Fifteen years of GH replacement increases bone mineral density in hypopituitary patients with adult-onset GH deficiency

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Abstract

Objective: Few studies have determined the effects of more than 5–10 years of GH replacement in adults on bone mineral content (BMC) and bone mineral density (BMD).

Design/patients: In this prospective, single-centre, open-label study, the effects of 15 years of GH replacement on BMC and BMD, measured using dual-energy X-ray absorptiometry, were determined in 126 hypopituitary adults (72 men) with adult-onset GH deficiency (GHD). Mean age was 49.4 (range 22–74) years at the initiation of the study.

Results: The mean initial GH dose of 0.63 (S.E.M. 0.03) mg/day was gradually lowered to 0.41 (0.01) mg/day after 15 years. The mean serum IGF1 SDS increased from 1.69 (0.11) at baseline to 0.63 (0.16) at the study end (P<0.001 vs baseline). The 15 years of GH replacement induced a sustained increase in total body BMC (+5%, P<0.001) and BMD (+2%, P<0.001). Lumbar (L2–L4) spine BMC increased by 9% (P<0.001) and BMD by 5% (P<0.001). In femur neck, a peak increase in BMC and BMD of 7 and 3%, respectively, was observed after 7 years (both P<0.001). After 15 years, femur neck BMC was 5% above the baseline value (P<0.01), whereas femur neck BMD had returned to the baseline level. In most variables, men had a more marked response to GH replacement than women.

Conclusions: Fifteen-year GH replacement in GHD adults induced a sustained increase in total body and lumbar (L2–L4) spine BMC and BMD. In femur neck, BMC and BMD peaked at 7 years and then decreased towards baseline values.

Introduction

Young adults with GH deficiency (GHD) have reduced bone mineral content (BMC) and bone mineral density (BMD) (1, 2, 3, 4). This reduction in BMC and BMD is more marked in patients with childhood-onset (CO) GHD than in patients with adult-onset (AO) GHD (5). Elderly GHD adults do not have a reduced bone mass and density compared with age-matched healthy controls (6, 7). However, adult GHD patients without GH replacement are at a higher risk of fractures than healthy controls of the same age (8, 9, 10). It is therefore possible that other factors than reduced BMD, such as increased number of falls due to visual deficits caused by pituitary tumours or their treatment, could contribute to the increased fracture risk in GHD adults.

Initiation of GH replacement in adults with GHD induces an initial increase in bone resorption that may result in unchanged or even reduced bone mass (2, 3). This is followed by increased bone formation and a net increase in bone mass seen after 12–18 months of GH replacement (2, 3). GH exerts direct effects on bone (2, 11), but it has been questioned whether the direct effects can fully explain the effects of GH on bone. Indirect effects, such as increased muscle performance by GH, could also be of importance (12). Furthermore, the responsiveness to GH replacement is dependent on the group of patients studied. The increase in BMC and BMD is larger in patients with CO GHD compared with patients with AO GHD (5) and also more marked in men compared with women (13, 14, 15, 16, 17, 18). Finally, the response to GH is most marked at weight-bearing locations and the increase in BMC is greater than that of BMD (15, 17).

GH replacement has been shown to induce a progressive increase in bone mass and density up to 5–6 years of treatment (15, 19, 20, 21). One study of 7 years of GH replacement showed an increase in BMC and BMD up to 4 years and after that BMC and BMD reached a plateau (13). In another study, total body and lumbar (L2–L4) spine BMC and BMD increased progressively up to 10 years of GH replacement, whereas femur neck BMC and BMD reached a peak value after 5–7 years of treatment (22).
GHD in adults is a chronic disease and GH replacement may therefore continue over decades. Long-term studies on the effects of GH treatment are therefore important. There are at present no studies of more than 10 years of GH replacement. The aim of this study was to investigate the effects of 15 years of GH replacement therapy on BMC and BMD in patients with AO GHD recruited at a single centre.

