The syndrome of gastric carcinoid and hyperparathyroidism: a family study and literature review

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Abstract

Objective: To present evidence supporting the hypothesis that the coexistence of gastric carcinoids (GCs) and hyperparathyroidism may represent a distinct clinical entity, not related to multiple endocrine neoplasia type 1 (MEN1).

Methods: We studied a cohort of five young siblings (age range 26–42 years), one of whom had been found to have GC and hyperparathyroidism. All siblings underwent serial gastroscopies for the assessment of gastric neuroendocrine cell proliferations over a mean follow-up period of 31.2 months. Imaging, biochemical and hormonal as well as molecular genetic investigations were performed in the direction of MEN1 syndrome. The literature was searched for cases with coexistence of GCs and hyperparathyroidism not associated with MEN1.

Results: Four of the siblings, all male, were found to have GCs in a background of Helicobacter pylori-associated chronic atrophic gastritis and pernicious anaemia, with no serological evidence of gastric autoimmunity. In two of them, asymptomatic hyperparathyroidism was also present. Screening for MEN1 gene mutations or large deletions was negative, and hormone and imaging investigations did not support a diagnosis of familial MEN1 syndrome. A literature search revealed sporadic reports of cases with GC and hyperparathyroidism not attributable to MEN1.

Conclusions: The association of GCs and hyperparathyroidism appears to constitute a distinct syndrome that can be encountered in genetically predisposed individuals, and should not be regarded as ‘atypical’ or ‘incomplete’ expression of MEN1. Its prevalence and aetiology should be the subject of future studies. Screening for hyperparathyroidism seems to be justified in patients with GC of any type.

Introduction

In recent years, well-differentiated gastric neuroendocrine tumors (carcinoids) have been diagnosed with increasing frequency as a result of the widespread availability of upper gastrointestinal endoscopy and the development of novel immunohistochemical protocols for the recognition of endocrine cells in gastric biopsies (1). Type I gastric carcinoids (GCs), seen in the setting of hypergastrinaemia associated with chronic atrophic gastritis (CAG), are the most common (~80%) gastric neuroendocrine tumors. They originate from the enterochromaffin-like (ECL) cells of the gastric body and fundus and their pathogenesis follows the sequence hyperplasia–dysplasia–neoplasia, in which gastrin plays a central role as trophic stimulus for ECL cell growth. A similar pathogenetic mechanism is thought to be operating in the rare (~5%) type II GCs that are ECL tumors associated with gastrinoma in the setting of multiple endocrine neoplasia type 1 syndrome (MEN1). The remaining (type III or ‘sporadic’) GCs are not associated with hypergastrinaemia and can originate from any endocrine cell of the gastric wall.

There have been sporadic reports of GCs coexisting with hyperparathyroidism (HPTH), where a diagnosis of MEN1 could not be substantiated (2–7). Although the possibility of ‘atypical’ or ‘incomplete’ MEN1 could not be theoretically excluded, the question was raised if such cases might represent a new association (3, 5). In the present study, we describe multiple cases of type I GCs in a family of five siblings with CAG, two of whom were also found to have HPTH. By contrast to type II GCs, multiple cases of which can occur in MEN1 families, familial occurrence of type I GCs appears to be exceedingly rare (8, 9). These cases evoke hypotheses to explain the association between gastric ECL cell and parathyroid proliferations.
Subjects and methods

Patients

The index patient is the eldest of five siblings (four male, one female, age range 26–42 years). He was found to have a GC at age 38, when an upper gastrointestinal tract (GI) endoscopy was performed for investigation of persistent dyspepsia. He gave a history of a younger brother with dyspepsia and ‘stomach polyps’ discovered endoscopically at age 31. Personal and family history was otherwise unremarkable. Routine biochemical investigations of the index patient revealed hypercalcaemia, hypophosphataemia and high plasma parathormone (PTH) levels, leading to a diagnosis of HPTH. In order to establish or exclude a diagnosis of MEN1 syndrome, all siblings gave informed consent to undergo endoscopic, imaging, hormone and genetic investigations.

