Desmopressin increases IGF-binding protein-1 in humans

S B Catrina 1,2, R Rotarus 2, I R Botusan 1,2, M Coculescu 2 and K Brismar 1

1Department of Molecular Medicine and Surgery, Karolinska Institute, L1:O1, Stockholm S-17176, Sweden and 2Department of Endocrinology, University of Medicine and Pharmacy 'Carol Davila', Bucharest 011863, Romania

(Correspondence should be addressed to S-B Catrina; Email: sergiu-bogdan.catrina@ki.se)

Abstract

Context: IGF binding protein-1 (IGFBP-1) is essential for IGF-I bioavailability. High levels of IGFBP-1 are encountered in critically ill patients and are a good predictor marker in acute myocardial infarction. The mechanisms responsible for the elevated IGFBP-1 levels in these conditions are still unclear. Interestingly, high levels of vasopressin have been reported in the above-mentioned conditions.

Objective: To study the effect of vasopressin on IGFBP-1 in humans.

Design: Placebo-controlled cross-over study in patients with central diabetes insipidus (CDI) in whom potential interference from endogenous vasopressin secretion is minimized. After a 3-day desmopressin washout period, each patient received i.v. saline on day 1 and desmopressin (3 μg) on day 2. Blood samples were taken after administration, every 2 h during the whole night, starting at 2000 h.

Patients and setting: Fourteen inpatients with CDI in an endocrinology department of a university hospital.

Results: Serum IGFBP-1 increased within 4 h after 1-desamino-8-D-arginine vasopressin (DDA VP) by 375 ± 73%, compared with a spontaneous fasting increase by 252 ± 46% following placebo administration (P < 0.05). No changes were registered in the levels of either classically regulators of IGFBP-1 (insulin, glucagon, and cortisol) or of IGF-I and glucose. The decrease in plasma osmolarity induced by DDAVP did not precede the increase in IGFBP-1.

Conclusions: DDAVP increases serum levels of IGFBP-1. Further investigation is essential to unravel the clinical potential of this interaction in conditions associated with high IGFBP-1 levels.

Introduction

Insulin-like growth factor-binding protein 1 (IGFBP-1) is one of six members of the IGFBP family, which specifically binds and regulates the bioavailability of IGF-I and IGF-II (1). In humans, IGFBP-1 is mainly produced in the liver and contributes in short-term metabolic regulation (1). Previous investigations suggested that insulin is the main determinant of IGFBP-1 expression through downregulation of the IGFBP-1 hepatic production (2). Several other hormones influence IGFBP-1 production either by stimulating it (e.g., glucagon (3), catecholamines (4), and cortisol (5)) or by suppressing it (e.g., growth hormone (6)). However, these hormones are able to modulate IGFBP-1 serum levels only if the insulin levels are low or absent. Nevertheless, several cytokines (interleukin (IL)-1β, IL-6, and tumour necrosis factor-α, TNF-α) are able to stimulate the IGFBP-1 production and overcome the effect of insulin (recently reviewed in (7)).

The ratio of IGFBP-1 to insulin is increased in highly stressful conditions (critically ill patients) and represents a good predictor marker for mortality as valuable as APACHE II score (8). The loss of the negative regulatory function of insulin at the hepatic level might explain the increased IGFBP-1 in serum. Hypoxia and cytokines (8) were proposed to induce IGFBP-1 in this clinical condition but the exact players are still unknown. Bearing in mind that vasopressin is a stress hormone and inappropriate high serum values are encountered in critically ill patients (9) we tested a direct interaction between vasopressin and IGFBP-1. Two recent reports have raised even more interest for the evaluation of a direct relation between AVP and IGFBP-1 as far as each of them was found to be predictor markers in acute myocardial infarction (AMI) (10, 11).

For this end, we conducted a placebo-controlled cross-over study with i.v. administration of desmopressin (1-desamino-8-D-arginine vasopressin, DDVAP) in 14 patients with central diabetes insipidus (CDI), who have inappropriate low endogenous secretion of vasopressin (AVP).

We suggest a direct interaction between IGFBP-1 and vasopressin which warrants further investigation in larger clinical material and in other clinical scenarios where a combination of high vasopressin levels with high IGFBP-1 levels is encountered, for example, diabetes.

Materials and methods

Patients

Fourteen inpatients with CDI were included in the study after giving informed consent. The major outcome of the
study has been already published (12). Briefly, nine patients had complete CDI, while five had CDI partially. CDI was diagnosed by water restriction test followed by i.v. injection of 1 μg desmopressin (Ferring,almö, Sweden) after 3 h of stable urine osmolality, despite a rising plasma osmolality following fluid restriction (12).

**Study design**

The study protocol was approved by the Ethical Committee of the University of Medicine ‘Carol Davila’ Bucharest. The patients were instructed to discontinue their treatment with desmopressin 3 days before admission to the clinic. At 1900 h, after a standard meal, an indwelling catheter was inserted and at 2000 h each patient received i.v. 1 ml saline in the first evening (day 1) and desmopressin (3 μg; Ferring) in the second evening (day 2). Blood samples were taken at 2-h intervals during the whole night.

