**CLINICAL STUDY**

**Late-night and low-dose dexamethasone-suppressed cortisol in saliva and serum for the diagnosis of cortisol-secreting adrenal adenomas**

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**Abstract**

**Objective:** In patients with adrenal incidentalomas, hormonally active masses need to be considered, particularly cortisol-producing adenomas (CPA), aldosterone-producing adenomas, and pheochromocytomas. The screening for hypercortisolism relies on confirming excess cortisol secretion and insufficient suppression after dexamethasone. Because of its high correlation with free cortisol and its stress-free collection, salivary cortisol (SaC) may offer advantages over serum cortisol (SeC). We evaluated the value of SaC and SeC for the diagnosis of CPA.

**Design:** Comparative study between 2001 and 2006.

**Methods:** Thirty-eight patients with confirmed CPA were compared with 18 healthy subjects as well as 48 control patients suffering from aldosterone-producing adenomas (n=13), pheochromocytomas (n=16), or nonfunctioning adenomas (n=19). Sampling of saliva and serum was performed at 2300 and at 0800 h following low-dose dexamethasone suppression. Receiver operating characteristics analysis was used to calculate thresholds with at least 95% sensitivity for CPA.

**Results:** Regarding the cutoffs for late-night cortisol, SaC (4.8 nmol/l, sensitivity 97%, specificity 69%) was slightly more specific than SeC (115 nmol/l, sensitivity 97%, specificity 63%). In contrast, the cutoff for dexamethasone-suppressed SaC (3.7 nmol/l, sensitivity 97%, specificity 83%) was slightly less specific than SeC (94 nmol/l, sensitivity 97%, specificity 88%). However, the latter cutoffs demonstrated greater specificity when compared with the cutoffs for late-night cortisol.

**Conclusion:** The diagnostic accuracy of SaC is as good as SeC. Owing to its higher specificity, dexamethasone-suppressed cortisol is preferable to late-night cortisol when screening for Cushing’s syndrome in patients with adrenal incidentalomas.

**European Journal of Endocrinology** 161 747–753

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**Introduction**

The overall frequency of incidentally recognized adrenal masses can be as high as 6%, as shown in a report on 19 studies summarizing 86 742 autopsies (1). Incidentalomas of the adrenal gland are detected with increased frequency, mainly due to the widespread use as well as the technical improvement of imaging techniques like computed tomography (CT), magnetic resonance imaging, and ultrasonography. For example, unexpected adrenal nodules are encountered in about 5% of abdominal CT scans performed for whatever reason (2, 3). The vast majority of these incidentalomas are either nonfunctioning adrenal adenomas (2, 4, 5) or adenomas, which are clinically and/or biochemically silent (‘subclinical’). With respect to the hormonally active tumors, biochemical evaluation is needed to differentiate between cortisol-producing adenomas (CPA), aldosterone-producing adenomas, and pheochromocytomas. The biochemical screening for CPA relies on confirming hypercortisolism, and three tests are commonly used for diagnostic assessment: measurement of late-night serum or salivary cortisol (SaC), dexamethasone suppression test, and 24-h urinary free cortisol (UFC) (6, 7).

Although routinely performed in endocrine practice, both blood and urine sampling are not only stressful, but also labor intensive and time consuming. In contrast, the measurement of SaC offers several advantages, such as its simple, cost-effective and stress-free collection (8). Thus, late-night SaC has been frequently used for diagnosing Cushing’s syndrome (CS) (9–16). Most studies included few patients with primary adrenal disease, and few data are available for the use of SaC during a dexamethasone suppression test. As a consequence, this prospective study was designed to evaluate the validity of SaC measurement for the diagnosis of CS in patients with clinically and...
biochemically confirmed CPA. Samples were collected both late at night and at 0800 h following an overnight low-dose dexamethasone suppression test (LDDST). Thresholds with at least 95% sensitivity for CS were calculated for each test, using receiver operating characteristics (ROC) curves.