Upper GI endoscopy

The siblings underwent upper GI endoscopy and all polypoid lesions detected were removed and examined histologically. Multiple biopsies from adjacent mucosa of the gastric fundus, corpus and antrum were also taken. Patients found to have ECL neoplasia or dysplasia were placed on a 6 monthly surveillance program with endoscopic removal of all subsequently appearing lesions.

Histology and immunohistochemistry

Gastric endocrine cell lesions were classified as: a) hyperplasia (simple, linear, micronodular, adenomatoid), b) dysplasia (enlarging or fusing micronodules, microinvasion, nodular growth) and c) neoplasia (carcinoid), according to the criteria of Solcia et al. (10). Despite its limitations, especially with regard to the definition of dysplasia (11), this classification offers clinical relevance without the need for expensive, cumbersome morphometric evaluations.

Immunohistochemical staining was carried out on formalin-fixed, paraffin-embedded sections by the one-step HRP polymer detection system, using a commercially available kit (Envision, Dako, Copenhagen, Denmark). The primary monoclonal antibodies or polyclonal sera applied were directed against chromogranin-A (CgA), synaptophysin, serotonin, ghrelin, pancreatic polypeptide (PP), vasoactive intestinal peptide (VIP), somatostatin, gastrin, glucagon, insulin, cholecystokinin (CCK) and the proliferation-related antigen MKI67.

Imaging

Endoscopic ultrasonography (EUS) was employed for examination of the pancreas and gastric wall. The pituitary was examined with magnetic resonance imaging (MRI). Ultrasonography and/or scintigraphy with Tc99m-sestamibi were employed for imaging of parathyroid glands in patients with elevated plasma PTH levels. Bone density (femoral neck and lumbar spine) was measured with the dual-energy X-ray absorptiometry (DEXA) method. Whole-body octreoscan was performed using octreotide labelled with Indium-111.

Hormonal and other assays

Serum CgA levels were measured using a RIA kit (CISBIO, Bagnols-sur-Cèze, France). RIA kits were also used for the measurement of serum levels of gastrointestinal peptides including gastrin and glucagon (Siemens Medical Solutions Diagnostics, Tarrytown, NY, USA), somatostatin, PP, and VIP (Euro-Diagnostica, Arnhem, The Netherlands) and C-peptide (BioSource, Nivelles, Belgium), while insulin was measured by chemiluminescence using an IMMULITE 2000 Siemens analyser. Various commercially available kits were employed for measurement of prolactin, growth hormone, gonadotropins, adrenocorticotropin, PTH, calcitonin, thyroid stimulating hormone, free thyroxine, antithyroid antibodies (anti-TPO, anti-TG), anti-parietal cell antibodies (APCA), vitamin B12 and 25(OH)-vitamin D3.

Screening for MEN1 gene mutations

DNA was extracted from peripheral blood leucocytes using a commercially available kit (QIAamp DNA Blood Mini Kit, Qiagen Inc). The coding exons (2–10) and respective splice junctions of the MEN1 gene were amplified by PCR and subjected to denaturing high performance liquid chromatography as described previously (12). The method has 100% sensitivity for mutation detection compared with direct sequencing. Quantitative multiplex PCR of short fluorescent fragments (QMPSF) was employed in order to exclude the presence of large deletions of one or more exons, and the products were analysed on PAGE using an automated Applied DNA sequencer system, according to previously published procedures (13).

