Des-acyl ghrelin (DAG) is one of the three preproghrelin gene-encoded peptides. Compared with ghrelin and obestatin, it has not received the attention it deserves. DAG has long been considered an inert degradation product of acyl ghrelin (AG). Recent evidence, however, indicates that DAG behaves like a separate hormone. DAG can act together with AG, can antagonize AG, and seems to have AG-independent effects. Therefore, it is believed that DAG must activate its own receptor and that it may also interact with AG at this receptor. Of potential clinical importance is that an increasing number of studies suggest that DAG might be a functional inhibitor of ghrelin and that DAG can suppress ghrelin levels in humans. Therefore, DAG or DAG analogs might be good candidates for future treatment of metabolic disorders or other conditions in which antagonism of AG actions could be beneficial, such as diabetes, obesity, and Prader–Willi syndrome.

Introduction

It is now more than 12 years since ghrelin was identified as the ligand of the GH secretagogue receptor type 1a (GHSR1a). The story of des-acyl ghrelin (DAG; Fig. 1) also began in 1999, when Kojima et al. described this peptide in the same report that introduced ghrelin to the scientific world (1, 2). The preproghrelin gene-derived peptides include acyl ghrelin (AG), DAG, and obestatin. AG is produced mainly by the stomach and exerts its central and peripheral effects through GHSR1a (1). The ghrelin peptide is acylated by the enzyme ghrelin O-acyl transferase (GOAT) (3, 4, 5), which is expressed predominantly in the stomach, gut, and pancreas but also at other sites (6).

The concept that emerged from Kojima’s paper and many others was that DAG should be considered as the degradation product of ghrelin without biological activities. The obvious reason for this is that DAG is unable to bind and activate the GHSR1a in the sub-micromolar, physiological range (1, 7). Therefore, DAG has never received the level of attention in research and the literature that it deserves, although a growing number of reports strongly suggest that DAG has intrinsic activities in a variety of physiological and pathophysiological situations. Depending on the experimental circumstance, DAG can either antagonize or support the activities of ghrelin, while in other situations, DAG works completely independently of ghrelin.

In this review, we will try to address some of these actions of DAG on (patho)physiological processes. We describe the properties of DAG not only as a functional antagonist of AG but also as a strong inhibitor of AG serum levels in vivo in animals and men. Finally, we will describe the situations in which DAG acts independently of AG.

The controversial aspects of the AG/DAG ratio

Although available data are still contradictory, several studies have indicated that obese mice and humans might have lower DAG levels than normal-weight subjects whereas AG levels are similar, indicating that obesity could reflect a relative DAG deficiency and that DAG levels might be regulated by body weight (8, 9, 10). Moreover, insulin-resistant obese subjects have an elevated AG/DAG ratio when compared with insulin-sensitive obese subjects (11, 12). St-Pierre et al. (9) found that the dysregulation of ghrelin secretion profiles during a euglycemic/hyperinsulinemic clamp (EHC) is associated with insulin resistance (IR). They determined whether insulin-sensitive overweight or obese (ISO) and insulin-resistant overweight or obese (IRO) individuals display different AG and DAG profiles during a EHC in 89 nondiabetic overweight and obese postmenopausal women. DAG levels were found to be significantly decreased for ISO and IRO individuals during the EHC, whereas only ISO subjects displayed a significant reduction of AG levels. Also, the ratio of AG/DAG

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Abstract

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(fasting and area under the curve (AUC)) was significantly decreased in ISO individuals (11).

Rodriguez et al. found that AG levels were increased, whereas DAG levels were decreased in obesity and obesity-associated type 2 diabetes. Body mass index, waist circumference, insulin, and homeostasis model assessment (HOMA) index were positively correlated with AG levels (13). However, it is noteworthy that in the latter study, blood samples were not collected in a way that prevents de-acylation of ghrelin; therefore, the data must be interpreted with caution.

Barazzoni et al. determined potential differential associations of ghrelin isoforms with IR and the impact of obesity on their plasma concentrations in 45 subjects with metabolic syndrome. Plasma insulin and HOMA-IR were negatively associated with DAG but positively with AG and AG/DAG ratios. Compared with the nonobese, obese metabolic syndrome patients had lower DAG but comparable AG and higher AG/DAG ratios. Apparently, obesity could alter circulating ghrelin profiles, and a relative AG excess or DAG deficiency can contribute to obesity-associated IR in metabolic syndrome (12).

