**Endothelial nitric oxide synthase intron 4a/b polymorphism and early atherosclerotic changes in hypopituitary GH-deficient adult patients**

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**Abstract**

**Objective:** Endothelial nitric oxide synthase (eNOS) intron 4a/b polymorphism is associated with plasma NO concentrations and coronary artery disease/hypertension in various populations. GH deficiency in adulthood predisposes to reduced NO concentrations and premature atherosclerosis. Our aim was to determine whether intron 4a/b polymorphism of eNOS gene influences endothelial function and early atherosclerotic changes in GH-deficient hypopituitary patients.

**Design:** Thirty-three hypopituitary GH-deficient patients on conventional replacement therapy other than GH and 43 age-, sex-, and body mass index (BMI)-matched controls were studied in this cross-sectional case–control study.

**Methods:** Early atherosclerotic changes were determined by flow-mediated dilation (FMD) of brachial artery and carotid artery intima-media thickness (IMT). eNOS4a/b polymorphism was detected by PCR.

**Results:** Hypopituitary patients had significantly higher total/low-density lipoprotein cholesterol and fat mass and lower IGF-I concentrations compared with controls. IMT was significantly higher in patients (0.777 ± 0.23 vs 0.639 ± 0.17 mm, P < 0.01). No significant difference was observed with respect to FMD measurements. eNOS4a/b genotype frequencies were similar between patients and controls. Patients carrying ‘a’ allele (a/a and a/b) had significantly higher IMT compared with controls carrying ‘a’ allele and bb genotype (P < 0.05). However, logistic regression analysis revealed that presence of hypopituitarism, age ≥45 years, and BMI ≥27.9 kg/m² were significant independent predictors of IMT ≥0.65 mm.

**Conclusion:** No compelling data are evident to suggest that eNOS4a/b polymorphism modifies the atherosclerotic process in GH-deficient situations. A large case–control study is needed to confirm our findings.

**Introduction**

Long-standing growth hormone (GH) deficiency in adulthood may predispose to the development of premature atherosclerosis (1–4). Epidemiological evidence indicated an increased incidence of cardiovascular and cerebrovascular diseases in hypopituitary patients treated with conventional replacement therapy other than GH (1, 3, 4). A causal relationship is suggested between GH deficiency and increased vascular mortality and morbidity (1, 3, 5). Several studies indicated improved atherosclerotic risk profile and vascular reactivity after GH replacement therapy in hypopituitary patients (6–9).

The endothelium modulates vascular tone by producing vasodilator and vasoconstrictor substances. Nitric oxide (NO) produced by endothelial cells modulates vascular tone and confers protection against atherosclerosis (10, 11). Endothelial dysfunction that results in reduced availability of NO is recently identified as an early marker of atherosclerosis and has value to predict future coronary artery disease before anatomical evidence of atherosclerosis appears (10, 11). Flow-mediated dilation (FMD) defined as a change in arterial diameter in response to reactive hyperemia is an endothelium-dependent process. Flow-mediated endothelium-dependent dilation evaluated noninvasively by ultrasonography has been used to determine endothelial function in medium-sized vessels and brachial artery measurements can be used as a surrogate marker for coronary endothelial function (12–14).

The endothelial cells produce NO by the enzyme NO synthase (eNOS) (10, 11). Decreased production of NO by eNOS may contribute to the endothelial dysfunction and early atherosclerotic changes. Previous studies indicated that polymorphisms of eNOS gene could lead to decreased
NO availability and play a role for the development of atherosclerotic cardiovascular disease (15, 16). A significant association between variable number of tandem repeat (VNTR) polymorphism in intron 4 of eNOS gene (eNOS4a/b polymorphism) and coronary artery disease/hypertension has been detected in various populations (17–19), although this association has not been confirmed in some studies (20–22). In addition, eNOS4a/b polymorphism has been reported to be associated with decreased plasma NO concentrations (15).

In a GH-deficient patient, endothelial NO production by eNOS may further diminish. Boger et al. (23) indicated decreased NO formation in untreated GH-deficient patient, which has been reversed after GH replacement therapy. Insulin-like growth factor-I (IGF-I) has been reported to directly simulate NO production in endothelial cells as a result of the activation of eNOS possibly by a tyrosine kinase-dependent mechanism (24).