### Patients and methods

**Patients**

In this study, 126 consecutive GHD adults (72 men) with a mean age of 49.4 (range 22–74) years were included between 1990 and 1994. All patients had AO pituitary disease and all had known pituitary disease or other anterior pituitary hormonal deficiencies. The pituitary deficiency was mainly caused by pituitary tumours and/or their treatment (Tables 1 and 2). The patients had been treated with pituitary surgery (n=49), surgery and radiotherapy (n=49), radiotherapy alone (n=11) and no treatment (n=17).

Most patients had multiple anterior pituitary hormonal deficiencies (Tables 1 and 2). Possibly due to late effects of radiotherapy, several patients had more pituitary hormonal deficiencies at study end compared with baseline (Tables 1 and 2). In 116 of the patients, the diagnosis of GHD was based on a peak GH < 3 μg/l during a stimulation test (insulin (n=112), GHRH (n=2) and glucagon (n=2)). In nine patients, the diagnosis was based on a 24-h GH profile (sampling every 30 min). In one patient, who had a known anterior pituitary disease and three additional hormonal deficiencies, the diagnosis was based on a low serum insulin-like growth factor 1 (IGF1) level. When required, patients received adequate and stable therapy with glucocorticoids, thyroid hormone and desmopressin. All the testosterone-deficient men received testosterone therapy. However, at baseline 52% (25 out of 48) and at study end 31% (15 out of 48) of the oestrogen-deficient women received oestrogen replacement therapy. Five patients received treatment with vitamin D, calcium and/or bisphosphonates after 5 (n=1), 11 (n=1) and 13 (n=3) years of GH replacement. In these patients, the measurements performed after the initiation of treatment with vitamin D, calcium and/or bisphosphonates were excluded in the statistical analysis, and the last value before treatment was carried forward according to the intention-to-treat approach used.

Fourteen of the patients died during the study period (cerebral infarction (n=3), myocardial infarction (n=2), pneumonia (n=2), sudden death of unknown cause (n=1), renal cancer (n=1), subarachnoid haemorrhage (n=1), chronic obstructive pulmonary disease (n=1), perimyocarditis (n=1), pulmonary embolism (n=1) and coronary artery disease (n=1)). Fourteen patients discontinued GH replacement (lack of compliance (n=7), old age (n=3), colon cancer (n=1), epilepsy after stroke (n=1), pulmonary cancer (n=1) and chronic lymphocytic leukaemia (n=1)). Eight patients were lost to follow-up because they moved to other cities or abroad. Thus, 90 of the 126 patients completed the 15 years of GH replacement. All patients were, however, retained in the statistical analysis according to the intention-to-treat approach used.

**Study protocol**

This is an ongoing, prospective, open-label treatment trial. The first 64 patients initially received a weight-based GH dose of 11.9 μg/kg per day (0.25 IU/kg per week). During the first 2–3 years of treatment, the GH dose was gradually lowered and individualised when the weight-based dose regimen was abandoned. In the remaining 62 patients, the dose of GH was individualised from the initiation of the treatment, with the aim of normalising serum IGF1 concentration and body composition in each patient.

At baseline, and after each year of GH replacement until 5 years, and then after 7, 10, 12 and 15 years, physical and laboratory including measurements of bone mass examinations were performed. Dose titration

### Table 1 Causes of pituitary deficiency in the study population of 126 adults with adult-onset GH deficiency.

<table>
<thead>
<tr>
<th>Type of deficiency</th>
<th>Men (n)</th>
<th>Women (n)</th>
<th>Total (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pituitary adenoma</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-secreting</td>
<td>44</td>
<td>22</td>
<td>66</td>
</tr>
<tr>
<td>Secreting</td>
<td>11</td>
<td>13</td>
<td>24</td>
</tr>
<tr>
<td>Craniopharyngioma</td>
<td>7</td>
<td>6</td>
<td>13</td>
</tr>
<tr>
<td>Empty sella</td>
<td>3</td>
<td>4</td>
<td>7</td>
</tr>
<tr>
<td>Sheehan’s syndrome</td>
<td>0</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>Idiopathic</td>
<td>0</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Other pathology*</td>
<td>7</td>
<td>4</td>
<td>11</td>
</tr>
<tr>
<td>Total</td>
<td>72</td>
<td>54</td>
<td>126</td>
</tr>
</tbody>
</table>

*One patient with a meningioma, two with pituitary apoplexia, one with histiocytosis X, two with sarcoidosis, one with a cholesteatoma, one with an ependymoma, one with a dysgerminoma and two with trauma.

