17%, 5/29). The mean dose in hydrocortisone equivalents was 17.4 ± 5.2 mg/m² (32.7 ± 12.4 mg) without differences between younger and older patients or genotype groups. Most patients (87%, 26/30) received fludrocortisone with a mean dose of 0.11 ± 0.06 mg, with similar doses in the different groups. Decreased BMD and osteocalcin levels compared with control subjects were present in the group of patients treated with prednisolone but not in patients on short-acting glucocorticoids in spite of similar doses in hydrocortisone equivalents. Increased serum IGF1 level was only present in those on short-acting glucocorticoids (Table 2). Androgen levels were not different.

The 17OHP curves varied considerably between the individual patients (Fig. 3). Forty-four percent (7/16) of prednisolone-treated patients had no diurnal variations compared with 14% (1/7) of those on hydrocortisone/cortisone acetate (\( P = \text{NS} \)). The one patient on dexamethasone also lacked diurnal variations. The area under the curve for 17OHP was 1995 nmol/l h (630–22 650) in patients on prednisolone, 2025 nmol/l h (600–16 470) for those on hydrocortisone/cortisone acetate, and 690 nmol/l h in one patient on dexamethasone. There were no significant differences between the three genotype groups (not shown).

**Poor control vs overtreatment**

Patients with poor control (\( n = 7 \)) had significantly higher femoral neck BMD and a tendency to decreased urinary NTX/creatinine compared with patients having suppressed 17OHP and/or urinary pregnanetriol (\( n = 10 \)) (\( T \)-score \(-0.70 \pm 0.53 \) vs \(-1.56 \pm 0.75 \) SDS, \( P = 0.025 \); \( 38 \pm 23 \) vs \( 61 \pm 17 \) nmol/mmol, \( P = 0.056 \)). BMD at other sites did not differ between those with poor control and those with overtreated (total body \( T \)-score \(-0.74 \pm 0.50 \) vs \(-1.44 \pm 1.03 \) SDS; lumbar spine \( T \)-score \(-0.74 \pm 0.47 \) vs \(-0.49 \pm 2.44 \) SDS; osteoporosis/osteopenia 5/7 vs 6/10; fractures 4/7 vs 6/10). The results were similar if BMD (g/cm²) or Z-score were used (data not shown).

**SW vs SV phenotype**

No differences between SW and SV were found in the parameters reported above (data not shown) apart from a tendency to higher urinary NTX in SW compared with SV (58 ± 18 vs 39 ± 16 nmol/l, \( P = 0.062 \)).
Table 2 BMD, frequency of osteopenia/osteoporosis, fractures and biochemical tests in adult males with 21-hydroxylase deficiency on different glucocorticoids and male controls (mean±s.d. or median and range).