Results

The main findings are summarized in Table 1. All four male siblings had similar gastroscopic appearances, characterized by multiple small (<7 mm) polyps of the body and fundus in a background of pangastritis with varying degrees of mucosal atrophy, intestinal metaplasia and chronic active inflammation. Histologically, most polyps were hyperplastic/inflammatory, but a significant proportion of them contained endocrine cell proliferations ranging from linear hyperplasia to neoplasia. Hyperplasia and occasionally dysplasia of endocrine cells was present in biopsies of the adjacent gastric mucosa. Small (not exceeding 5 mm in size), multifocal, well differentiated neuroendocrine tumors
(carcinoids) were found in the four male siblings (M-42, M-38, M-37, M-28). The tumors were seated deeply in the mucosa and were infiltrating the muscularis mucosae. Tumour cells were arranged in solid nodular or pseudoglandular formations, were CgA- and synaptophysin-positive, and showed low mitotic activity (MKI67 positivity < 2%). No consistent immunoreactivity pattern for enteropancreatic peptides, serotonin and ghrelin was seen in the tumor cells. Focal hyperchromic thickening of the deep mucosa was revealed by EUS in two cases (M-42, M-38), but the hypoechogenic thickening of the deep mucosa was not seen in the EUS. Clinical examination for skin lesions associated with MEN1 (lipomas, angiofibromas, collagenomas) or polyglandular autoimmune syndrome (vitiligo) was also negative. The rest of the imaging investigations in the direction of MEN1 gave negative results in all siblings. In particular, no pancreatic or adrenal lesions could be detected by EUS. Screening for germline mutations of the coding region of the MEN1 gene was negative, as was the search for large deletions by QMPSF.

The average duration of follow-up was 31.2 months (range 24–36 months). All siblings received Helicobacter pylori eradication treatment, the success of which was confirmed histologically. The hypergastrinaemia persisted during the observation period in all siblings with ECL neoplasia, of whom also developed vitamin B12 deficiency requiring parenteral cobalamin administration. Elevated serum CgA levels were not

**Table 1 Main findings in individual patients.**

<table>
<thead>
<tr>
<th></th>
<th>M-42</th>
<th>F-40</th>
<th>M-38</th>
<th>M-37</th>
<th>M-28</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Endoscopic appearances</strong></td>
<td>Polyps body-fundus. Erosive gastritis antrum</td>
<td>AG body-fundus. Erythematous gastritis antrum</td>
<td>Polyps body-fundus. AG body-fundus</td>
<td>Polyps and AG body-fundus. Erosive gastritis antrum</td>
<td>Polyps body-fundus. AG fundus</td>
</tr>
<tr>
<td><strong>Most advanced ECL cell lesion detected</strong></td>
<td>Carcinoid</td>
<td>Linear hyperplasia</td>
<td>Carcinoid</td>
<td>Carcinoid</td>
<td>Carcinoid</td>
</tr>
<tr>
<td><strong>Other histologic findings</strong></td>
<td>Extensive CAG with IM</td>
<td>Chronic active gastritis with IM</td>
<td>Moderate CAG with IM, Focal active gastritis</td>
<td>Chronic active gastritis with IM</td>
<td>Extensive CAG with IM, Focal active gastritis</td>
</tr>
<tr>
<td><strong>H. pylori infection</strong></td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td><strong>Serum APCA</strong></td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
</tr>
<tr>
<td><strong>Serum gastrin (pg/ml) NR: &lt;110</strong></td>
<td>1129</td>
<td>29</td>
<td>1108</td>
<td>&gt;1263</td>
<td>&gt;1263</td>
</tr>
<tr>
<td><strong>Serum B12 (pg/ml) NR: 200–900</strong></td>
<td>89</td>
<td>486</td>
<td>173</td>
<td>151</td>
<td>99</td>
</tr>
<tr>
<td><strong>Serum CgA (ng/ml) NR: &lt;98</strong></td>
<td>211</td>
<td>48</td>
<td>123</td>
<td>101</td>
<td>145</td>
</tr>
<tr>
<td><strong>Serum Ca (NR: 8.8–10.5 mg/dl)</strong></td>
<td>10.9</td>
<td>9.3</td>
<td>9.6</td>
<td>9.8</td>
<td>9.8</td>
</tr>
<tr>
<td><strong>Serum PTH (pg/ml) NR: 15–65</strong></td>
<td>119</td>
<td>39</td>
<td>106</td>
<td>48</td>
<td>31</td>
</tr>
<tr>
<td><strong>Other abnormalities</strong></td>
<td>Simple goitre</td>
<td>Simple goitre</td>
<td>Simple goitre</td>
<td>Simple goitre</td>
<td>Simple goitre</td>
</tr>
</tbody>
</table>

M, male; F, female; AG, atrophic gastritis; CAG, chronic atrophic gastritis; IM, intestinal metaplasia; APCA, anti-parietal cell antibodies; CgA, chromogranin-A; PTH, parathyroid hormone; NR, reference range.
apparently affected by the systematic endoscopic removal of all newly appearing gastric polypoid lesions. During the same period, the HPTH remained asymptomatic in both cases.