Thickening of the intima and tunica media of the arteries is another marker of initial asymptomatic atherosclerosis. The increase in intima-media thickness (IMT) in the extracranial carotid arteries is reported to be associated with a higher prevalence of coronary heart disease and described as a surrogate marker of coronary atherosclerosis (25, 26).

The aim of this study is to determine whether the intron 4a/b polymorphism of eNOS gene modulates endothelial function and early atherosclerotic changes in GH-deficient hypopituitary patients compared with controls.

Materials and methods

Patients and controls

Thirty-three hypopituitary patients with at least three pituitary hormone deficits (adrenocorticotrophin; ACTH, thyrotrophin; TSH, and gonadotrophins) other than GH (12 males and 21 females) and forty-three healthy controls (17 males and 26 females) were studied.

Subjects affected by diabetes mellitus, hypertension or any other established cardiovascular disease, and other chronic diseases that could affect endothelial function were excluded.

The patients and controls were matched for age, sex, and body mass index (BMI; Table 1). Patients were recruited from the Endocrinology Clinic of Internal Medicine at Istanbul Faculty of Medicine. Controls were recruited from hospital staff and friends. The study was performed according to the Declaration of Helsinki and Institutional Ethical Committee approved the study. Written informed consent was obtained from all study participants (both patients and controls).

Causes of hypopituitarism were tumors of sellar and suprasellar region (macroadenomas in 3 cases, craniopharyngiomas in 3 cases, germinomas in 2 cases, hypotalamic glioma in 1 case, GH-secreting pituitary adenoma in 1 case, ACTH-secreting pituitary adenoma in 1 case, and non-functional lesions in 7 cases) treated by surgery, radiotherapy, or both in 18 patients. Sheehan’s syndrome in 7 patients, primary empty sella in 2 patients, septo-optic dysplasia in 5 patients, and Langerhans cell granulomatosis of hypothalamic–pituitary region in 1 patient. The patient with a diagnosis of GH-secreting pituitary adenoma was operated transsphenoidally 17 years before. The patient with a diagnosis of ACTH-secreting pituitary adenoma was operated transsphenoidally 12 years before. In both of these patients, panhypopituitarism developed after the operation. Mean duration of hypopituitarism was 123.06 ± 101.97 months (median: 132.00 months, range: 12–360 months). All patients were on conventional replacement therapy consisting of L-thyroxine, glucocorticoids, sex hormones (except for

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Study parameters in patients and controls.</th>
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</thead>
<tbody>
<tr>
<td><strong>Gender (male/female)</strong></td>
<td>12/21</td>
</tr>
<tr>
<td><strong>Age (years)</strong></td>
<td>44.09 ± 16.71</td>
</tr>
<tr>
<td><strong>Systolic BP</strong></td>
<td>117.72 ± 18.33</td>
</tr>
<tr>
<td><strong>Diastolic BP</strong></td>
<td>76.06 ± 12.23</td>
</tr>
<tr>
<td><strong>Glucose mmol/l</strong></td>
<td>4.50 ± 0.92</td>
</tr>
<tr>
<td><strong>Cholesterol mmol/l</strong></td>
<td>5.47 ± 0.91</td>
</tr>
<tr>
<td><strong>Triglycerides</strong></td>
<td>1.56 ± 0.84</td>
</tr>
<tr>
<td><strong>HDL cholesterol mmol/l</strong></td>
<td>1.30 ± 0.32</td>
</tr>
<tr>
<td><strong>LDL cholesterol mmol/l</strong></td>
<td>3.38 ± 0.82</td>
</tr>
<tr>
<td><strong>Free T4 pmol/l</strong></td>
<td>15.10 ± 3.69</td>
</tr>
<tr>
<td><strong>IGF-I µg/l</strong></td>
<td>59.85 ± 31.01</td>
</tr>
<tr>
<td><strong>BMI kg/m²</strong></td>
<td>28.65 ± 5.88</td>
</tr>
<tr>
<td><strong>Fat mass (%)</strong></td>
<td>43.28 ± 11.33</td>
</tr>
<tr>
<td><strong>WHR</strong></td>
<td>0.88 ± 0.07</td>
</tr>
<tr>
<td><strong>FMD (%)</strong></td>
<td>5.88 ± 3.53</td>
</tr>
<tr>
<td><strong>IMT (mm)</strong></td>
<td>0.777 ± 0.23</td>
</tr>
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</table>

BP, blood pressure; BMI, body mass index; WHR, waist-to-hip ratio; FMD, flow-mediated dilation; IMT, carotid artery intima-media thickness; IGF-I, insulin-like growth factor-I; NS, not significant.