<table>
<thead>
<tr>
<th></th>
<th>Prednisolone</th>
<th></th>
<th>HC or CoAc</th>
<th></th>
<th>Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(n=18)</td>
<td>P value</td>
<td>(n=8)</td>
<td>P value</td>
<td>(n=32)</td>
</tr>
<tr>
<td>Age (years)</td>
<td>35.6±12.0</td>
<td>NS</td>
<td>37.4±12.8</td>
<td>NS</td>
<td>36.5±11.9</td>
</tr>
<tr>
<td>Total body BMD (g/cm²)</td>
<td>1.13±0.10</td>
<td>&lt;0.001</td>
<td>1.25±0.11*</td>
<td>NS</td>
<td>1.27±0.10</td>
</tr>
<tr>
<td>Total body (T-score)</td>
<td>−1.25±1.71</td>
<td>&lt;0.001</td>
<td>−0.14±1.22*</td>
<td>NS</td>
<td>0.53±1.15</td>
</tr>
<tr>
<td>Total BMC (g)</td>
<td>2.54±0.49</td>
<td>&lt;0.001</td>
<td>2.88±0.33†</td>
<td>0.009</td>
<td>3.39±0.50</td>
</tr>
<tr>
<td>Lumbar BMC (g/cm²)</td>
<td>1.12±0.18</td>
<td>0.031</td>
<td>1.22±0.24</td>
<td>NS</td>
<td>1.23±0.16</td>
</tr>
<tr>
<td>Lumbar (T-score)</td>
<td>−0.94±1.44</td>
<td>0.026</td>
<td>−0.30±2.14</td>
<td>NS</td>
<td>−0.01±1.33</td>
</tr>
<tr>
<td>Femoral neck BMD (g/cm²)</td>
<td>0.94±0.12</td>
<td>0.006</td>
<td>0.99±0.17</td>
<td>NS</td>
<td>1.05±0.14</td>
</tr>
<tr>
<td>Femoral neck (T-score)</td>
<td>−1.22±0.84</td>
<td>0.002</td>
<td>−0.83±1.38</td>
<td>NS</td>
<td>−0.21±1.10</td>
</tr>
<tr>
<td>Osteoporosis</td>
<td>4/18</td>
<td>0.047</td>
<td>0/8</td>
<td>NS</td>
<td>1/31</td>
</tr>
<tr>
<td>Osteoporosis/osteopenia</td>
<td>12/18</td>
<td>(0.069)</td>
<td>6/8</td>
<td>NS</td>
<td>12/31</td>
</tr>
<tr>
<td>Individuals with fractures</td>
<td>9/18</td>
<td>NS</td>
<td>4/8</td>
<td>NS</td>
<td>14/32</td>
</tr>
<tr>
<td>Osteoporotic</td>
<td>4/18</td>
<td>NS</td>
<td>1/8</td>
<td>NS</td>
<td>5/32</td>
</tr>
<tr>
<td>CaD supplementation</td>
<td>7/18</td>
<td>0.001</td>
<td>2/9</td>
<td>0.036</td>
<td>0/32</td>
</tr>
<tr>
<td>S-25OH-vit D</td>
<td>47±16</td>
<td>NS</td>
<td>55±31</td>
<td>NS</td>
<td>45±24</td>
</tr>
<tr>
<td>S-crea (µmol/l)</td>
<td>73.1±10.2</td>
<td>0.004</td>
<td>79.3±11.9</td>
<td>NS</td>
<td>82.8±10.9</td>
</tr>
<tr>
<td>S-ostecalcin (µg/l)</td>
<td>16.6±4.6</td>
<td>&lt;0.001</td>
<td>19.5±9.1</td>
<td>NS (0.058)</td>
<td>25.6±8.5</td>
</tr>
<tr>
<td>S-CTX (ng/l)</td>
<td>426±129</td>
<td>NS</td>
<td>461±173</td>
<td>NS</td>
<td>455±247</td>
</tr>
<tr>
<td>U-NTX/crea</td>
<td>52±21</td>
<td>NS</td>
<td>56±14</td>
<td>NS</td>
<td>51±34</td>
</tr>
<tr>
<td>S-estradiol (pg/ml)</td>
<td>91 (26–298)</td>
<td>NS</td>
<td>79 (49–169)</td>
<td>NS</td>
<td>75 (43–148)</td>
</tr>
<tr>
<td>S-prolactin (µg/l)</td>
<td>12.1±5.8</td>
<td>(0.091)</td>
<td>10.7±4.9</td>
<td>NS</td>
<td>9.9±3.1</td>
</tr>
<tr>
<td>S-IGF1 (µg/l)</td>
<td>237±72</td>
<td>NS</td>
<td>305±41*</td>
<td>&lt;0.001</td>
<td>237±45</td>
</tr>
<tr>
<td>S-IGF1 (SDS)</td>
<td>0.17±1.20</td>
<td>NS</td>
<td>1.29±1.01*</td>
<td>&lt;0.001</td>
<td>0.34±0.54</td>
</tr>
<tr>
<td>HCeq/m² (mg/m²)</td>
<td>16.8±4.6</td>
<td>NS</td>
<td>19.9±6.3</td>
<td>NS</td>
<td>0.10</td>
</tr>
<tr>
<td>Months on GC</td>
<td>423 (231–635)</td>
<td>NS</td>
<td>391 (284–470)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AUC 17OHP (nmol/l×h)</td>
<td>1995 (630–22 650)</td>
<td></td>
<td>2025 (600–16 470)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*P<0.05, †P=0.05–0.099 compared with the Prednisolone group. NS, not significant; HC, hydrocortisone; CoAC, cortisone acetate; Lumbar, lumbar spine; HCeq, hydrocortisone equivalents; GC, glucocorticoid; AUC, area under the curve.