Discussion

All five members of this unique sibship had gastric ECL cell proliferations of varying degrees, corresponding to the intensity of the associated atrophic gastritis. The latter could be aetiologically related to \textit{H. pylori} infection, which often shows familial clustering. It is now recognized that long-standing \textit{H. pylori} infection may lead to CAG and pernicious anaemia (PA) \cite{14, 15}, which should be differentiated from the classic autoimmune CAG. Moreover, there is evidence from studies in animals and humans that \textit{H. pylori} infection may also lead to development of GCs, at least in certain ethnic groups \cite{1–18}. Proposed pathogenetic mechanisms involve the tropic stimulus of hypergastrinaemia and a possible direct mitogenic effect of \textit{H. pylori} lipopolysaccharide on gastric ECL cells.

The development of HPTH in two out of four siblings with GC is unlikely to be caused by chance alone, due to the rarity of these conditions in young individuals. One has to postulate the presence of a specific genetic background predisposing to independent development of either condition, or, alternatively, a pathophysiological mechanism leading to the development of HPTH when a GC is present. The first hypothesis was adopted in some older reports of the combination of GC with HPTH, which was thought to represent an incomplete expression of MEN1 syndrome \cite{2, 4–6}. In fact, some of the reported cases had clinical features highly suggestive of MEN1 \cite{6}. After the MEN1 gene was identified in 1997, it was shown that GC and HPTH could coexist outside the MEN1 setting \cite{3}. This is further supported by our negative genetic, biochemical and imaging investigations for MEN1 syndrome in this family with GC and HPTH. Apart from MEN1, familial predisposition to HPTH may also result from germline mutations in the genes for MEN2 (\textit{RET}), familial hypocalciuric hypercalcaemia/neonatal severe HPTH (\textit{calcium-sensing receptor, CASR} gene), HPTH – jaw tumor syndrome (\textit{CDC73}), and familial isolated HPTH that has a mixed genetic basis. However, these conditions are not known to be associated with GC development. A literature search revealed five previous reports of HPTH associated with GC in patients not fulfilling the diagnostic criteria for MEN1 syndrome \cite{2–6}. Four of them occurred in patients with CAG/PA (Table 2).

It has been suggested that a mutation in exon 3 of the \textit{CTNNB1} gene, encoding \textit{\beta}-catenin, may be involved in the oncogenesis of both carcinoids \cite{19} and parathyroid tumors \cite{20} through \textit{\beta}-catenin accumulation and activation of the Wnt signalling pathway. However, the recently reported failure to detect \textit{CTNNB1} exon 3 mutations in large series of gastrointestinal (including gastric) carcinoids \cite{21} and parathyroid adenomas \cite{22} makes it unlikely that this mutation constitutes the genetic basis of ECL cell and parathyroid neoplasia in our family.

There is evidence that an unrecognized autoimmune mechanism might be participating in the pathogenesis of a proportion of cases of HPTH due to adenoma or hyperplasia \cite{23}. Moreover, it is known that \textit{H. pylori} has the ability to trigger autoimmunity, possibly through mechanisms of molecular mimicry \cite{24, 25}. Therefore, despite the absence of APCA, an autoimmune mechanism leading to the development of both CAG and HPTH cannot be entirely excluded, in the view of lack of detailed gastric and parathyroid autoantibody studies in our patients and those reported by others.