Finally, Prader–Willi syndrome (PWS) is a leading genetic cause of obesity, characterized by hyperphagia, endocrine, and developmental disorders. It is believed that the hyperphagia results from impaired gut hormone signaling. Purcell et al. reported that compared with adiposity-matched controls, hyperphagia in PWS is not related to a lower postprandial glucagon-like peptide-1 or peptide YY response. However, elevated ghrelin levels in PWS did show a relation with increased hunger, which was unrelated to insulin levels (14).

Goldstone et al. investigated whether differences in appetite hormones may explain the development of abnormal eating behavior in 42 young (7 months–5 years) children with PWS. No significant relationship was found between eating behavior and the levels of any of the hormones measured or IR, independent of age. However, even in these young children, PWS was associated with low levels of the anorexigenic pancreatic polypeptide (PP), as has been described in older children and adults. Only later in life do hyperghrelinemia or hypoinsulinemia begin to contribute to the hyperphagia of PWS (15). It was hypothesized that abnormal development or sensitivity of pathways required for activity of these appetitive hormones (e.g. in the CNS) may be responsible for the emergence of hyperphagia in PWS.

**DAG is a functional ghrelin antagonist**

*The first indications*

It was the group of Broglio et al. (16) that shifted the existing paradigm by suggesting that DAG should be considered as a separate hormone. Others later supported their findings. Broglio et al. studied the interaction of the combined administration of AG and DAG in six normal young volunteers. As expected, AG administration alone markedly increased circulating GH, prolactin, ACTH, and cortisol levels. AG administration was also followed by a decrease in insulin levels and an increase in plasma glucose levels. While DAG administration alone had no effects, DAG together with AG diminished the insulin and glucose response to AG. Their findings indicated for the first time that DAG can be metabolically active by counteracting the effects of AG on insulin secretion and glucose metabolism.

Inhoff et al. (17) have also shown that the metabolic effects of AG (administered i.p.) could be abrogated by coadministration of DAG in nonfasted rats. DAG alone did not alter food intake, as was also reported by others, as, e.g. by Neary et al. (18). Double labeling revealed that nesfatin-1 immunoreactive neurons in the arcuate nucleus are activated by simultaneous injection of AG and DAG. These results suggest that DAG suppresses AG-induced food intake by preventing the increase in neuronal activity in the arcuate nucleus and recruiting nesfatin-1 immunopositive neurons (17). Kumar et al. also reported the antagonistic properties of DAG on AG (19).

By using isolated mouse pancreatic islets, they studied suppression of spontaneous secretion of PP by AG and obestatin. Interestingly, the PP-releasing effect of AG, but not obestatin, was counteracted by coadministration of DAG. Apparently, AG and obestatin inhibit spontaneous PP secretion at physiologically
relevant concentrations by acting through separate receptors (19).

Gauna et al. (20) reported that administration of AG in humans reduces insulin sensitivity, whereas the combination of AG plus DAG strongly improves insulin sensitivity. They studied the effects of acute administration of AG, DAG, and their combined administration in eight adult-onset GH-deficient patients. AG, which was rapidly cleared from the circulation, induced a rapid rise in glucose and insulin levels. AG and DAG coadministration, however, prevented the AG-induced rise in insulin and glucose. Moreover, cotreatment with DAG prevented AG-induced IR for at least 6 h after administration (20).

In vitro studies
Granata et al. (21) investigated the role of AG in pancreatic β-cell proliferation as well as apoptosis induced by serum starvation or cytokine (interferon γ/tumor necrosis factor α) treatment. They found that HIT-T15 cells (a hamster insulinoma cell line) expressed Ghrl but not the GHSR1a mRNAs. However, both DAG and AG recognized common high-affinity binding sites on these cells. Furthermore, both AG and DAG stimulated insulin secretion by these cells.

Interestingly, both DAG- and AG-induced cell survival and protection against apoptosis in isolated human islets of Langerhans. Insulin-positive β-cells in these islets expressed GHSR1a, explaining the mechanism of action for AG. However, competitive binding studies using radiolabeled DAG suggest that islets also express DAG and AG binding sites that are probably not the classical GHSR1a. Therefore, the effects of DAG and AG may also occur via mechanisms independent of the GHSR1a, likely mediated by specific AG and DAG binding sites (21).

