The diagnosis of GH deficiency in obese patients: a reappraisal with GHRH plus arginine testing after pharmacological blockade of lipolysis

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Abstract

Background: The diagnosis of GH deficiency (GHD) in obese patients is complicated by the reduced GH secretion associated with overweight. A GH response to GHRH + arginine lower than 4.2 μg/l is currently considered indicative of GHD in obesity. The aim of the study was to investigate the effect of acute pharmacological blockade of lipolysis on the GH response to GHRH + arginine in obese patients.

Patients and methods: Two groups of patients were studied: 12 obese patients with proven GHD and 14 patients with essential obesity. On separate occasions, two tests were carried out in each patient: GHRH + arginine and GHRH + arginine preceded by acipimox.

Results: The mean GH peak after GHRH + arginine was significantly lower in hypopituitary patients than in subjects with essential obesity. Acipimox significantly increased the mean GH response in patients with essential obesity, but not in hypopituitary subjects. All hypopituitary patients and 7/14 patients with essential obesity displayed GH peaks lower than 4.2 μg/l after GHRH + arginine: the GH response to the test increased after acipimox pretreatment in five of these seven essentially obese subjects. After acipimox administration, free fatty acids (FFAs) significantly fell in both groups with comparable mean absolute decreases. All IGF1 values were normal in both groups of subjects.

Conclusions: Our study has demonstrated that the acipimox-induced acute reduction of circulating FFAs levels increases mean somatotropin response to GHRH + arginine in patients with essential obesity, whereas it has no effect in hypopituitary subjects. The current criterion for the diagnosis of GHD in obese patients may be misleading. Indeed, subjects affected by third degree obesity, like most of our patients, may be erroneously classified as really GH-deficient and started on an expensive unjustified treatment. It appears therefore that the current criteria for the diagnosis of GHD in obesity should be reconsidered in the light of further studies also taking into account different body mass index groups.

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Introduction

GH deficiency (GHD) syndrome in adults is characterized by abnormal body composition (visceral adiposity, reduced muscle mass, and decreased bone mineral density), atherogenic lipid profile, impaired physical performance and quality of life, and reduced life expectancy (1, 2). Such clinical and metabolic derangements improve during recombinant human GH therapy (3, 4).

Based on a clinical suspicion, GHD must be confirmed by the assessment of GH secretion performed using pharmacological challenges such as insulin tolerance test (ITT) or GHRH + arginine. The latter is currently considered the favorite diagnostic tool due to its high specificity and sensitivity, as well as tolerability (5). Adipose tissue represents a major negative determinant of somatotropin secretion (6), and indeed obesity is characterized by profound impairment of spontaneous and stimulated GH release. As a consequence, obese subjects may display GH responses to provocative tests overlapping those of patients with severe GHD (7). Thus, obesity represents the most important confounding factor in the diagnosis of GHD.

Two Italian studies (8, 9) have recently addressed this issue investigating large groups of hypopituitary patients and establishing body mass index (BMI)-related cut-off values for the somatotropin response to GHRH + arginine by percentile analysis and receiver-operating characteristic curves. In particular, GH cut-offs for the diagnosis of severe GHD in adults with BMI > 30 kg/m² were 4.2 μg/l in the experience of Corneli et al. (8) and 5.5 μg/l in the hands of Colao et al. (9) respectively. Indeed, GH response lower than 4 μg/l is considered
diagnostic for severe GHD in obese patients by the recently revised guidelines of several scientific societies (GH Research Society, European Society for Pediatric Endocrinology, Lawson Wilkins Society, European Society of Endocrinology, Japan Endocrine Society, and Endocrine Society of Australia) (10).

Several lines of experimental evidence indicate that high circulating free fatty acids (FFAs) play an important role in the impairment of spontaneous and stimulated GH secretion in obesity, given the improvement of somatotropin release observed in obese patients after the administration of the antilipolytic agent acipimox (11).

Based on the above, we performed a study aimed at investigating the influence of acute pharmacological blockade of lipolysis on the GH response to GHRH + arginine in obese patients with proven GHD and in subjects classified as having essential obesity.