### Table 2 Number of pituitary deficiencies at baseline and after 15 years of GH replacement (study end) in the study population of 126 adults with adult-onset GH deficiency (GHD).

<table>
<thead>
<tr>
<th>Type of deficiency</th>
<th>Baseline (n)</th>
<th>Study end (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Isolated GHD</td>
<td>7</td>
<td>3</td>
</tr>
<tr>
<td>No. of additional deficiencies</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>7</td>
<td>1</td>
</tr>
<tr>
<td>2</td>
<td>10</td>
<td>14</td>
</tr>
<tr>
<td>3</td>
<td>48</td>
<td>54</td>
</tr>
<tr>
<td>Diabetes insipidus</td>
<td>22</td>
<td>23</td>
</tr>
</tbody>
</table>

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and safety monitoring were performed every third month during the first year and every sixth month thereafter. Body weight was measured in the morning to the nearest 0.1 kg and body height was measured to the nearest 0.01 m. Body mass index (BMI) was calculated as the weight in kilograms divided by the height in meters squared. No effort was made to influence the patients’ physical activity level during the study period.

**Ethical considerations**

Informed consent was obtained from all patients. The study was approved by the Regional Ethics Review Board at the University of Gothenburg and the Swedish Medical Products Agency (Uppsala, Sweden).

**BMC and BMD**

BMC and BMD were measured by dual-energy X-ray absorptiometry (DXA) in the total body, lumbar spine and femur neck as described previously (23). From the initiation of the study until the end of 1999, a LUNAR DPX-L scanner was used (Scanex, Helsingborg, Sweden). Software versions were changed several times (from 1.1 to 1.35), version 1.33 being used during the major part of this period of the study. In-house precision error on the scanner used, as determined from duplicate examinations in ten healthy subjects, was 1.9% for total body BMC. From January 2000, a LUNAR Prodigy scanner (Scanex) was used. Software versions were upgraded several times during data collection, from version 5.70 to 8.10. The precision of the scanner was estimated from repeated measurements on different days in 30 subjects with coefficients of variation (CV) of total body BMC of 1.4%. Before the change of scanner in 2000, 31 subjects (19 males and 12 females), ranging in weight from 51 to 112 kg, were scanned on both scanners on the same day. Total body BMC was not significantly different between scanners (mean difference, 0.019 g/cm²; 95% confidence interval for the difference, −0.0079 to 0.00458 g/cm²).

Daily quality control was performed according to the manufacturer’s protocol. A spine phantom was measured at least once a week. Every single spine phantom measurement was compared with a baseline value based on a mean of ten repeated measurements. A maximum 1.5% deviation from the baseline value was accepted. A European phantom (COMAC-BME Quantitative Assessment of Osteoporosis Study Group) was measured once a year. BMC z-score, which is the difference in s.d. of age- and sex-matched healthy subjects, and t-score, which is the difference in s.d. of sex-matched young (20–39 years) healthy subjects, were determined using the Lunar DPX-L Software program. The reference database used was the LUNAR USA reference population for the region examined.

**Biochemical analysis**

Serum IGF1 concentration, until June 2004, was determined using a hydrochloric acid–ethanol extraction RIA (Nichols Institute Diagnostics. San Juan Capistrano, CA, USA). Inter- and intra-assay CV were 5.4 and 6.9% respectively, at a mean serum IGF1 level of 126 µg/l, and 4.6 and 4.7% respectively, at a mean serum IGF1 level of 327 µg/l. From June 2004 to August 2006, serum IGF1 concentration was determined using a chemiluminescence immunoassay (Nichols Advantage; Nichols Institute Diagnostics) (24). From September 2006, serum IGF1 level was determined using an automated chemiluminescent assay system (IMMULITE 2500, Diagnostic Products Corp., Los Angeles, CA, USA). The standard used for calibration of the IGF1 assays was the WHO NIBSC 1st IRR 87/518 throughout the study period. The individual serum IGF1 values were compared with age- and sex-adjusted values obtained from a reference population (25), and the individual IGF1 SDSs, taking into account age and gender, were then calculated (26).