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postmenopausal women), and desmopressin where necessary. Out of 21 and 26 women in hypopituitary and control groups, 10 were postmenopausal in each group respectively ($P>0.05$). In the control group, the postmenopausal women were not receiving sex hormone preparation and the premenopausal women had regular menstrual cycles. None of the patients received GH replacement therapy for at least 12 months before the study. All patients were stable on conventional replacement treatment for at least 6 months before the study. Replacement doses had been optimized previously for the individual patients on the basis of clinical and biochemical evaluation and target hormone concentrations. This regimen was continued in the present study. Biochemical markers of thyroid, liver, and kidney functions were within normal range in all subjects. Only 2 out of 33 hypopituitary patients and 5 out of 43 controls had positive family history of coronary heart disease ($P>0.05$).

Subjects were classified as those who had never smoked, ex-smoker, and current smokers. Number of the patients in these subgroups were as follows: patient group, 26/33, 1/33, and 6/33 respectively; control group, 30/43, 3/43, and 10/43 respectively ($P>0.05$).

GH deficiency was determined according to the following criteria: at least three pituitary hormone deficiencies together with 1) IGF-I concentrations lower than mean$–2$ s.d. of the control group ($<67$ $\mu$g/l) (27, 28) and/or 2) maximum stimulated serum GH concentrations lower than 3 $\mu$g/l in response to previously administered glucagon (29). All patients were GH deficient according to these criteria.

**Study protocol**

Patients and controls attended the Endocrinology Clinic of Internal Medicine Department after an overnight fast of 12 h. The investigation was carried out on an outpatient basis. Menstruating women (patients and controls) were studied during the early follicular phase of menstrual cycle. Venous blood was obtained for genotype determination and measurements of glucose, triglycerides, total cholesterol, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL) cholesterol, routine hematological, liver, renal function tests, free thyroxine ($T_4$), and IGF-I concentrations. Samples were stored at $−70^\circ$C until eNOS4a/b polymorphism analysis; all other parameters were determined on the same day of collection. After blood sampling, anthropometric measurements were performed. Body composition was determined by bioimpedance analysis. FMD of brachial artery and carotid artery IMT measurements were performed on the same day after anthropometric measurements were taken.

**Anthropometric and physical parameters**

BMI was calculated as the ratio of weight (kg) divided by the square of height (m$^2$). Waist-to-hip ratio (WHR) was calculated as waist circumference (cm) divided by hip circumference (cm).

Body composition was measured by a bioelectrical impedance analyser (Bodystat 1500, Bodystat Ltd, Douglas, Isle of Man, British Isles, Great Britain) after overnight fast with an empty bladder in the morning of the anthropometric measurements and blood samples were taken.

Systolic and diastolic blood pressures were measured on the right arm of the subject in an upright sitting position after at least 5 min rest using a mercury sphygmomanometer with appropriate cuff size.

**Metabolic and hormone parameters**

All biochemical analyses including glucose, total cholesterol, HDL cholesterol, LDL cholesterol, and triglycerides concentrations were performed at Roche/Hitachi Modular Analytics System in the Central Biochemistry Laboratory. Glucose was measured by an oxidase-based technique. LDL and HDL cholesterol were determined by direct method (homogeneous enzymatic assay for the direct quantitative determination of LDL and HDL cholesterols) at Roche/Hitachi Modular Analytics System.

Free $T_4$ concentrations were analyzed at this system by electrochemiluminescence immunoassay (ECLIA; E170 Module, Tokyo, Japan).

Serum IGF-I concentrations were measured on an Immulite and Immulite 1000 Analyzer using an enzyme-labeled chemiluminescent immunometric assay (EURO/DPC Ltd, Gwynedd, UK).

**Measurement of endothelial function**

Brachial artery FMD (endothelium dependent process) was performed as previously reported by an ultrasonic vessel wall-tracking system (10 MHz linear transducer attached to standard Vingmed System Five, Horten, Norway) (12). The brachial artery of the dominant arm was identified and a two-dimensional longitudinal B-mode image of the brachial artery was obtained. The brachial artery internal diameter was assessed at end diastole (timed by the QRS complex) and arterial flow volume was measured using the pulse Doppler sample volume at $\leq70^\circ$ angle in the center of the artery. Forearm ischemia was caused by inflating a pneumatic arm cuff up to 200 mmHg for 5 min. The cuff was deflated and the arterial flow volume was immediately recorded. Arterial diameter was measured at 60 s after deflation. Three consecutive measurements were taken before inflation and after deflation. Mean of these three measurements was used for calculation: (mean arterial diameter 60 s after deflation$–$(mean baseline diameter))/((mean baseline diameter) $\times$ 100).