Patients and methods

Patients

Two groups of patients were studied: i) 12 obese patients with proven GHD (6 men and 6 women, mean age 53.1 ± 17.36 years) due to diseases of the hypothalamic–pituitary region and/or to their treatment (see Table 1 for details); ii) 14 patients classified as having essential obesity (4 men and 10 women, mean age 49.9 ± 16.83 years). Mean BMI was significantly higher in essentially obese subjects than in GHD patients (44.2 ± 5.73 vs 38.7 ± 5.45 kg/m², P < 0.05).

Hypopituitary patients were studied before the instatement of biosynthetic GH therapy; hormone deficiencies other than GHD were treated with adequate replacement therapy, i.e. levothyroxine (Eutirox, Bracco SpA, Milan, Italy), cortisone acetate (Cortone, Istituto Chimico Internazionale Rende, Rome, Italy), and testosterone enanthate (Testo Enant, Geymonat SpA, Anagni, Italy) in men, and oestrogen–progestogen association in women, desmopressin (Minirin DDA VP, Milan, Italy), cortisone acetate (Cortone, Istituto Chimico Internazionale Rende, Rome, Italy), and testosterone enanthate (Testo Enant, Geymonat SpA, Anagni, Italy) in men, and oestrogen–progestogen association in women, desmopressin (Minirin DDA VP, Ferring), as appropriate. At the time of the study, none of the patients suffered from diseases known to affect GH–insulin-like growth factor 1 (IGF1) secretion, such as diabetes mellitus, chronic renal failure, and liver insufficiency. All patients gave their informed consent to participate in the study, which was approved by the Ethics Committee of our Institution.

Test procedures

On separate occasions and in random order, two tests were carried out for each patient: GHRH + arginine and GHRH + arginine preceded by acipimox (Olbetam 250 mg, Pfizer Italia srl), orally, at −270 and −60 min.

On both occasions, an indwelling catheter was placed into a forearm vein and continuously flushed with saline. Two baseline samples were collected prior to injection of 1 µg/kg body weight GHRH (Geref, Serono) as an i.v. bolus and infusion of 0.5 g/kg body weight (maximal dose 30 g) arginine (S.A.L.E., Turin, Italy) over 30 min through a separate i.v. access. Blood samples were then collected at 15, 30, 45, 60, and 90 min after the beginning of the infusion. Blood samples were centrifuged at 1000 g for 10 min at 4 °C; serum was collected by aspiration and stored at −20 °C until assayed for GH, IGF1, FFA, and insulin concentrations were measured in serum samples collected at baseline.

Biochemical assays

Serum GH concentrations were measured by chemiluminescent assay (CLIA, Nichols Institute Diagnostics, San Juan Capistrano, CA, USA). Intra-assay coefficients of variation (CV) for mean GH levels of 0.8, 5.9, and 17.1 µg/l were 2.8, 3.7, and 5.4% respectively. Inter-assay CV for mean GH levels of 0.1, 6.2, and 16.2 µg/l were 7.5, 6.2, and 8.7% respectively. The sensitivity of the assay is 0.02 µg/l. The assay was calibrated against the IS 98/574 recombinant standard.

IGF1 serum concentrations were determined by CLIA (Nichols Institute Diagnostics) after IGF1

Table 1 Demographics and etiology of GH deficiency (GHD) in hypopituitary patients.