**Statistical methods**

All the descriptive statistical results are presented as the mean and s.e.m. For all variables, within-group differences were calculated using a repeated measures ANOVA, with all data obtained from all time points, and with time as the independent variable. Post hoc analysis was performed using Student–Newman–Keuls test. Gender differences and differences between women on oestrogen replacement and women without oestrogen replacement were calculated by a two-way ANOVA, with all data obtained from all time points, and with gender or oestrogen treatment as the independent variables. In order to eliminate the baseline differences, data were transformed into percent change or change from baseline before the between-group analyses. All analyses were performed according to the intention-to-treat principle (using the carry forward principle). A two-tailed P < 0.05 was considered significant.

**Results**

**GH dose and serum IGF1**

The dose of GH prescribed at the baseline visit was 0.63 (0.03) mg/day (Table 3). The dose was then gradually reduced to 0.41 (0.01) mg/day at study end. Serum IGF1 levels increased during the first years of the study and then decreased to some extent. Mean IGF1 SDS (adjustment for age and gender) was above the normal range during the first 3 years of the study, but after that, it was within the normal range (±2 s.d.).
Table 3 The dose of GH during 15 years of GH replacement in 126 GH-deficient adults and the effects of this treatment on serum IGF1 concentration and IGF1 SDS. All values are shown as the mean (S.E.M.). The statistical analyses are based on a one-way ANOVA followed by Student–Newman–Keuls post hoc test.

<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>3 years</th>
<th>5 years</th>
<th>7 years</th>
<th>10 years</th>
<th>12 years</th>
<th>15 years</th>
<th>P value (10–15 years)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dose of GH (mg/day)</td>
<td>0.63 (0.03)</td>
<td>0.51 (0.02)</td>
<td>0.49 (0.02)</td>
<td>0.46 (0.02)</td>
<td>0.44 (0.02)</td>
<td>0.42 (0.02)</td>
<td>0.41 (0.01)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Serum IGF1 (μg/l)</td>
<td>103 (6)</td>
<td>298 (10)</td>
<td>273 (10)</td>
<td>246 (8)</td>
<td>206 (8)</td>
<td>188 (7)</td>
<td>183 (7)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>IGF1 SDS</td>
<td>−1.69 (0.11)</td>
<td>2.28 (0.21)</td>
<td>1.89 (0.21)</td>
<td>1.47 (0.18)</td>
<td>0.84 (0.17)</td>
<td>0.56 (0.16)</td>
<td>0.62 (0.16)</td>
<td>&lt;0.05</td>
</tr>
</tbody>
</table>

P values (10–15 years) are based on the statistical analysis between the 10- and 15-year values. NS, non-significant. †P<0.001 (for the dose of GH vs initial GH dose; for other variables vs baseline).

**Height, weight and BMI**

Mean body height decreased from 171.8 (0.94) cm at baseline to 171.3 (0.97) cm after 15 years (P<0.001; data not shown). Mean body weight increased from 81.5 (1.5) to 83.7 (1.5) kg (P<0.001) and BMI increased from 27.6 (0.44) kg/m² at baseline to 28.5 (0.44) kg/m² at the end of the study (P<0.001).

**BMC and BMD**

GH replacement gradually increased total body BMC during the first 5 years of treatment as measured using DXA (Table 4). Thereafter, total body BMC increased slowly throughout the study period. After 15 years, mean total body BMC was 5% above the baseline value (P<0.001). Total body BMD had increased by 2% after 10 years (P<0.001 vs baseline) and then reached a plateau. Total body t-score, after an initial decrease, increased up to 10 years and then stayed at a constant level. Total body z-score tended to decrease during the first 5 years of treatment and then increased during the remaining part of the study.

Lumbar (L2–L4) spine BMC increased throughout the study period and was 9% above the baseline value after 15 years of GH replacement (P<0.001 vs baseline; Table 4). Lumbar (L2–L4) spine BMC and t-score increased up to 10 years of GH treatment and remained at a constant level between 10 and 15 years. After 15 years, lumbar (L2–L4) spine BMC was 5% above the baseline value (P<0.001 vs baseline). Lumbar (L2–L4) spine z-score increased progressively throughout the study period.