Results were expressed as percentage changes from baseline. Subjects did not smoke or drink caffeine rich...
beverages for 12 h before testing. Intra-observer variability of FMD measurement was 3.3%.

**Ultrasound measurement of the common carotid artery**

Bilateral carotid ultrasound was carried out using an ultrasound system with a high-resolution 10 MHz linear array scan head (attached to standard Vingmed System Five). The common carotid arteries were scanned longitudinally. The bulb dilation served as a landmark to indicate the border between distal common carotid artery and the carotid bulb. Images were obtained from the distal portion of the common carotid artery. 1–2 cm proximal to the carotid bulb. Images were saved and stored on S-VHS videotape. The two bright echogenic lines in the arterial wall were identified as the intima and media lines. The first echogenic line represents the lumen–intima interface and the second echogenic line represents the media–adventitia interface. The intimal plus medial thickness was measured as the distance from the main edge of the first to main edge of the second echogenic line. Each measurement was repeated thrice, and the mean of the left and right common carotid arteries was taken and used for further analysis. None of the subjects had atheromatous plaque, localized lesion of thickness > 2.0 mm, or stenosis in this region. Intra-observer variability of IMT measurement was 3.6%.

**Genetic polymorphism analysis**

eNOS intron 4a/b polymorphism was determined by PCR using oligonucleotide primers (sense: 5'-AGGCC-TATGGTATTGCTTT-3'; antisense: 5'-TCTCTTATGGC-TGTGCTAC-3') that flank the region of the 27 bp VNTR in intron 4. Reactions were performed in a total volume of 50 µl containing 100 ng genomic DNA, 10 pmol of each primer, 0.2 mM dNTP, 1 U Taq DNA polymerase, 5 µl PCR buffer (500 mmol/l KCL, 100 mmol 3-hydroxy-methyl-aminomethane chloride and 0.8% Nonidet P40). The thermocycling procedure consisted of initial denaturation at 94 °C for 1 min, annealing at 58 °C for 1 min, and extension at 72 °C for 1 min. The PCR products were analyzed using 3% agarose gel electrophoresis 90 V for 1 h and visualized by ethidium bromide staining. The large allele eNOS4b contains five tandem 27 bp repeats, and smaller allele eNOS4a was shown in Table 2. The genotype frequencies within groups was in accordance with the distribution predicted by the Hardy–Weinberg equilibrium model. Continuous variables among the genotypes were compared with the use of ANOVA and a post hoc Tukey test. Independent sample t-test was used to compare continuous variables between patients and controls. Categorical data were compared by χ² and Fisher's exact test where appropriate. The triglyceride and systolic and diastolic blood pressure measurements significantly deviated from a normal distribution by Kolmogorov–Smirnov test. Therefore, log-transformed values were used for these parameters. Univariate and logistic regression analyses were used to evaluate the significant associations between study parameters and genotypes. All tests were two sided and a P value < 0.05 was considered as significant.

**Results**

Baseline characteristics of patients and controls were summarized in Table 1. Hypopituitary GH-deficient patients had significantly higher total and LDL cholesterol and lower IGF-I concentrations compared with controls. Percent fat mass determined by bioelectrical impedance analysis was significantly higher in patients. Lower FMD measurements were observed in patients, although difference did not reach statistical significance. IMT of common carotid arteries was significantly higher in patients compared with controls.

**Genotype frequencies**

The distribution of eNOS4a/b genotype frequencies were shown in Table 2. The genotype frequencies were in agreement with Hardy–Weinberg equilibrium. Frequencies were not significantly different between patients and controls (χ²: 3.21, P = 0.20).