**Subjects and methods**

**Subjects**

Eighty-six patients with adrenal masses were prospectively enrolled at the time when they were referred for minimally invasive surgery from various endocrine centers in Germany. The 38 CPA patients presented with some clinical signs of CS. Their initial biochemical diagnosis in the primary endocrine center was made with elevated 24-h UFC, insufficient cortisol suppression after dexamethasone, and low ACTH levels (individual results are not provided because UFC reference ranges, criteria for dexamethasone suppressive tests, and laboratory procedures varied among the different primary endocrine centers). The 48 control patients (AdTu) were initially investigated in the primary endocrine center either because of incidentally discovered adrenal masses or clinical findings suggestive of an adrenal tumor (e.g. low potassium concentrations or uncontrolled hypertension). Nineteen of these patients were found to have nonfunctioning adenomas, 16 patients had histologically confirmed pheochromocytomas, and 13 patients with elevated plasma aldosterone concentration to plasma renin activity ratios as well as positive suppression tests, and/or typical clinical symptoms were diagnosed as having aldosterone-producing adenomas. Of note, careful diagnostic evaluation did not reveal any biochemically and clinically apparent features of subclinical CS coexisting with the underlying disease. All patients were reevaluated before surgical treatment: none of the females were on contraceptives or estrogens, and medications known to affect glucocorticoid metabolism were omitted for at least 24 h before testing. Moreover, none of the patients had a history of alcohol abuse or psychiatric problems. The control group consisted of 18 healthy subjects. None of these subjects suffered from endocrine disorders or was on medication (including systemic corticosteroids and hormonal contraceptives). The local ethics committee approved the study protocol, and all participants gave their written informed consent.

**Collection of samples**

While patients were always tested during a hospital stay, samples from healthy control subjects were taken in an outpatient setting. An indwelling i.v. canulla was inserted into a forearm vein, followed by a recovery period of 15 min in order to avoid stress-induced cortisol raises. Blood samples for ACTH measurement were obtained at 0800 h from all patients. Patients as well as healthy controls underwent simultaneous sampling of saliva and serum at 2300 h. Afterwards, 1 mg dexamethasone was administered orally, and matched saliva and serum samples were taken at 0800 h the next morning. Saliva was collected using the Salivette device (Sarstedt, Germany). If measurement was not performed after testing, samples were stored at −20°C.

**Measurement of samples**

Plasma ACTH was determined by solid-phase two-site sequential chemiluminescent immunometric assays (Immulite 2000, Siemens, Eschborn, Germany). SaC was assayed using a modification of the ‘GammaCoat’ RIA (DiaSorin, Stillwater, MN, USA), decreasing the sample volume from 200 to 100 μl. The lower detection limit of this assay was 0.6 nmol/l, and the intra- and interassay coefficients of variation (CV) were 2.6 and 4.6% respectively. Serum cortisol (SeC) was measured by a competitive immunoassay (Advia Centaur, Bayer, Fernwald, Germany), with a lower detection limit of 5.5 nmol/l. The intra- and interassay CV were <3.8 and 5.5% respectively, as determined at various cortisol concentrations (ranging from 107 to 1025 nmol/l). The cortisol antiseraum of this kit had a 0.2% cross-reactivity with dexamethasone (as described by the manufacturer). All hormonal measurements were performed in a single laboratory.

**Statistical analysis**

Results are expressed as the mean ± S.E.M. The diagnostic accuracy of each test was investigated using ROC curves, comparing both patient groups (CPA versus AdTu). Kruskal–Wallis tests, followed by Dunn’s multiple comparison tests, and Mann–Whitney tests were performed where appropriate. Significance was set at P < 0.05. Statistical analysis was performed using GraphPad Prism 5.0 software (GraphPad Software Inc., San Diego, CA, USA).

**Results**

The clinical characteristics of the three study groups are shown in Table 1. Patients of both groups (CPA and AdTu) demonstrated comparable age, body mass index, and tumor size, whereas sex distribution was significantly different (P < 0.05). All patients of the CPA group had plasma ACTH levels of <10 pg/ml.