Femur neck BMC reached a maximum level of 7% above baseline (P<0.001) after 7 years (Table 4). After that, femur neck BMC decreased and was 5% above the baseline value after 15 years (P<0.01). Femur neck BMD increased to a maximum level of 3% above baseline after 7 years of treatment (P<0.001). Femur neck BMD then decreased and returned to the baseline level after 15 years. Femur neck z-score reached a maximum after 7 years and then decreased but remained significantly elevated over the baseline level after 15 years.

Table 4 Effects of 15 years of GH replacement in 126 adults with GH deficiency on BMC, BMD, z-score and t-score as measured using DXA. All values are shown as the mean (S.E.M.). The statistical analyses are based on a repeated measures ANOVA followed by Student–Newman–Keuls post hoc test.

<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>3 years</th>
<th>5 years</th>
<th>7 years</th>
<th>10 years</th>
<th>12 years</th>
<th>15 years</th>
<th>P value (10–15 years)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DXA total body</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>BMC (kg)</td>
<td>2.73 (0.06)</td>
<td>2.77 (0.06)</td>
<td>2.81 (0.06)†</td>
<td>2.83 (0.06)†</td>
<td>2.83 (0.07)†</td>
<td>2.83 (0.07)†</td>
<td>2.87 (0.07)†</td>
<td>NS</td>
</tr>
<tr>
<td>BMC (g/cm²)</td>
<td>1.17 (0.01)</td>
<td>1.16 (0.01)</td>
<td>1.16 (0.01)</td>
<td>1.17 (0.01)</td>
<td>1.19 (0.01)</td>
<td>1.19 (0.01)</td>
<td>1.19 (0.02)</td>
<td>NS</td>
</tr>
<tr>
<td>z-Score (s.d.)</td>
<td>−0.05 (0.13)</td>
<td>−0.19 (0.14)</td>
<td>−0.18 (0.14)†</td>
<td>0.04 (0.15)</td>
<td>0.19 (0.16)†</td>
<td>0.22 (0.17)†</td>
<td>0.19 (0.17)†</td>
<td>NS</td>
</tr>
<tr>
<td>z-Score (s.d.)</td>
<td>−0.11 (0.11)</td>
<td>−0.20 (0.10)</td>
<td>−0.19 (0.11)</td>
<td>0.06 (0.11)†</td>
<td>0.36 (0.19)†</td>
<td>0.43 (0.14)†</td>
<td>0.42 (0.13)†</td>
<td>NS</td>
</tr>
<tr>
<td>DXA lumbar (L2–L4) spine</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>BMC (g)</td>
<td>55.5 (1.5)</td>
<td>58.1 (1.7)†</td>
<td>59.1 (1.8)†</td>
<td>59.5 (1.8)†</td>
<td>60.3 (1.9)†</td>
<td>60.3 (1.9)†</td>
<td>60.7 (2.0)†</td>
<td>NS</td>
</tr>
<tr>
<td>BMC (g/cm²)</td>
<td>1.17 (0.02)</td>
<td>1.20 (0.02)†</td>
<td>1.22 (0.02)†</td>
<td>1.22 (0.02)†</td>
<td>1.24 (0.02)†</td>
<td>1.24 (0.02)†</td>
<td>1.23 (0.02)†</td>
<td>NS</td>
</tr>
<tr>
<td>z-Score (s.d.)</td>
<td>−0.44 (0.15)</td>
<td>−0.13 (0.17)†</td>
<td>0.03 (0.18)†</td>
<td>0.06 (0.18)†</td>
<td>0.19 (0.20)†</td>
<td>0.13 (0.20)†</td>
<td>0.18 (0.21)†</td>
<td>NS</td>
</tr>
<tr>
<td>z-Score (s.d.)</td>
<td>−0.01 (0.14)</td>
<td>0.19 (0.14)</td>
<td>0.35 (0.15)†</td>
<td>0.39 (0.16)†</td>
<td>0.63 (0.18)†</td>
<td>0.62 (0.18)†</td>
<td>0.77 (0.19)†</td>
<td>0.01</td>
</tr>
<tr>
<td>DXA femur neck</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>BMC (g)</td>
<td>4.87 (0.14)</td>
<td>5.04 (0.14)</td>
<td>5.14 (0.12)†</td>
<td>5.20 (0.12)†</td>
<td>5.15 (0.13)†</td>
<td>5.13 (0.13)†</td>
<td>5.12 (0.13)†</td>
<td>NS</td>
</tr>
<tr>
<td>BMC (g/cm²)</td>
<td>0.94 (0.01)</td>
<td>0.96 (0.01)*</td>
<td>0.96 (0.02)†</td>
<td>0.97 (0.02)†</td>
<td>0.96 (0.02)</td>
<td>0.95 (0.02)</td>
<td>0.94 (0.02)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>z-Score (s.d.)</td>
<td>−0.71 (0.12)</td>
<td>−0.45 (0.14)†</td>
<td>−0.49 (0.11)†</td>
<td>−0.45 (0.12)†</td>
<td>−0.52 (0.12)*</td>
<td>−0.63 (0.12)</td>
<td>−0.75 (0.12)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>z-Score (s.d.)</td>
<td>−0.23 (0.10)</td>
<td>0.10 (0.12)†</td>
<td>0.17 (0.10)†</td>
<td>0.24 (0.10)†</td>
<td>0.24 (0.10)†</td>
<td>0.24 (0.10)†</td>
<td>0.18 (0.11)†</td>
<td>0.15 (0.11)†</td>
</tr>
</tbody>
</table>