Baseline characteristics were compared among genotype (a/a, a/b, and b/b) carriers in the whole group and in the patients. None of the subjects carried the a/a genotype in the control group. No significant difference was observed with respect to baseline characteristics.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>Endothelial nitric oxide synthase (eNOS) 4a/b genotype distribution in hypopituitary growth hormone (GH)-deficient patients and controls.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>eNOS4a/b</td>
</tr>
<tr>
<td>a/a n (%)</td>
<td>2 (6.1)</td>
</tr>
<tr>
<td>a/b n (%)</td>
<td>9 (27.3)</td>
</tr>
<tr>
<td>b/b n (%)</td>
<td>22 (66.6)</td>
</tr>
<tr>
<td>Total n (%)</td>
<td>33 (100)</td>
</tr>
<tr>
<td>χ²:</td>
<td>3.21, P = 0.20</td>
</tr>
</tbody>
</table>
among three genotypes in the patients and the whole group (data not shown). The patients and controls were further grouped into genotype carriers containing ‘a’ allele (a/a and a/b) and b/b. No significant difference was observed in terms of study parameters between controls carrying ‘a’ allele and b/b genotype (data not shown). In the patient group, diastolic blood pressure was the only significantly different parameter between patients carrying ‘a’ allele (a/a and a/b) and b/b genotype; being significantly higher in patients carrying eNOS ‘a’ allele (a/a and a/b) compared with patients carrying b/b genotype (82.72 ± 11.90 mmHg versus 72.72 ± 11.20 mmHg respectively, \( P = 0.029 \)).

Patients carrying ‘a’ allele (a/a and a/b) and b/b genotype and controls carrying ‘a’ allele (a/a and a/b) and b/b genotype were compared (Table 3). IMT was significantly higher in patients carrying ‘a’ allele compared with controls carrying ‘a’ allele and b/b genotype. Although patients carrying b/b genotype had significantly higher cholesterol concentrations, IMT was not significantly different between patients carrying b/b genotype and controls. No significant difference was observed with respect to FMD measurements between patients and controls carrying ‘a’ allele or b/b genotype.

**Correlations between FMD/IMT and study parameters**

In the patients, IMT was significantly correlated with age, BMI, diastolic, and systolic blood pressure. No significant association between FMD and study parameters was observed in hypopituitary patients. In the controls, IMT correlated significantly with age, BMI, WHR, and glucose. No significant association was observed between FMD and study parameters in the controls either (Table 4). Significant univariate relationships were further evaluated in a multivariate logistic regression analysis model including genotypes. Patients and controls carrying ‘a’ allele (a/a or a/b) were labeled as (1) and b/b genotype was labeled as (0) for this analysis. For continuous parameters (age, BMI, systolic and diastolic blood pressure, WHR, and glucose), median values of the parameters in the whole group were chosen as limits. Patients and controls having values greater than these limits were labeled as (1) and values less than these limits were labeled as (0).

Presence of hypopituitarism (regression coefficient = 1.622, \( P = 0.006 \)), BMI > 27.9 kg/m\(^2\) (regression coefficient = 1.669, \( P = 0.004 \)), and age ≥ 45 years (regression coefficient = 1.326, \( P = 0.019 \)) had significantly and independently predicted study subjects with an IMT ≥ 0.65 mm. Carrying an ‘a’ allele did not significantly enter in this regression model. The ratio for accurately classifying the study subjects with an IMT ≥ 0.65 mm according to this model was 72.4%.

**Discussion**

In this study, although BMI-, age-, and sex-matched controls were used, percent fat mass, total, and LDL cholesterol concentrations were significantly higher in the patients compared with controls. Our findings supported previous observations indicating increased fat mass and a more atherogenic lipid profile in GH-deficient adults (6, 8, 9, 30). Carotid artery IMT was significantly higher in our patients compared with controls (0.777 ± 0.23 mm versus 0.639 ± 0.17 mm, \( P < 0.01 \)). Increased IMT, intimal plaque formation, and reduced arterial compliance were reported previously in hypopituitary patients.