<table>
<thead>
<tr>
<th>Patient</th>
<th>Gender</th>
<th>Age (years)</th>
<th>BMI (kg/m²)</th>
<th>Hypothalamic–pituitary disease</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>F</td>
<td>68</td>
<td>30.5</td>
<td>Panhypopituitarism after surgical removal of a non-functioning pituitary macroadenoma</td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>55</td>
<td>41.4</td>
<td>Panhypopituitarism after surgical removal and radiotherapy for a non-functioning pituitary macroadenoma</td>
</tr>
<tr>
<td>3</td>
<td>F</td>
<td>36</td>
<td>38.5</td>
<td>Persistent GHD after transphenoidal surgery for Cushing’s disease</td>
</tr>
<tr>
<td>4</td>
<td>F</td>
<td>63</td>
<td>33.8</td>
<td>Partial hypopituitarism in empty sella</td>
</tr>
<tr>
<td>5</td>
<td>F</td>
<td>51</td>
<td>36.1</td>
<td>Idiopathic GHD</td>
</tr>
<tr>
<td>6</td>
<td>F</td>
<td>78</td>
<td>48.0</td>
<td>Partial hypopituitarism after radiotherapy for sellar meningioma</td>
</tr>
<tr>
<td>7</td>
<td>M</td>
<td>48</td>
<td>35.3</td>
<td>Partial hypopituitarism in empty sella</td>
</tr>
<tr>
<td>8</td>
<td>M</td>
<td>27</td>
<td>32.0</td>
<td>Persistent GHD after transphenoidal surgery for Cushing’s disease</td>
</tr>
<tr>
<td>9</td>
<td>M</td>
<td>67</td>
<td>42.5</td>
<td>Partial hypopituitarism following spontaneous apoplexy of pituitary macroadenoma</td>
</tr>
<tr>
<td>10</td>
<td>M</td>
<td>75</td>
<td>46.5</td>
<td>Partial hypopituitarism following spontaneous apoplexy of pituitary macroadenoma</td>
</tr>
<tr>
<td>11</td>
<td>M</td>
<td>36</td>
<td>40.5</td>
<td>Partial hypopituitarism due to sellar germinoma</td>
</tr>
<tr>
<td>12</td>
<td>M</td>
<td>33</td>
<td>39.0</td>
<td>Panhypopituitarism after surgical removal of a craniopharyngioma</td>
</tr>
</tbody>
</table>
separation from IGF-binding proteins by sample acidification. Intra-assay CV for mean IGF1 levels of 63, 208, and 766 μg/l were 4.8, 5.2, and 4.4% respectively. Inter-assay CV for mean IGF1 levels of 62, 215, and 811 μg/l were 7.1, 5.7, and 7.4% respectively. Assay sensitivity is 6 μg/l. Data are expressed as IGF1 SDS values, calculated according to Brabant et al. IGF1 SDS was used as a variable in all statistical analyses involving IGF1. IGF1 SDS values less than −1.88 (third percentile) were considered abnormally low.

Serum FFA concentrations were determined by RANDOX colorimetric method.

Serum insulin concentrations were measured by electrochemiluminescent immunoassay (ECLIA, Roche Diagnostics GmbH). Intra- and inter-assay CV were 2.6 and 4.9% respectively. Assay sensitivity is 0.2 mU/l. The reference range is 2.6–24.9 mU/l.

**Statistical analysis**

Results are presented as mean ± s.d. Kolmogorov–Smirnov test was used to establish normality, and Student’s t-test for unpaired or paired data was used for inter- and intra-group comparisons respectively. Linear regression analysis was used to evaluate associations between variables. Statistical analyses were performed with commercially available software packages (Statview, Abacus Concepts, Berkeley, CA, USA; MedCalc Software, Mariakerke, Belgium). A P value <0.05 was considered statistically significant.

**Results**

The mean GH peak in response to GHRH+arginine was significantly lower in obese hypopituitary patients than in subjects with essential obesity (1.63 ± 1.37 vs 5.99 ± 4.38 μg/l, P < 0.01 respectively; Fig. 1). Acipimox pretreatment improved the GH response to the combined test in patients with essential obesity (mean GH peak from 5.99 ± 4.38 to 9.31 ± 3.86 μg/l, P < 0.05), but not in hypopituitary subjects (mean GH peak from 1.63 ± 1.37 to 2.20 ± 1.70 μg/l, P = NS). As a consequence, the mean absolute increase in GH peaks after acipimox was significantly higher in non-hypopituitary than in hypopituitary patients (3.33 ± 3.71 vs 0.57 ± 0.91 μg/l, P < 0.05). Notably, in all hypopituitary patients, single GH peaks were below 4.2 μg/l, the proposed cut-off value for the diagnosis of severe GHD in subjects with BMI > 30 kg/m² (8). Out of 14 patients with essential obesity, 7 patients also displayed single GH responses lower than 4.2 μg/l after GHRH + arginine (Fig. 2). In this subgroup of seven patients, acipimox premedication enhanced the response to the combined test, with a mean GH peak rising from 2.41 ± 0.93 to 7.76 ± 3.51 μg/l (P < 0.01; Fig. 3). Conversely, the mean GH peak of the seven obese patients with single GH responses > 4.2 μg/l was not significantly affected by acipimox (from 9.57 ± 3.28 to 10.87 ± 3.79 μg/l, P = NS).
When considering single subjects, acipimox pretreatment was unable to induce GH responses > 4.2 µg/l in any of the hypopituitary patients. Conversely, it caused a significant improvement in the somatotropin response of five out of the seven obese patients showing a GH peak lower than 4.2 µg/l, while in the remaining two cases, no change in GH peak was observed (Figs 2 and 4).