P values (10–15 years) are based on the statistical analysis between the 10- and 15-year values. NS, non-significant; n, number of patients on GH replacement at different time points; however, all 126 patients were included in the statistical analysis according to the intention-to-treat approach used. A separate analysis of the patients (n=90) who completed the 15 years of GH replacement showed approximately similar results (data not shown). *P<0.05; †P<0.01; ‡P<0.001 vs baseline.
Effects of 15 years of GH replacement on bone

Women on oestrogen replacement vs women without oestrogen replacement

Forty-eight of the 54 included women were gonadotrophin deficient. Twenty-five (52%) women at the initiation of the study and 15 (31%) at the study end received oestrogen replacement therapy. The reason for fewer women receiving oestrogen replacement at study end compared with baseline was discontinuation because of age. The women receiving oestrogen replacement at baseline were significantly younger (mean age 44.5 (2.1) years) than the women without oestrogen therapy (mean age 57.5 (2.2) years, P<0.001). Women on oestrogen replacement therapy received a significantly higher dose of GH than women without oestrogen replacement (P<0.001). There was no difference in serum IGF1 level or IGF1 SDS between women with and without oestrogen replacement. There was also no difference in treatment response in BMD or BMC at any skeletal site measured.

Osteopenia and osteoporosis

At baseline, 37.8 and 38.8% of the patients had osteopenia (t-score < −1.0 S.D.) in the lumbar (L2–L4) spine and femur neck respectively (Table 5). After 15 years of GH replacement, 30.6% of the patients had osteopenia in the lumbar (L2–L4) spine and 40.8% in the femur neck (Table 5). The patients with remaining osteopenia at study end, as measured in the femur neck, were older (mean age 67.6 (1.7) vs 62.4 (1.5) years respectively, P<0.05) and included a larger proportion of women (61.1 vs 38.7% respectively). Among the patients with osteopenia in the lumbar (L2–L4) spine after 15 years of GH replacement, 69.0% were women compared with 37.7% women among patients without osteopenia, but there was no age difference.

Table 5 Osteopenia (t-score < −1.0 S.D.) and osteoporosis (t-score < −2.5 S.D.) at baseline and after 15 years of GH replacement therapy in 126 GH-deficient adults. Patients with an increase in t-score of >0.5 were arbitrarily considered to have an increase in t-score. Likewise, patients with a decrease in t-score of >0.5 were considered to have a decrease in t-score and patients with an increase or decrease in t-score of <0.5 were considered to have an unchanged t-score.