**Table 3 Baseline characteristics of hypopituitary patients and controls carrying ‘a’ allele (genotypes a/a and a/b) and bb genotype.**

<table>
<thead>
<tr>
<th>Patients carrying eNOS4a/a and a/b n: 11</th>
<th>Controls carrying eNOS4a/a and a/b n: 16</th>
<th>Patients carrying eNOS4b/b n: 22</th>
<th>Controls carrying eNOS4b/b n: 27</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender (male/female)(^a)</td>
<td>4/7</td>
<td>6/10</td>
<td>8/14</td>
</tr>
<tr>
<td>Age (years)</td>
<td>44.81 ± 18.58</td>
<td>45.25 ± 13.41</td>
<td>43.72 ± 16.15</td>
</tr>
<tr>
<td>Systolic BP(^b) mmHg</td>
<td>125.90 ± 19.65</td>
<td>119.32 ± 16.52</td>
<td>113.63 ± 16.48</td>
</tr>
<tr>
<td>Diastolic BP(^b) mmHg</td>
<td>82.72 ± 11.90</td>
<td>76.25 ± 10.87</td>
<td>72.72 ± 11.20</td>
</tr>
<tr>
<td>Glucose mmol/l</td>
<td>4.55 ± 0.47</td>
<td>4.65 ± 0.47</td>
<td>4.47 ± 1.08</td>
</tr>
<tr>
<td>Cholesterol mmol/l</td>
<td>5.15 ± 0.60</td>
<td>5.05 ± 0.97</td>
<td>5.63 ± 1.01*</td>
</tr>
<tr>
<td>Triglycerides(^c) mmol/l</td>
<td>1.54 ± 0.82</td>
<td>2.06 ± 1.56</td>
<td>1.56 ± 0.87</td>
</tr>
<tr>
<td>HDL cholesterol mmol/l</td>
<td>1.24 ± 0.35</td>
<td>1.29 ± 0.37</td>
<td>1.33 ± 0.30</td>
</tr>
<tr>
<td>LDL cholesterol mmol/l</td>
<td>3.17 ± 0.52</td>
<td>2.95 ± 0.75</td>
<td>3.49 ± 0.92</td>
</tr>
<tr>
<td>Free T(_3) pmol/l</td>
<td>15.69 ± 3.47</td>
<td>13.58 ± 1.69</td>
<td>14.80 ± 3.83</td>
</tr>
<tr>
<td>IGF-I (\mu g/l)</td>
<td>60.40 ± 32.56(^d)</td>
<td>148.97 ± 44.22</td>
<td>59.55 ± 30.99(^d)</td>
</tr>
<tr>
<td>BMI kg/m(^2)</td>
<td>30.88 ± 6.98</td>
<td>26.52 ± 3.95</td>
<td>27.53 ± 5.05</td>
</tr>
<tr>
<td>Fat mass (%)</td>
<td>45.18 ± 12.71</td>
<td>39.58 ± 11.28</td>
<td>42.34 ± 10.77</td>
</tr>
<tr>
<td>WHR</td>
<td>0.892 ± 0.07</td>
<td>0.870 ± 0.07</td>
<td>0.879 ± 0.06</td>
</tr>
<tr>
<td>FMD (%)</td>
<td>6.49 ± 4.65</td>
<td>6.83 ± 5.52</td>
<td>5.58 ± 2.91</td>
</tr>
<tr>
<td>IMT (mm)</td>
<td>0.845 ± 0.21(^e)</td>
<td>0.616 ± 0.11</td>
<td>0.742 ± 0.24</td>
</tr>
</tbody>
</table>

\(^a\)Values were mean ± s.d. \(^b\)BP, blood pressure; BMI, body mass index; WHR, waist-to-hip ratio; FMD, flow mediated-dilation; IMT, carotid artery intima-media thickness; IGF-I, insulin-like growth factor-1. \(^P < 0.006\) compared with b/b genotype controls. \(^P < 0.001\) compared with controls carrying ‘a’ allele and b/b genotype. \(^P = 0.027\) compared with controls carrying ‘a’ allele. \(^P = 0.049\) compared with b/b genotype controls.

\(^c\)Values were mean ± s.d. Statistical significance was determined by ANOVA and a post hoc Tukey test (\(\chi^2\) test).

\(^d\)For comparison log transformed values were used.
hypopituitary adults (31–33). In our study, older age, higher BMI, and presence of hypopituitarism significantly predicted higher IMT measurements in a logistic regression model. The eNOS4a/b polymorphism did not enter this model.