Baseline circulating FFAs were superimposable in patients with essential obesity and in hypopituitary subjects (0.73 ± 0.18 vs 0.84 ± 0.42 mmol/l, P = NS respectively). After acipimox administration, serum FFAs significantly fell in both groups (from 0.73 ± 0.18 to 0.16 ± 0.17 mmol/l, P < 0.001, in essential obesity, and from 0.84 ± 0.42 to 0.38 ± 0.30 mmol/l, P < 0.01, in hypopituitarism) with comparable mean absolute decreases (0.59 ± 0.20 vs 0.46 ± 0.24 mmol/l, P = NS respectively). When considering the two subgroups of patients with essential obesity displaying GH responses to GHRH+arginine respectively lower and > 4.2 µg/l, FFA values were superimposable under basal conditions and fell in a comparable manner after acipimox. In these two subsets of patients, BMI and serum insulin were also superimposable.

All IGF1 SDS values were normal in both groups of subjects. In particular, mean IGF1 SDS was superimposable in non-hypopituitary and hypopituitary patients (−0.66 ± 0.81 vs −0.63 ± 0.84, P = NS). Furthermore, in patients with essential obesity, mean IGF1 SDS values were comparable in subjects showing GH responses respectively greater and lower than 4.2 µg/l (−0.5 ± 0.8 vs −0.7 ± 0.7, P = NS).

No significant correlations were found between BMI values and GH peaks after both tests in patients with essential obesity and in hypopituitary subjects.

**Discussion**

Our study has demonstrated that the acute reduction of circulating FFA levels induced by acipimox increases the mean somatotropin response to GHRH+arginine in patients with essential obesity, whereas it has no effect in subjects with GHD. More important, in a relevant percentage of patients with essential obesity, the GH response to the combined test turned out to be lower than 4.2 µg/l, i.e. the cut-off proposed by Corneli et al. (8) for the diagnosis of severe GHD in subjects with BMI > 30 kg/m² and accepted by several scientific societies (10). Interestingly, acipimox was particularly able to improve mean GHRH+arginine-induced somatotropin release in this peculiar subgroup of obese patients, while it was ineffective in those displaying better responses to the combined test. These results may appear at variance with those observed by Maccario et al. (13) in only six obese
patients who, like seven of our patients, already displayed a significant GH rise after GHRH+arginine administration without acipimox. It appears from both our present experience and the literature (13) that in obese patients showing adequate, possibly maximal, GH rises after GHRH+arginine, acipimox premedication does not further improve these responses. We had the opportunity to compare this subgroup of obese patients with the subset comprising those characterized by the acipimox-induced improvement of an originally insufficient GH response to GHRH+arginine. The two subgroups displayed superimposable BMI values, as well as serum levels of FFAs and insulin; they also presented comparable FFA decreases after acipimox.

Considering that this antilipolytic drug is known to strongly reduce serum insulin in obese patients (14) and that hyperinsulinemia might be involved in the hyposomatotropism of obesity (11), different reductions of insulin levels in our two subsets of patients might explain different GH behaviors. Unfortunately, we lack a sufficient number of insulin determinations following acipimox administration to establish comparisons with baseline in both groups.