<table>
<thead>
<tr>
<th>Gender</th>
<th>Baseline (%)</th>
<th>After 15 years (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Osteopenia</td>
<td>Osteoporosis</td>
</tr>
<tr>
<td></td>
<td>(t-score &lt; −1.0 S.D.)</td>
<td>(t-score &lt; −2.5 S.D.)</td>
</tr>
<tr>
<td>Total body</td>
<td>23.5</td>
<td>5.1</td>
</tr>
<tr>
<td>Lumbar (L2–L4) spine</td>
<td>37.8</td>
<td>9.2</td>
</tr>
<tr>
<td>Femur neck</td>
<td>38.8</td>
<td>7.1</td>
</tr>
</tbody>
</table>

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serum IGF1 and total body BMC (a variable reflecting bone mass and density). There was no correlation between the increase in this variable after 15 years of GH replacement and femur neck BMC (r = 0.26, P < 0.01) and BMD (r = 0.30, P < 0.01).

The percent change in serum IGF1 correlated positively with the percent change in femur neck BMC (r = 0.25, P < 0.01), indicating that those patients with the highest increase in serum IGF1 had the greatest treatment response in terms of femur neck BMC. No correlation was seen at study end between the percent change in serum IGF1 and the percent change in BMC or BMD at other skeletal sites.

There was no correlation between the GH dose at study end and the percent change in any of the DXA variables measured.

Discussion

This single-centre study is the longest and one of the largest studies of the effects of GH replacement therapy in patients with AO GHD on bone mass and density. The results show that 15 years of GH replacement induced a sustained increase in total body and lumbar (L2–L4) spine BMC and BMD. At study end, all z-scores were normalised (above zero).

Total body and lumbar (L2–L4) spine BMC, BMD, t-score and z-score values were significantly above the baseline levels after 15 years of GH replacement. The main increase occurred during the first 7–10 years. The results of previous studies suggest, taken together, that GH replacement increases BMC and BMD during the first 5–10 years of therapy, whereas after that a plateau is reached in the absolute values of BMC and BMD (13, 15, 19, 20, 22). There are no previous studies over a longer period than 10 years. In this study, no further gain in absolute values of total body and lumbar (L2–L4) spine BMC and BMD occurred between 10 and 15 years of GH replacement. However, lumbar (L2–L4) spine z-score was increased at 15 years compared with the 10-year value. In the femur neck, the response to 15 years of GH replacement was different compared with the responses in total body and lumbar (L2–L4) spine. Femur neck BMC and BMD increased to a maximum after 7 years and then started to decrease. After 15 years, femur neck BMC and t-score had returned to the baseline value.

The reason why bone mass and density decreased between 10 and 15 years of GH replacement in the femur neck and not in the lumbar spine is not fully clear. Femur neck is composed of more cortical bone whereas lumbar spine is composed of more trabecular bone (27). It is well known that the trabecular bone in the lumbar spine is sensitive to sex steroids, which is noticed for instance in postmenopausal women (27). In the femur neck, with predominantly cortical bone, BMD decreases with increasing age resulting in senile osteoporosis, which affects both elderly men and women (27). It is...
therefore possible that the sex steroid replacement used in this study contributed to the increase in bone density at lumbar (L2–L4) spine whereas it had only a lesser effect on bone mass in femur neck. Furthermore, the dose of GH was gradually reduced during the study period and the possibility cannot be excluded that the dose of GH used at study end was not sufficiently high to maintain the increase in femur neck bone mass and density. In some support of this assumption, the percent change in serum IGF1 correlated positively with the percent change in femur neck BMC after 15 years, indicating that the patients with the highest increase in serum IGF1, and therefore likely the highest dose of GH, had the greatest treatment response in terms of femur neck BMC.

In all variables at all skeletal sites, except for femur neck BMC, men had a greater treatment response to GH replacement than women. Similar gender differences in responsiveness have been observed in several previous studies (13, 15, 16, 18, 19). The mechanisms underlying these gender differences are not fully understood, but sex hormones might play a role (19).