Another indication that DAG acts as an AG antagonist was demonstrated by Gauna et al. (22). They reported that AG stimulates, whereas DAG inhibits glucose output by primary hepatocytes. AG, but not the GH secretagogue hexarelin, dose dependently induced glucose output. However, DAG dose dependently inhibited glucose release and completely reversed AG-induced glucose output. Interestingly, GHSR1a gene expression was not detectable in the hepatocyte preparations. The observed effects of DAG, in the absence of GHSR1a expression and lack of a response to hexarelin treatment, again indicate the involvement of an uncharacterized ghrelin receptor (subtype) (22).

More recently, Miegueu et al. (23) reported the direct effects of ghrelin gene family peptides on preadipocyte proliferation, differentiation, and adipocyte lipid and glucose metabolism in 3T3-L1 cells. In these adipocytes, fatty acid (FA) uptake was increased at sub-nanomolar concentrations in a concentration-dependent manner by obestatin and DAG, in particular. Interestingly, DAG stimulation of FA uptake was blocked by the [D- Lys3]-GHRP-6, which suggests that this GHSR1a antagonist (although not very selective for the GHSR1a (24, 25)) can block the hypothetical DAG receptor as well (23). An independent report on the effects of AG and DAG in adipocytes by Rodriguez et al. (13) also shows that AG and DAG can stimulate lipid accumulation in human visceral adipocytes.

In agreement with earlier studies in primary adipocytes (26), DAG and obestatin significantly decreased lipolysis. Baragli et al. (27) also reported that AG and DAG block isoproterenol and forskolin-induced lipolysis; a process in which phosphodiesterase (PDE) seems to be involved. In line with this, cilostamide, a specific PDE3B inhibitor, blocked the effects of AG and DAG on isoproterenol-induced lipolysis. In particular, selective inhibition of PI3K iso-enzyme γ prevented AG and DAG effects on isoproterenol-stimulated lipolysis, impeding AKT phosphorylation (Table 1) (27).

Lear et al. (28) also reported that DAG has specific binding sites and different metabolic effects in the murine HL-1 adult cardiomyocyte cell line. They compared the effects of AG and DAG on glucose and FA uptake and attempted to identify DAG-specific binding sites in cardiomyocytes. AG and DAG had opposing metabolic effects: DAG increased medium-chain FA uptake, whereas neither AG alone nor in combination with DAG did so. In HL-1 cells and primary cultures of neonatal rat cardiomyocytes, DAG but not AG increased insulin-induced translocation of glucose transporter-4 from nuclear to cytoplasmic compartments. HL-1 and primary-cultured mouse and rat cardiomyocytes each possessed two independent, specific, binding sites for DAG that were not recognized by AG. Neither AG nor DAG affected cell viability, whereas both isoforms were equally protective against apoptosis. Apparently, in cardiomyocytes, DAG binds to specific receptors and has effects on glucose and medium-chain FA uptake that are distinct from those of AG (28).

Moazed et al. (29) also suggested the presence of a non-GHSR1a that transduces DAG signals. They reported that DAG evokes endothelium-dependent vasodilatation of rat mesenteric vascular bed via activation of potassium channels. However, this vasodilator response in the perfused rat mesenteric vascular bed was not mediated by the classical GHSR1a (29).

In vivo results in rodents
In 2005, Asakawa et al. (30) reported that DAG appeared to decrease food intake and gastric emptying rate through actions on the hypothalamus. DAG transgenic mice exhibit a decrease in body weight, food intake, and fat pad mass accompanied by moderately decreased linear growth, along with a decrease in gastric emptying. This again indicates, but now in vivo, that in contrast to AG, DAG induces decrease in gastric emptying.
negative energy balance by decreasing food intake and delaying gastric emptying (30).

Following on from the study of Asakawa et al., Zhang et al. studied the effects on adiposity and glucose metabolism of overexpressing DAG in the fat depots of mice, using the FA-binding protein-4 (FABP4) promoter to drive expression of a murine preproghrelin transgene (31). In the presence of high DAG levels, epididymal and perirenal fat masses decreased. FABP4-ghrelin transgenic mice appeared to be resistant to obesity induced by high-fat diet. Glucose tolerance tests showed significantly faster clearance of glucose in FABP4-ghrelin transgenic mice than in controls. Insulin sensitivity testing showed that FABP4-ghrelin transgenic mice had a significantly greater hypoglycemic response to insulin, indicating markedly improved insulin sensitivity (31). As the FABP4 promoter is also active in the hypothalamus (32), it is very likely that FABP4-ghrelin transgenic mice overexpress DAG in the hypothalamus as well and that part of the effect may be through actions of DAG at a central level. Several other studies, however, could not find an appetite-inhibiting effect of DAG (e.g. Inhoff et al. (17) and Neary et al. (18)).