In our hands, although with the limitations due to the relatively small number of patients studied, the GHRH+arginine test displayed, with a cut-off value of 4.2 μg/l, a 100% sensitivity, while its specificity rose from 50 to 85.7% after acipimox premedication. However, the possibility that the two obese patients who failed to increase GH levels above 4.2 μg/l in the GHRH+arginine+acipimox test were really GH-deficient cannot be excluded, and this would clearly improve the specificity of the test.

The diagnosis of GHD in obese patients is complicated by the markedly reduced spontaneous and stimulated GH secretion that characterizes this clinical condition, and indeed over weight is the main confounding factor in the interpretation of the GH response to all provocative tests.

In obesity, a reduction in the half-life of GH as well as a significant decrease in the synthesis and secretion of the hormone has been reported (11, 15) with the net result of low plasma GH levels. The pathophysiology of this hyposomatotropism involves neuroendocrine and metabolic alterations. Among the former, dysregulation of GHRH, somatostatin, and ghrelin pathways has been demonstrated; among the latter, hyperinsulinemia and excess of circulating FFAs seem to play a major role.

The role of elevated FFAs in the GH hyposecretion of obesity has been investigated using the antilipolytic drug acipimox, whose acute and chronic administration effectively reduces circulating FFA levels. The improvement in the somatotropin response to GHRH+pyridostigmine observed by Lee et al. (16) in obese subjects pretreated with acipimox suggested that elevated FFA levels and somatostatinergic hypertonie might play a leading role.

In the experience of Cordido et al. (17), although unable to modify basal GH secretion, acipimox improved the hormonal responsiveness to pyridostigmine, GHRH, and GHRH+GHRP6, with a synergistic effect, confirming the role played by high FFAs. Comparable results were obtained by Pontiroli et al. (18) in a similar experimental setting. Prolonged acipimox administration, leading to sustained FFA suppression, has proved even more effective than the acute treatment in improving GH responses to pharmacological challenges (13, 19).

A few studies (13, 20) have evaluated the usefulness of combined administration of acipimox and GHRH for the diagnosis of GHD in obese adults, using the pharmacological blockade of lipolysis to refine the differential diagnosis between true GHD and the reversible somatotropin deficit secondary to obesity. However, according to the findings of Cordido et al. (21), GHRH+GHRP6 was superior to GHRH+acipimox in distinguishing the two situations.

None of these studies investigated the association of acipimox with GHRH+arginine, now considered the best alternative to ITT (the gold standard for the diagnosis of GHD in adults, but limited by contraindications) thanks to its reproducibility, reliability, safety profile, high sensitivity, and specificity. Moreover, GHRH+arginine is the only test for which BMI-dependent variability of GH responsiveness has been investigated (8, 9).

In our experience, half of the patients with essential obesity displayed a somatotropin response to the combined test lower than 4.2 μg/l; thus, they would have been considered as affected by true GHD according to the current guidelines (6). On the contrary, the ability of acipimox to improve GH responsiveness in most of these patients (5/7) excluded the diagnosis of hypopituitarism. The two patients whose GH response was not improved by the antilipolytic drug were investigated for possible undiagnosed hypopituitarism. However, the remaining pituitary function was normal, and the sellar magnetic resonance imaging was negative in both cases. Furthermore, the acipimox-induced decrease in FFA levels was comparable with that observed in the other patients. Thus, the clinical significance of this finding in these two patients remains puzzling, and a replacement therapy with biosynthetic GH has to be taken into account.

In keeping with the limited number of patients examined, this study was not designed to define cut-off values for the somatotropin response to GHRH+arginine+acipimox. On the other hand, our observations suggest that the current criterion for the diagnosis of GHD in obese patients, i.e. a GH peak lower than 4.2 μg/l in response to GHRH+arginine, may be misleading. Indeed, subjects affected by third degree obesity (BMI > 40 kg/m²), like most of our patients, may be erroneously classified as really GH-deficient and started on an expensive unjustified
treatment. Therefore, it appears that the current criteria for the diagnosis of GHD in obesity should be reconsidered in the light of further studies also taking into account different BMI groups.

If acipimox use in combination with GHRH + arginine may be of help in defining GHD in selected doubtful cases, further studies on larger series of subjects are needed to allow the extensive use of this test as an additional tool in the diagnosis of GHD.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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