Endothelial dysfunction evaluated by FMD was also a feature of adult GHD (7, 8, 10, 11). Although FMD of brachial artery was lower in our patients, the difference did not reach statistical significance. No significant association was observed between FMD and study parameters. Relatively small sample size may be responsible for the lack of significant difference between FMD measurements. Methodological differences and skill of the operator have effects on FMD; however, in our study, all of the measurements were performed by one experienced investigator (H O). Brachial artery FMD is a functional parameter of endothelium, therefore, is subject to dynamic changes (12). Carotid artery IMT is an anatomical parameter of subclinical atherosclerosis (25). Previous studies also indicated discordant FMD and IMT results in hypopituitary GH-deficient adults; a more atherogenic IMT while FMD was similar to controls or vice versa (34, 35). Recently, it is suggested that although IMT and FMD are used as surrogate markers of preclinical asymptomatic atherosclerosis, they reflect different and independent stages of early atherosclerotic process and are not predicted by each other (36, 37). Measurement of FMD has been reported to have less significance in comparison with IMT in individual cardiovascular risk prediction (38).

Polymorphism of eNOS gene may influence the functional activity of the enzyme and has modulating effects on atherogenesis. Previous studies indicated functional/phenotypic significance of eNOS4a/b polymorphism in various conditions and indirect effects on atherogenesis (39–42). In addition, Wang et al. (43) reported that this polymorphism could influence transcriptional activity of eNOS gene. Tsukada et al. (15) showed that plasma NO level of healthy subjects with the ‘a’ allele was significantly lower than in those without ‘a’ allele (P <0.05).

Fatini et al. (44) showed that the 4a allele and the combined genotypes of ‘a’ allele with other eNOS gene polymorphisms were significantly associated with carotid atherosclerosis. Interestingly, in this study, a relatively high incidence of the 4a allele and combined genotype of ‘a’ allele with another polymorphism (T-786C) was observed in a subset of patients with no traditional risk factors for atherogenesis (n = 30). This observation has been confirmed by Ischihara et al. (18). In Ischihara’s study, eNOS4a allele was an independent risk factor for myocardial infarction, particularly in patients lacking other conventional risk factors (n = 104). Therefore, deleterious effects of eNOS4a allele on atherosclerosis may occur independently from conventional risk factors.

Asakimori et al. (45) indicated that odds ratio for carotid plaque positivity was increased by a factor of 3.72 in the presence of ‘a’ allele of intron 4 polymorphism in non-diabetic hemodialysis patients. However, Lembo et al. (46) could not find any significant difference between the frequency of intron 4a/b polymorphism in hypertensive patients with and without carotid plaques. A different phenotype – established atherosclerosis – was investigated in last two cited studies. We aimed to investigate early vascular manifestation of atherosclerosis in our study.

In our study, although eNOS4a/b polymorphism did not enter in logistic regression model to predict IMT and no significant difference was observed between FMD measurements in patients and controls carrying ‘a’ allele and b/b genotype, ‘a’ allele of eNOS in patients might offer more detrimental effects on early atherosclerotic changes compared with controls. Patients with ‘a’ allele had significantly higher IMT compared with controls carrying ‘a’ allele and b/b genotype. However, no significant difference with respect to IMT was observed between hypopituitary patients carrying ‘a’ allele and bb genotype. The small sample size of patients carrying ‘a’ allele (n: 11) limited us to comment further.

A larger number of patients would be needed to clearly identify the deleterious/protective effects and pathogenetic mechanisms of 4a/b polymorphism on atherogenesis in GH-deficient situations.

The limitations of our study are relatively small sample size and its cross-sectional design. Although conventional replacement therapy was given to the hypopituitary group, we cannot entirely rule out the effects of under or over treatment of cortisol deficiency and hypogonadism on study parameters. In addition, NO plasma concentrations were not directly measured. Markers of oxidative stress and inflammation were not included in study parameters. Other eNOS gene polymorphisms associated with atherosclerotic vascular phenotypes in studies with a wide range of subject numbers (n: 36–879), such as −786T→C and
Glu298→Asp could not be investigated in our study (20, 44–47).

As a conclusion, this study confirms that GH-deficient hypopituitary patients on conventional replacement therapy other than GH have early atherosclerotic changes and a more atherogenic milieu. Although eNOS4a allele in patients seems to have a more detrimental effect compared with ‘a’ allele carrying controls on early atherosclerotic changes, our data are not sufficient to suggest that NOS4a/b polymorphism modifies the atherosclerotic process in GH-deficient situations.

Prospective studies with larger groups of patients and different polymorphisms of eNOS gene are needed to determine the pathogenetic link between eNOS gene polymorphism and atherogenesis in this special group of patients. Future studies are also needed to determine whether improving effects of GH replacement therapy on atherogenesis could be influenced by eNOS4a/b genotypes.

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References


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