Relationships between BMD and other parameters

BMD was correlated with our previously reported data on anthropometric parameters, body composition, metabolic profile, testicular ultrasound, and sperm analysis (25, 26). In patients, total body BMD was positively correlated to weight (r=0.567, P=0.001), BMI (r=0.448, P=0.013), total lean body mass (r=0.665, P<0.001), triglycerides (r=0.412, P=0.024), and negatively to prolactin (r=−0.412, P=0.012), and total sperm count and concentration (r=−0.544 and −0.533; P=0.044 and 0.049 respectively). No correlations were found between BMD and dose of hydrocortisone equivalents/m² or other biochemical markers. In multiple linear regression, the strongest correlation with total body BMD was total lean mass and prolactin (r=0.746, P<0.001).

In controls, total body BMD was positively correlated to weight (r=0.706, P<0.001), BMI (r=0.556, P=0.001), waist (r=0.433, P=0.027), hip (r=0.650, P<0.001), total fat and lean body mass (r=0.414, P=0.023 and r=0.675, P<0.001 respectively), osteocalcin (r=0.412, P=0.024), calcium (r=0.448, P=0.001), and testosterone/estradiol ratio (r=0.414, P=0.026). In multiple linear regression, the strongest correlation with total body BMD was weight and osteocalcin (r=0.797, P<0.001).

Total body BMD was positively correlated to lumbar spine and femoral neck BMD: CAH males r=0.434, P=0.017 and r=0.716, P<0.001; controls r=0.841 and r=0.741, both P<0.001. In patients, lumbar spine BMD was also correlated positively with fludrocortisone dose (r=0.382, P=0.037) and negatively with total functional testicular volume (r=−0.434, P=0.019).

Cases

A 61-year-old male with karyotype 46,XX (12 splice) was diagnosed and received glucocorticoids at age 7, and initiated testosterone replacement at 12 years. Current doses were prednisolone 6.25 mg daily and testosterone 250 mg i.m. every 3rd week. DXA measurement using the male reference range suggested good BMD (T-score lumbar spine 0.6 and femoral neck −0.1 SDS). His 17OHP levels indicated over-substitution.

The 22-year-old patient, with 3β-HSD type II deficiency, used prednisolone 7.5 mg daily and had good BMD (T-score lumbar spine 0.7 and femoral neck 1.3 SDS). His DHEAS level was high. None of them reported any fractures.

Discussion

This is a comprehensive study reporting on BMD and markers of bone metabolism in adult CAH males. We found reduced BMD and almost threefold increase in osteopenia/osteoporosis in patients older than 30 years.
compared with age-matched controls. Biochemical tests indicated suppressed bone formation and a tendency to increased bone resorption. There was no evidence of vitamin D deficiency. The frequency of fractures was not increased but FRAX evaluation indicated increased fracture risk in CAH males on glucocorticoid supplementation. Bone health appeared less affected in patients taking short-acting glucocorticoids than in those taking more long-acting preparations.

Among our CAH male patients, 67% had osteoporosis/osteopenia according to the WHO criteria. Only a few studies report BMD in adult CAH individuals with the results for men and women demonstrated separately (13, 14, 17, 22, 23). Our finding of a low BMD is in agreement with the results in some (17, 22, 23), but not all (13, 14), previous studies. In a recent report from Norway, 23 males were included who had a similar age and glucocorticoid doses as our cohort and the frequency of osteopenia/osteoporosis was 57% (23). Compared with controls, our patients had decreased bone formation markers and similar resorption markers indicating a negative balance with risk for further decrease of BMD. Instead, Sciannamblo et al. (17) found a higher bone turnover than in controls, studying a cohort of CAH males where some patients were still growing, while a smaller CAH study with extreme heterogeneity demonstrated decrease of both formation and resorption markers (11).