**Cortisol levels at 2300 h**

Mean cortisol was highest in CPA patients, with individual levels ranging from 1.1 to 48.1 nmol/l SaC.
and from 108 to 845 nmol/l SeC respectively (Fig. 1). The AdTu patients tended to have the lowest individual values, although the ranges overlapped those seen in healthy controls (2.0–16.2 vs 2.0–18.7 nmol/l for SaC; 11–556 vs 42–552 nmol/l for SeC; Fig. 1). Mean salivary and SeC levels are listed in Table 1. Highly significant differences were found between the CPA and the AdTu group ($P < 0.001$ for salivary as well as SeC), and between the CPA group and the controls ($P < 0.001$ for SaC; $P < 0.01$ for SeC). The SeC levels found in the AdTu and the control group were slightly different ($P < 0.05$), whereas the SaC values were comparable (n.s). In general, salivary and SeC levels were highly correlated ($r = 0.77$, $P < 0.0001$). ROC analysis cutoffs with a high sensitivity for the detection of CS patients of more than 95% were 4.8 nmol/l for SaC (specificity 69%) and 115 nmol/l for SeC (specificity 63%) respectively (Table 2).

**Cortisol levels at 0800 h after 1 mg dexamethasone**

Again, mean salivary and SeC were highest in CPA patients. Individual SaC levels ranged from 2.7 to 49.3 nmol/l, whereas SeC ranged from 80 to 845 nmol/l (Fig. 2). With respect to the AdTu group, salivary and SeC ranges were 0.8–10.0 and 6–138 nmol/l respectively, while ranges of 0.9–2.7 and 13–71 nmol/l were detected in healthy controls (Fig. 2). All mean levels are summarized in Table 1. Cortisol values in the three study groups differed significantly: CPA versus AdTu, each $P < 0.001$ for salivary and serum; CPA versus controls, each $P < 0.001$ for salivary and serum. In contrast, the differences between the AdTu and the control group were not statistically significant. Salivary and SeC levels were again highly correlated ($r = 0.81$, $P < 0.0001$).

**Table 1**

<table>
<thead>
<tr>
<th></th>
<th>CPA</th>
<th>Adrenal Tumors</th>
<th>Healthy Controls</th>
</tr>
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<tbody>
<tr>
<td>Subjects (n)</td>
<td>38</td>
<td>48</td>
<td>18</td>
</tr>
<tr>
<td>Sex</td>
<td>6 M, 32 F</td>
<td>20 M, 28 F</td>
<td>9 M, 9 F</td>
</tr>
<tr>
<td>Age (years)</td>
<td>50.2±2.1</td>
<td>50.4±2.1</td>
<td>37.1±1.9</td>
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<tr>
<td>BMI (kg/m²)</td>
<td>27.6±0.8</td>
<td>27.1±0.7</td>
<td>25.5±1.1</td>
</tr>
<tr>
<td>Tumor size (cm)</td>
<td>3.09±0.13</td>
<td>3.42±0.36</td>
<td>–</td>
</tr>
<tr>
<td>Salivary cortisol (nmol/l)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2300 h</td>
<td>19.6±1.9</td>
<td>4.7±0.4</td>
<td>6.8±1.2</td>
</tr>
<tr>
<td>0800 h (after Dex)</td>
<td>19.7±1.8</td>
<td>2.8±0.2</td>
<td>1.8±0.1</td>
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<tr>
<td>Serum cortisol (nmol/l)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>2300 h</td>
<td>441±31</td>
<td>115±15</td>
<td>210±37</td>
</tr>
<tr>
<td>0800 h (after Dex)</td>
<td>432±30</td>
<td>44±5</td>
<td>27±3</td>
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</table>

**Figure 1**

Individual salivary (A) and serum cortisol levels (B) at 2300 h in patients with adrenal masses and healthy control subjects. The study groups are patients with cortisol-producing adenomas (CPA), patients with aldosterone-producing adenomas (APA), patients with nonfunctioning adenomas (NFA), patients with pheochromocytomas (PHEO), and healthy controls (C).

ROC cutoffs affording high sensitivity (> 95%) for CS were 3.7 nmol/l for SaC (specificity 83%) and 94 nmol/l for SeC (specificity 88%) (Table 2).

**Combination of midnight and dexamethasone-suppressed cortisol**

In addition to the isolated analysis of each of the four single parameters, we also investigated the combined use of midnight and dexamethasone-suppressed cortisol, applying the thresholds that had been identified before. The combined analyses of either serum or saliva were found to have slightly higher sensitivity (both 100%), but lower specificity (serum 56%, saliva 65%) than the single parameters.