As early as 1982, Selking \textit{et al}. noted an increased prevalence of PA in 441 patients operated for primary HPTH \cite{26}, although the same authors could not find a

<table>
<thead>
<tr>
<th>Sex/age\textsuperscript{a}</th>
<th>Associated conditions</th>
<th>Presentation of HPTH</th>
<th>Parathyroid pathology</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>M/54</td>
<td>CAG/PA</td>
<td>Urolithiasis</td>
<td>Single adenoma</td>
<td>Alberti-Flor \textit{et al}. \cite{2}</td>
</tr>
<tr>
<td>F/50</td>
<td>CAG/PA</td>
<td>Asymptomatic</td>
<td>Single adenoma</td>
<td>Rode \textit{et al}. \cite{6} and Stockbrugger \textit{et al}. \cite{7}</td>
</tr>
<tr>
<td>F/50</td>
<td>CAG/PA, diabetes mellitus type \textit{i}, Graves' disease</td>
<td>Hypercalcaemia</td>
<td>Single adenoma</td>
<td>Ollenschlager \textit{et al}. \cite{5}</td>
</tr>
<tr>
<td>F/63</td>
<td>CAG/PA, Hashimoto's disease</td>
<td>Hypercalcaemia</td>
<td>Single adenoma</td>
<td>Corleto \textit{et al}. \cite{3}</td>
</tr>
<tr>
<td>M/42</td>
<td>CAG/PA, goitre</td>
<td>Asymptomatic</td>
<td>Not operated, imaging studies suggest adenoma</td>
<td>Present report</td>
</tr>
<tr>
<td>M/38</td>
<td>CAG/PA</td>
<td>Asymptomatic</td>
<td>Not operated</td>
<td>Nores \textit{et al}. \cite{4}</td>
</tr>
<tr>
<td>F/33</td>
<td>–</td>
<td>Urolithiasis</td>
<td>Single adenoma</td>
<td>Present report</td>
</tr>
</tbody>
</table>

\textsuperscript{a}At the time of diagnosis of HPTH.

\textsuperscript{b}Reports referring to the same patient.

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consistent relationship between serum gastrin levels and parathyroid hyperfunction in 32 patients with PA. Peracchi et al. reported HPTH in 3 out of 52 patients with CAG. 31 out of whom were also harbouring ECL cell proliferations (27). The fact that six out of the seven documented cases with GC and HPTH unrelated to MEN1 (Table 2) were seen in patients with hypergastrinaemia secondary to CAG evokes the question of whether gastrin could be playing a role in the pathogenesis of HPTH in these cases. A direct effect of gastrin on the parathyroids is unlikely, as gastrin receptors are typically absent from normal and neoplastic parathyroid tissue (28, 29). In chickens, hypergastrinaemia induced by administration of omeprazole or infusion of gastrin resulted in increased parathyroid gland weight and PTH gene expression (30). This is in contrast with the findings in fundectomised rats, where hypergastrinaemia had no effect on PTH (31). One would therefore be tempted to postulate that the ECL cells, absent in the case of fundectomy, are the link between gastrin and parathyroids. Histamine, the main product of ECL cells, has a known stimulatory effect on the parathyroids (32), and its serum levels can be found slightly elevated in patients with GC (8), although there is no evidence that this finding is clinically significant. It should be noted, however, that at least one other ECL cell product – basic fibroblast growth factor (NUDT6) – is known to exert a mitogenic effect on the parathyroids (33, 34). At variance with this hypothesis are the data of Gagnemo-Persson et al. (35), showing that stimulation of ECL cells by hypergastrinaemia in rats did not result in elevation of plasma PTH levels or PTH mRNA expression in the parathyroid gland.

It is theoretically conceivable that, in the setting of CAG, hyperparathyroidism might be reactive to calcitonin hypersecretion caused by stimulation of C-cells via their gastrin receptor (36). However, this mechanism is unlikely according to published data indicating that serum gastrin has no influence on calcitonin release in hypergastrinaemic patients with PA (37, 38).

Whatever the underlying mechanism, the evidence presented here points to the existence of an association between GC and HPTH, not attributable to MEN1. This association should be regarded as a syndrome seen mostly, but not exclusively in the clinical setting of CAG/PA. It is characterized by usually asymptomatic HPTH due to a parathyroid adenoma discovered incidentally during laboratory work-up of patients with GC. Future studies may establish the prevalence and elucidate the aetiology and clinical significance of this intriguing association. In the meantime, screening for HPTH seems justified in patients with GC of any type.

Declaration of interest

The authors declare no conflict of interest that could be perceived prejudicing the impartiality of this scientific work.

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