In spite of the difference in BMD between our patients and control subjects, the frequency of fractures did not differ. Regrettably, we did not ask the source of the trauma that had caused the fractures, and the proportions of low-energy fractures in patients and control subjects are unknown. The patients, however, often spontaneously informed us that the fractures had occurred during sports activities. The only previous report on fractures is from Koetz et al. (22) who studied bone health in primary adrenocortical insufficiency and found only one fracture in 14 CAH males. However, BMD decreases by age in the general population and is associated with increasing fracture risk, and the peak incidence of osteoporotic fractures occurs in old age in males. In our patients, the femoral neck BMD was lower in older than in younger patients. Assessment of fracture risk in our patients of age 40 years and older using the FRAX tool indicated an increased fracture risk. Moreover, it can be assumed that FRAX underestimates the fracture risk as it has not been developed for life-long glucocorticoid supplementation but only calculates glucocorticoid use for more than 3 months as a risk factor. It is of concern that a 41-year-old CAH male with osteoporosis had obtained a spontaneous vertebral fracture. The patient was, however, lactose intolerant which might have contributed to the poor bone health.

Fractures were more prevalent in the milder genotype group I172N compared with the null group despite the fact that BMD at all sites and the mean present glucocorticoid and mineralocorticoid doses were similar (25). A possible explanation for the increased fracture frequency in the I172N group might be that the doses of corticosteroids were too high considering the milder disease. We have previously found that the I172N group

Figure 3 Diurnal 17-hydroxyprogesterone (17OHP) curve analyzed from dried blood spots in adult males with 21-hydroxylase deficiency. Upper panel shows prednisolone supplementation, middle panel shows hydrocortisone/cortisone acetate supplementation, and lower panel shows patients with no regular glucocorticoid supplementation. The 17OHP curve of the male with dexamethasone supplementation is not illustrated; all values were at or less than the level of detection (5 nmol/l).
also had a more unfavorable cardiovascular/metabolic profile (25).

A positive finding was that younger CAH males (<30 years) had similar BMD to the control subjects. Hopefully, this means that younger CAH patients have a lower risk of future fractures. The reason for the normal BMD may be improved care during childhood resulting in increased peak bone mass.

An adequate supply of vitamin D is essential for bone health. Studies on 25OHD levels in CAH patients are rare and have shown subnormal values (22, 23), while in this study 25OHD did not differ from age- and sex-matched controls. This is in accordance with a recent report showing values in adult classic CAH similar to the overall population (24). Prescribing vitamin D should be considered in CAH patients, especially in countries with high risk for low 25OHD and osteoporosis, such as Sweden.

As described previously (19, 22), we found no relationship between BMD and current glucocorticoid dose. However, correlations between BMD and cumulative glucocorticoid doses have been demonstrated previously (16, 31). We found in multiple regression analysis that the strongest correlation with BMD was with lean body mass and prolactin. Prolactin levels have shown to be negatively correlated to BMD in males and in about half of the studies there was no correlation with testosterone levels (32). Prolactin may possibly affect BMD by inducing some degree of hypogonadism but may also have a direct effect on bone.

The most important issue for bone health in the male patients with CAH is to optimize the glucocorticoid therapy. The ideal is to establish normal diurnal cortisol values and androgen concentrations. In this study, a number of patients had suppressed diurnal 17OHP curves without diurnal variation as well as variable reduction of androgen levels indicating that the glucocorticoid doses could be reduced. However, some patients demonstrated very high levels of DHEAS, androstenedione, and diurnal 17OHP. These patients with poor control had better BMD, probably due to long-term high-androgen concentrations and lower exposure to glucocorticoids. We also report herein BMD levels in a patient with 3β-HSD type II deficiency for the first time. He had a good BMD and the high levels of DHEAS most likely contributed to the favorable outcome. We for the first time also studied BMD in a CAH male with karyotype 46,XX. He had normal values according to the male reference population despite modest overtreatment with glucocorticoids, probably due to regular testosterone replacement since puberty.