The potential clinical importance of inhibiting AG has recently been reported in rodents. Barnett et al. (33) reported the design, synthesis, and characterization of GO-CoA-Tat, a peptide-based bi-substrate analog that antagonizes GOAT. I.p. administration of GO-CoA-Tat improved glucose tolerance and reduced weight gain in wild-type (WT) mice. Interestingly, they did not observe this reduction in weight gain in ghrelin-deficient mice that lack both AG and DAG. Apparently, GO-CoA-Tat administration reduces weight gain provided that the ghrelin gene is still present (so ghrelin effect is low and DAG effect is present) and that in ghrelin KO mice this effect is absent (so both ghrelin and DAG effects are low). This might suggest that the presence of DAG is at least partly responsible for the inhibition in weight gain (as acyl ghrelin effects are already low due to the GO-CoA-Tat antagonizes GOAT. I.p. administration of GO-CoA-Tat, a peptide-based bi-substrate analog that antagonizes GOAT.

![Innate immune response](image)

**Table 1** Summary of *in vivo* and *in vitro* study results of DAG effects in animals and in humans

<table>
<thead>
<tr>
<th>Reported observations in which DAG effects were found</th>
<th><em>In vitro</em> animal studies</th>
<th><em>In vivo</em> animal studies</th>
<th><em>In vivo</em> human studies</th>
<th><em>In vivo</em> human studies</th>
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<tbody>
<tr>
<td>DAG has specific binding sites and different metabolic effects in the murine HL-1 adult cardiomyocyte cell line</td>
<td>(27)</td>
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<td>DAG dose-dependently inhibits glucose release and reverses ghrelin-induced glucose output in primary porcine hepatocytes</td>
<td>(22)</td>
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<tr>
<td>Direct effects of ghrelin gene family peptides on preadipocyte proliferation, differentiation, and adipocyte lipid and glucose metabolism in 3T3-L1 cells</td>
<td>(23)</td>
<td></td>
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<tr>
<td>DAG can improve cell survival and provide protection against apoptosis in isolated islets of Langerhans</td>
<td>(21, 56)</td>
<td></td>
<td>(21)</td>
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<tr>
<td>DAG probably has its own receptor(s)</td>
<td></td>
<td>(5, 36-38)</td>
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<tr>
<td>DAG evokes GHSR1a-independent endothelium-dependent vasodilatation of rat mesenteric vascular bed via activation of potassium channels</td>
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<td>Both ghrelin and DAG can block isoproterenol- and forskolin-induced lipolysis</td>
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<td>DAG exhibits pro-anabolic and anticaotic effects on C2C12 myotubes exposed to cytokines and reduces burn-induced muscle proteolysis in rats</td>
<td>(26)</td>
<td></td>
<td>(39)</td>
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<td>The metabolic effects of ghrelin can be abrogated by coadministration of DAG</td>
<td></td>
<td></td>
<td>(17, 19)</td>
<td>(57, 58)</td>
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<tr>
<td>Insulin-resistant obese subjects have an elevated ghrelin/DAG ratio</td>
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<td></td>
<td>(11, 12)</td>
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<tr>
<td>Obese mice and humans have lowered DAG levels</td>
<td></td>
<td></td>
<td>(8, 9)</td>
<td>(10)</td>
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<tr>
<td>DAG can decrease food intake and gastric emptying rate via the hypothalamus</td>
<td></td>
<td>(29)</td>
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<tr>
<td>Insulin sensitivity was significantly increased in FABP4-ghrelin transgenic mice that become resistant to obesity induced by high-fat diet</td>
<td>(30)</td>
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<tr>
<td>In obesity and obesity-associated type 2 diabetes, ghrelin levels are increased and DAG levels decreased</td>
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<td></td>
<td>(34)</td>
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<tr>
<td>DAG administration strongly inhibits ghrelin levels</td>
<td></td>
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<td>(13)</td>
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<tr>
<td>Improvements in body weight and body composition are associated with an increase in DAG</td>
<td></td>
<td></td>
<td>(35)</td>
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<tr>
<td>DAG can rescue endothelial progenitor cell function in individuals with type 2 diabetes</td>
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<td>(40)</td>
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Another report on the potential beneficial effects of antagonizing AG showed that the ghrelin receptor antagonist [δ-Lys3]-GHRP-6, after 7 days of subcutaneous treatment, markedly decreased food intake in ovariectomized mice fed both high-fat and standard diets. Furthermore, Maletinska et al. (34) reported that this AG antagonist reduced body weight, blood glucose, insulin, and leptin.