**Discussion**

Several procedures are recommended for the initial screening for hypercortisolism, such as repeated 24-h UFC determination, low-dose dexamethasone suppression, and measurement of late-night cortisol (6, 7).
The finding of increased UFC levels and decreased ACTH concentrations is usually sufficient to establish the diagnosis of hypercortisolism in patients with clinical signs of CS referred for an adrenal tumor. However, adequate urine collection is difficult, especially in children and the elderly. Moreover, it may require several collections to rule out intermittent cortisol excess, which may be inconvenient for patients. Finally, measurement of UFC by immunoassay is often influenced by cross-reactivity with cortisol metabolites or exogenous glucocorticoids, and normal ranges are not very well established for some of the newer immunoassays. Consequently, ACTH measurements combined with late-night and/or dexamethasone-suppressed cortisol levels are potential alternatives when screening for CS.

Our late-night SeC cutoff of 115 nmol/l falls within the range provided by previous studies. While some authors showed that thresholds between 207 (17) and 229 nmol/l (18) had both high sensitivity and specificity, Newell-Price et al. (19) suggested a single midnight sleeping SeC cutoff of 50 nmol/l. This test, however, required inpatient admission for at least 48 h to avoid false-positive results due to the stress of hospitalization, and blood had to be drawn within the first minutes of waking the patient. Our observations are comparable with findings by Görges et al. (20) who found a sensitivity of 100% and a specificity of 77% for a cutoff of 140 nmol/l. Of note, we observed some unusually high cortisol levels at 2300 h in both the AdTu and the control group. This may point to potential influences of stress, which are more likely to occur in random samples than in samples collected during dynamic suppression tests (with defined sampling conditions).

In comparison with blood and urine sampling, the noninvasive collection of saliva is stress-free and much easier to perform in an outpatient setting (8). SaC is not influenced by cortisol-binding globulin levels, which are often altered by severe diseases or certain drugs (21–23). Therefore, SaC represents an excellent index of the free and biologically active rather than the protein-bound hormone (24). Its concentrations are independent of salivary flow, storage conditions, dental care, and food respectively (25–28). Furthermore, it has been shown that the impact of blood leakage due to microinjury on the oral mucosa is negligible (29). In this context, late-night SaC measurement has been repeatedly described as a convenient and reliable screening test for CS (10, 12–16). These studies reported sensitivities and specificities from 92 to 100% and from 93 to 100% respectively with thresholds varying from 3.6 to 15.2 nmol/l. Besides, a recent publication by Nunes et al. (30) demonstrated that late-night SaC is of great diagnostic value when screening for CS. This was true regardless of whether patients were investigated on an inpatient or an outpatient basis. However, most papers published to date included only small groups with CPA. With respect to our prospective evaluation, a cutoff of 4.8 nmol/l had similar sensitivity, but lower specificity in patients with adrenal masses.

Table 2

<table>
<thead>
<tr>
<th>Cutoff (nmol/l)</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>AUC</th>
<th>Significance (P)</th>
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<tbody>
<tr>
<td>2300 h Serum</td>
<td>115</td>
<td>97</td>
<td>63</td>
<td>0.94</td>
</tr>
<tr>
<td>0800 Serum (after Dex)</td>
<td>94</td>
<td>97</td>
<td>88</td>
<td>0.99</td>
</tr>
<tr>
<td>2300 h Saliva</td>
<td>4.8</td>
<td>97</td>
<td>69</td>
<td>0.93</td>
</tr>
<tr>
<td>0800 h Saliva (after Dex)</td>
<td>3.7</td>
<td>97</td>
<td>83</td>
<td>0.97</td>
</tr>
</tbody>
</table>

Figure 2

Individual salivary (A) and serum cortisol levels (B) at 0800 h (after 1 mg dexamethasone) in patients with adrenal masses and healthy control subjects. The study groups are patients with cortisol-producing adenomas (CPA), patients with aldosterone-producing adenomas (APA), patients with nonfunctioning adenomas (NFA), patients with pheochromocytomas (PHEO), and healthy controls (C).
Several studies found dexamethasone tests to be the most appropriate screening tools for CS due to CPA (2, 5, 31–34). Various doses have been used, but most groups preferred the LDDST with 1 mg dexamethasone, which has also been recommended by the National Institutes of Health Consensus Development Program (35). The optimal threshold is still debated, but reported cutoffs for SeC suppression usually range from 100 to 200 nmol/l (36–38). More recently, some experts suggested lower SeC cutoffs for the LDDST, mainly with the intention to raise sensitivity (6, 7, 38–40). In our prospective analysis of patients with adrenal masses, however, a threshold of 50 nmol/l was found to have only slightly higher sensitivity, but resulted in profound loss of specificity.