The glucocorticoid preparations used varied. Hydrocortisone is the preferred glucocorticoid in children while prednisolone is the preferred option in many adults (2). All our patients with osteoporosis (aged 24–52 years) and the patient with the most serious fracture, a spontaneous vertebrae fracture, used prednisolone. This is in accordance with Jaaskelainen et al. (15), who reported that adult CAH patients substituted with hydrocortisone had better BMD than those on prednisone, prednisolone, or dexamethasone, and the recent study by Koetz et al. (22) describing that hydrocortisone-treated patients with primary adrenal insufficiency had better BMD compared with prednisolone-treated patients. A number of circumstances may contribute to a more negative effect on bone health with prednisolone than with short-acting glucocorticoids. Prednisolone-treated patients received a dose in the evening to suppress the early morning rise of ACTH, 17OHP, and androgens. This may result in a supraphysiological glucocorticoid exposure to the skeleton during late night and early morning hours with a negative impact on BMD. About 40% of our patients on prednisolone lacked diurnal variations of 17OHP.

Moreover, in our cohort, we have previously found that fat mass was increased compared with controls in hydrocortisone/cortisone acetate-treated patients but not in prednisolone treated (25). It is generally accepted that an increased fat mass is beneficial for the skeleton (33).

IGF1 is a growth factor important for bone tissue. Hydrocortisone/cortisone acetate-treated patients had higher IGF1 SDS compared with controls. This difference was not found in prednisolone-treated patients. It can be speculated that the increased IGF1 might be advantageous. We also found elevated IGF1 levels in adult CAH females (19). It has also been reported in endogenous Cushing’s syndrome (34), and in pharmacological glucocorticoid therapy (35, 36). Glucocorticoids probably stimulate IGF1 and the extent may vary with glucocorticoid dose and preparation. There is certainly a need for further studies on which glucocorticoid to choose in adult CAH patients for optimal balance between beneficial effects and adverse effects.

The outcome with regard to bone health in CAH males differs from our previous results in female CAH patients. Although the reduction in bone density and the frequency of osteopenia/osteoporosis were similar in males and females, the number of fractures differed. As expected, both CAH males and their controls had more fractures than their female counterparts in line with the higher prevalence of fractures in males than in females before the age of 50–55 years (37, 38). The difference has been attributed to dissimilarities in lifestyle, with males in general having more risky behavior/more participation in rough sports compared with females. However, the fracture frequency was almost ten times increased in CAH females compared with their controls (30 vs 3%), whereas there was no difference between CAH males and controls (53 vs 44%). Although many women with CAH are involved in rough sports (39), it is unlikely to be the only explanation for this outcome. Other circumstances can also be important for the gender difference, for instance compliance with treatment and the dissimilarities in sexual hormone.
concentrations. Moreover, the sensitivity of the glucocorticoid receptors to endogenous and exogenous glucocorticoids is highly individual (40).

The Endocrine Society clinical practice guidelines recommend against the routine evaluation of BMD in children and do not recommend regular monitoring of BMD in adult CAH patients (41). Our findings of low BMD in CAH males and previously in CAH females emphasize the importance of monitoring BMD in at least adult patients starting from young adult age. We have recently recommended DXA measurements of BMD to be done when the patient is transferred from pediatric to adult care and, depending on the BMD levels and the patient’s individual risk, repeated every second to fifth year (2). Osteoporosis prophylaxis with recommended physical activities, appropriate nutrition, calcium and vitamin D supplementation, and bisphophonates may be considered. The primary option is, however, optimizing the glucocorticoid supplementation.

The major limitation of this study is its sample size despite being larger than most and including older CAH males and age- and sex-matched controls in contrast to most other studies. Negative findings must therefore be interpreted with caution. Another limitation is assessing the impact of steroid treatment. Neither type of steroids previously used nor their cumulative lifetime doses were available and we used the dose of the present steroid in the calculations. Steroid excess at an early age may certainly have a continuing negative impact on bone health in adult age.

In conclusion, adult CAH males with 21-hydroxylase deficiency have decreased BMD and bone formation markers compared with age- and sex-matched controls. Genotype and age at diagnosis did not affect these values. Fractures, bone resorption markers, and vitamin D did not differ between groups. However, overtreatment with glucocorticoids and prednisolone treatment appeared to affect BMD more negatively than poor control and short-acting glucocorticoids.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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