In vivo results in humans

Benso et al. (35) recently reported that i.v. DAG administration improves glucose metabolism and inhibits lipolysis in healthy volunteers. Apparently, and in contrast to the diabetogenic action of AG, DAG displays hypoglycemic properties. They studied the effects of a 16-h overnight infusion of DAG or saline in eight normal subjects. The overall insulin levels (as AUC) were not significantly modified by DAG infusion. However, postprandial insulin levels after both dinner and breakfast were significantly increased following DAG treatment compared with placebo.

We recently performed a clinical study in eight overweight patients with controlled type 2 diabetes on the effects of a continuous overnight infusion of two doses of DAG vs placebo in a double blind crossover study on AG concentrations and the glucose and insulin responses to a standard breakfast meal (SBM).

Early morning AG levels (during the overnight DAG infusion, just before the SBM) dropped significantly. DAG administration also markedly decreased AG levels following breakfast.

Moreover, overnight infusions of DAG significantly decreased postprandial glucose levels after the SBM (using continuous glucose monitoring, as well as peak serum glucose levels). There was a strong correlation between fasting AG levels and the magnitude of suppression of postprandial glucose levels by DAG administration. The higher the fasting AG levels, the more pronounced the decrease in glucose levels. This close correlation between the effect of DAG infusion on postprandial glycemia and the preprandial AG levels suggests a potential clinical benefit of DAG administration, especially in subjects with relatively high levels of AG.

The concept that low AG/DAG ratios are accompanied by an improved metabolic state was supported by Cederberg et al. They reported that after 6 months of an intensive long-term physical exercise by 552 young men undergoing military service, improvements in body weight and body composition were associated with an increase in DAG (36). However, the samples in this study were collected in such a way that acyl ghrelin might easily convert to DAG, although this would be the case in both the baseline and the 6-month data (36).

GHSR1a- and ghrelin-independent effects of DAG

Delhanty et al. studied the GH$^{-}$SR1a-independent effects of DAG in mice in vivo. Using microarrays, they examined rapid effects of DAG on genome-wide expression patterns in fat, muscle, and liver of GH$^{-}$SR1a-deficient mice. The expression data were analyzed using Ingenuity Pathways Analysis and Gene Set Enrichment Analysis. Regulation of subsets of these genes was verified by quantitative PCR in independent experiments. They found that DAG acutely regulated clusters of genes involved in glucose and lipid metabolism in all three tissues, consistent with enhancement of insulin sensitivity. This pivotal set of studies demonstrates that DAG rapidly modulates the expression of metabolically important genes in these tissues, which are central to the control of lipid and glucose homeostasis. These results strongly indicate a direct, GH$^{-}$SR1a-independent, action of DAG that improves insulin sensitivity and metabolic profile (5).

Gauna et al. (37) reported the results of a study on the interactions between AG and DAG on ghrelin receptors in INS-1E rat insulinoma cells, using insulin secretion after 30-min static incubation as a read-out. Both DAG and AG dose dependently stimulated insulin release in the nanomolar range. As expected, the AG-induced insulin output was antagonized by the two GH$^{-}$SR1a antagonists [δ-Lys3]-GHRP-6 and BIM28163. These GH$^{-}$SR1a blockers, however, did not block DAG-induced insulin secretion. These data strongly suggest the existence of a specific receptor for DAG, other than the GH$^{-}$RS-R1a, that DAG might share with AG.

Halem et al. in 2005 (38) also reported on the potential presence of a non-GH$^{-}$SR1a. They demonstrated that the GH$^{-}$SR1a blocker BIM-28163 that was also used in the Gauna study above blocked AG activation of the GH$^{-}$SR1a and inhibited AG-induced GH secretion in vivo. To their surprise, they observed that BIM-28163 also acted as an agonist with regard to stimulating weight gain. Again Halem’s results suggest the presence of an unknown ghrelin receptor that modulates ghrelin actions on weight gain (38). The great clinical importance of these studies is the notion that GH$^{-}$SR1a may not be the most appropriate target for antiobesity strategies. An example of this is a report by Chen et al. (39) who observed a potential role for the CRF type 2 receptor in DAG’s action in regulating food intake.