In contrast to late-night SaC, less is known about SaC determination after low-dose dexamethasone suppression. Up to now, only few groups investigated the validity of this method, providing cutoffs between 1.5 and 2.8 nmol/l (11, 16, 41, 42). Of note, these studies have mainly focused on patients with Cushing’s disease, while only few subjects with CPA were included. To the best of our knowledge, this is the first study in which cutoffs were calculated by comparison with a group of patients with adrenal masses of other etiologies. In contrast, comparison with healthy controls may give misleading results, especially since screening for hypercortisolism is usually initiated when clinical suspicion is high. In our study, the diagnostic performance of the LDDST was somewhat superior to late-night cortisol measurement, mainly because of its higher specificity. Although SeC showed slightly better accuracy, the difference to SaC was negligible. The combination of midnight and dexamethasone-suppressed cortisol in either serum or saliva yield improved sensitivity to 100%, whereas specificity decreased. As a consequence, we regard the determination of SaC during the LDDST as first-line test for evaluating patients with adrenal incidentalomas. Performed in an outpatient setting, this test is a simple, convenient, and economical screening tool.

It is important to keep in mind that there are also some disadvantages to the LDDST. For instance, Crapo et al. (43) reported that the rate of false-positive results is considerably higher in the presence of obesity, psychiatric disorders, chronic illness, and certain drugs respectively. In these cases, further confirmatory testing may be required. Besides, although we did not observe this phenomenon in our recent study, some CS patients retain cortisol suppression to dexamethasone (20, 40, 44). This unusual sensitivity may be due to individual differences in the absorption and metabolism of dexamethasone. For instance, patients with liver and/or adrenal failure may suffer from impaired glucocorticoid clearance, whereas drugs as well as alcohol may induce its hepatic metabolism (7). Some experts suggested simultaneous measurement of dexamethasone levels in order to minimize the number of false-positive or false-negative tests, but this approach may not be feasible because of the limited availability and the relatively high costs of dexamethasone assays. Consequently, if clinical suspicion of CS remains high, other diagnostic procedures may be more helpful.

It also has to be kept in mind that patients with only mild or even subclinical CS are difficult to detect, since it was noted that none of the common testing procedures achieved sensitivity > 80% (4). Therefore, the gold standard for establishing the diagnosis of milder forms of CS remains uncertain and is a subject of ongoing debate (5, 45). It is unclear whether an early diagnosis made on low cutoffs is helpful, considering that further diagnostic evaluation is needed in a large number of subjects with false-positive screening results. Apart from that, even if diagnosis is confirmed, its adequate therapy may still be uncertain. In this context, follow-up studies that address the outcome after adrenalectomy in comparison with the preoperative biochemical presentation would be of great interest.

With respect to the large variability of cutoffs provided for SaC, the choice of assay should be considered carefully. When testing the same saliva samples in two different assays, both Raff et al. (46) and Papanicolaou et al. (12) observed a wide variability in the absolute cortisol concentrations. Accordingly, determination of SaC requires careful evaluation of the specific procedure applied. This is especially true since its concentrations are at far lower levels than in serum, being close to the functional limit of detection of most assays.

In conclusion, the diagnostic accuracy of SaC is at least as good as SeC. This was true regardless of whether samples were collected at 2300 h or at 0800 h after 1 mg dexamethasone. However, owing to its higher specificity, dexamethasone-suppressed cortisol may be preferred to late-night cortisol. In comparison with their diagnostic accuracies in Cushing’s disease, both tests have lower specificity for differentiating cortisol-secreting adenomas from other adrenal tumors, emphasizing the need for additional tests to confirm the diagnosis.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding

This research did not receive any specific grant from any funding agency in the public, commercial, or not-for-profit sector.

Acknowledgements

We wish to thank Bianca Ueberberg, Ina Binse, and Ricarda Althoff for their support of this work.
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Received 4 August 2009
Accepted 11 August 2009