In terms of oestrogens, there was no difference in the treatment responses in BMD or BMC at any skeletal site measured between gonadotrophin-deficient women receiving or not receiving oestrogen replacement at baseline. However, a smaller number of women received oestrogen replacement at study end (n=15) than at baseline (n=25), which could have contributed to the lack of difference between women with and without oestrogen therapy. Further studies are therefore needed to clarify the importance of oestrogen replacement during long-term GH replacement. Moreover, androgens may interact with GH, resulting in increased bone mass (19). The possibility cannot be excluded that the testosterone replacement contributed to increased bone mass and density in the GHD men, although this could not be evaluated in more detail as most men were gonadotrophin-deficient and all gonadotrophin-deficient men received testosterone replacement.

A main question is whether GH replacement reduces the risk of fractures. Patients with GHD that do not receive GH replacement have an increased risk of fractures (8, 9, 10). There are some indications that GH replacement can reduce the incidence of fractures (10, 28). Although increased number of falls due to visual impairment caused by pituitary tumours or their treatment could be of importance, BMD t-score has been shown as an important predictor of fracture risk (29, 30). Therefore, an increase in bone mass and density, and especially BMD t-score, probably means that 15 years of GH replacement can reduce the fracture risk in GHD patients. Two fractures were reported in this study: one hip fracture and one symptomatic vertebral fracture. X-ray examinations were not performed to determine asymptomatic vertebral fractures. However, it has been estimated that two-thirds to three-quarters of vertebral fractures are asymptomatic and therefore remain undiagnosed (31). Patients with vertebral fractures have a mean height loss of around 5 cm (32, 33). No patient in our study had a height loss of 5 cm or more whereas six patients (two men) had a height loss of 3–4.5 cm. Therefore, we cannot exclude the possibility that a few of the GHD patients included in this study had asymptomatic vertebral fractures. Furthermore, this study is too small to evaluate fracture incidence. This would need large, probably multi-centre studies because GHD is a relatively rare condition and most centres do not have enough patients to evaluate the risk of fractures.

A limitation of the study is the lack of a control group. For ethical reasons, the study cannot be made with a control group of GHD patients without GH replacement. The use of t-scores and z-scores may to some extent compensate for the lack of a longitudinal control group. At the end of 1999, the DXA machine used was changed. The new DXA was, however, calibrated to show as similar values as possible compared with the old DXA. In terms of t-scores and z-scores, the LUNAR USA reference population was used as the reference database throughout the study period. Furthermore, the level of physical activity was not recorded. However, no effort was made to influence the patients’ physical activity level during the study period. Another limitation of the study is that the IGF1 assay was changed two times. Although the WHO NIBSC 1st IRR 87/518 standard was used for calibration throughout the study period, it cannot be excluded that the changes in IGF1 assay could have influenced the results of the IGF1 measurements. Finally, all patients were retained in the statistical analysis according to the intention-to-treat principle used. A separate analysis of the patients (n=90) who completed the 15 years of GH replacement was made and showed similar results (data not shown).

In conclusion, this study shows that 15 years of GH replacement to GHD adults induces a sustained increase in total body and lumbar (L2–L4) spine BMD and BMC. Femur neck BMC and BMD peaked after 7 years of treatment and then decreased towards baseline values, which might be due to the mean GH dose being relatively low during the last years of the study. Our study is the first that has studied the effect of 15 years of GH replacement and the results give further support to the usefulness of long-term GH replacement to GHD adults. It remains to be investigated in studies larger than the present one whether 15 years of GH replacement reduce the risk of fractures. Men had a greater treatment response than women despite women receiving a higher dose of GH. Further studies that look in more detail into the effect of sex hormones are needed to clarify the mechanisms behind these gender differences.
Declaration of interest

G Johannsson has received lecture fees from Pfizer, Novo Nordisk, Merck Serono and Eli Lilly and is a member of the SAB for KIMS Pfizer. B-A Bengtsson has previously received a research grant from Pharmacia/Pfizer. None of the other authors has any conflict of interest.

Funding

This study was supported by the Swedish Research Council, the Sahlgrenska Academy at the University of Gothenburg and Novo Nordisk.

Acknowledgements

We are indebted to Lena Wiren, Ingrid Hansson, Annika Alklind, Ann-Charlotte Olofsson, Annika Reibring and Anna Olsson at the Research Centre for Endocrinology and Metabolism for their skillful technical support.

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Received 15 December 2011
Revised version received 29 January 2012
Accepted 8 February 2012