Non-metabolic actions of AG and DAG

Intriguing reports have appeared recently showing that AG and DAG have effects on several non-metabolic conditions. An example of this is a study by Sheriff et al. They observed that DAG exhibits pro-anabolic and anticatabolic effects on C2C12 myotubes exposed to...
cytokines and reduces burn-induced muscle proteolysis in rats (40).

Another study on the effects of DAG suggests that it can rescue endothelial progenitor cell (EPC) function in individuals with type 2 diabetes (41). Togliatto et al. investigated the potential of DAG to reverse diabetes-associated pathology in individuals with type 2 diabetes, and ob/ob mice. Systemic administration of DAG, but not AG, prevented diabetes-induced damage to EPCs by modulating the NADPH oxidase regulatory protein Rac1 and improved vasculogenic potential both in individuals with type 2 diabetes and in ob/ob mice. In addition, unlike AG, DAG facilitated the recovery of bone marrow EPC mobilization (41).

In line with this, a recent report by Zhang et al. (42) reported that low (total) ghrelin levels are closely associated with severity and morphology of angiographically detected coronary atherosclerosis in patients with diabetes mellitus. Moreover, Yang et al. (43) studied the effect of ghrelin on angiotensin II (Ang II)-induced apoptosis of H9c2 cardiomyocytes. The results showed that ghrelin prevents Ang II-induced H9c2 cell death. Their data suggest that ghrelin may play an antagonistic role in Ang II-induced cardiomyocyte apoptosis by decreasing angiotensin receptor expression and inhibiting activation of the endoplasmic reticulum stress pathway (43).

Wu et al. (44) reported that ghrelin maintains the cardiovascular stability in severe sepsis. In their study, male adult rats were made septic by cecal ligation and puncture (CLP). At 5 h after CLP, a bolus i.v. injection of ghrelin was followed by continuous infusion of 12 nmol ghrelin via a primed mini-pump for 15 h. Treatment with ghrelin significantly augmented the maximal rates of ventricular pressure increase and decrease by 36 and 35% respectively. Ghrelin treatment also reversed the suppression of norepinephrine-induced vascular contraction and acetylcholine-induced endothelium-dependent vascular relaxation caused by CLP (44).

Another unexpected link between the ghrelin system and a clinical condition is that ghrelin only very recently has been introduced into the field of epilepsy and has already led to contradictory clinical publications. In humans, AG and DAG, on the whole, are found to decrease following an epileptic seizure. Furthermore, the majority of animal studies on this subject demonstrate that ghrelin has an anticonvulsant properties, but very little is known about the mechanisms by which ghrelin inhibits seizures. A recent valuable review by Portelli et al. addresses possible mechanisms of action for both AG (and DAG) in the prevention of seizures and indirectly gives an insight into modes of action of these peptides for researchers studying ghrelin in other fields of research (45).

Finally, the ghrelin system is also linked to gonadal function (46, 47), gut motility (39, 48, 49, 50), and cancer (51, 52, 53, 54), and it is highly likely that this relatively new hormone family will be linked to other medical conditions in the near future.

Conclusion
In conclusion, DAG is more than just a degradation product of acyl ghrelin. It appears to be a separate hormone, with its own actions, independent from the actions of ghrelin. DAG can only bind and activate the GHSR1a at supra-physiological concentrations. This low affinity interaction rules out the GHSR1a as a mediator of DAG activities at physiological concentrations. Increased AG/DAG ratios are linked to obesity and diabetes. An important caveat to interpretation of the available data, however, is that some of the reported studies did not measure directly DAG. DAG administration induces a rapid decrease in circulating AG levels, along with an improvement in glycemic control.

The insight that DAG can antagonize AG via several routes and should be considered as a natural antagonist of ghrelin could lead to the development of ghrelin inhibitors that might be helpful in the treatment of conditions in which high AG (actions) are linked with the severity of the disease, as in, e.g. PWS patients and in obese type 2 diabetes subjects.

Declaration of interest
P J D Delhanty and S J Neggers declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review reported. A J van der Lely is scientific advisor and shareholder of Aller Pharma SAS, Ecullly, France.

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References


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