**Assays**

The IGFBP-1 levels were measured in the serum using a polyclonal antibody that detects both the phosphorylated and non-phosphorylated forms. The detection limit was 3 ng/ml with inter- and intra-assay coefficients of variation (CVs) of 3 and 10% respectively (2). DDAVP at a concentration up to 4 mg/ml did not cross-react in the IGFBP-1 assay.

The IGF-I serum levels were measured by an RIA after acid–alcohol extraction and cryoprecipitation as described (2). The insulin levels were measured by an RIA commercial kit (Pharmacia & Upjohn Diagnostics AB) with a detection limit of 2 mU/ml and intra- and inter-CV of 5.8 and 6.4% respectively.

Glucagon concentrations were measured by an RIA commercial kit (Linco Research, St Charles, MI, USA) with a detection limit of 20 pg/ml and intra- and inter-CV of 4.6 and 13.4% respectively. Serum cortisol levels were determined by fluorescence immunoassay (AutoDELFIA, Wallac Oy, Turku, Finland). Intra- and inter-assay coefficients of variation were 4.9 and 6.4% respectively.

Blood glucose was determined by glucose oxidase method using YSI 2300 Stat Plus Analyzer.

**Statistical analysis**

The difference between the levels of the hormones measured after treatments was analyzed by repeated measurement of variance using Statistica 7.1 (StatStoft Inc., Tulsa, OK, USA). Differences of P<0.05 were considered significant. The data are represented as mean±S.E.M.

**Results**

**IGFBP-1**

Mean IGFBP-1 levels (2000 h) were 6.4±1.3 and 8.8±1.9 μg/l before injection of saline and DDAVP respectively. Intravenous injection of 3 μg DDAVP induced a significant increase in IGFBP-1 during the first 4 h, which reached a level of 27.1±8.3 μg/l as compared with the spontaneous night increase after saline 14.3±3.2 μg/l (P<0.05; Fig. 1). The IGFBP-1 levels remain higher after DDAVP treatment (43.3±10.1 μg/l) compared with saline (30.5±6.2 μg/l) until the morning (0600 h), however, not statistically significant (P=0.076).

**IGF-I and blood glucose and osmolarity**

Mean IGF-I levels were 160.6±40.2 and 177.6±52.9 μg/l before administration of saline and DDAVP respectively (P=NS). The serum levels did not change over time 177.6±52.9 and 187.4±57.8 μg/l (2200 h), 202.5±55.5 and 209.3±83.3 μg/l (2400 h) after treatment with saline and DDAVP respectively (Fig. 2A).

No significant changes were found in blood glucose levels during the study: 3.7±0.42 vs 4.6±0.7 mmol/l (2000 h), 3.6±0.4 vs 4.1±0.5 mmol/l (2200 h), 4±0.4 vs 4.1±0.3 mmol/l (2400 h) after saline and DDAVP infusion respectively.

We did not observe any difference in basal osmolarity levels between the 2 days of study (297.17±1.92 vs 292.29±1.97 mOsm/kg). On the other hand, DDAVP decreased the serum osmolarity (287.72±1.99 mOsm/kg at 2400 h).

**Glucagon, insulin, and cortisol**

Desmopressin compared with saline had no effect on the serum levels of either cortisol or insulin. A spontaneous
decrease in the mean concentration of both cortisol (Fig. 2B) and insulin (Fig. 2D) occurred during the first hours of follow-up (P < 0.05). The mean concentration of glucagon did not significantly change and there was no difference between groups (Fig. 2C).

**Discussion**

We report here for the first time that DDAVP increases IGFBP-1 blood levels. The study was performed on patients with CDI, in a clinical setting where a potential confounding effect from endogenous secretion of AVP is decreased. The increase in IGFBP-1 after DDAVP is specific and is not due to a change in water balance, which would have had an opposite effect through dilution.

Vasopressin acts via three subtypes of receptors that are classified according to the second messenger system to which they are coupled: V1a and V1b (also known as V3) receptors linked to the phosphoinositol signaling pathway and V2 receptors linked to the adenylate cyclase signaling pathway (13). DDAVP, originally developed as a selective agonist for V2 subtype receptors, has been shown to have a similar affinity for V1b receptors (14).

The main source of circulating IGFBP-1 in males and non-pregnant women is the liver where it is produced by hepatocytes (1) that express V1a and V1b receptors (15) but not V2. Thus, DDAVP might induce IGFBP-1 production through a direct liver effect mediated via a V1b-dependent pathway. In agreement with this, in vitro, DDAVP induced an increased IGFBP-1 secretion by HepG2 cells (human hepatic carcinoma cell line; data not shown). We cannot exclude that DDAVP stimulates IGFBP-1 from other sources as well, such as the kidneys, even though the renal production may not contribute substantially to the circulating IGFBP-1 level (1).

We did not observe a modulation of classical regulators of IGFBP-1 after administration of DDAVP. Nevertheless, due to the sampling schedule, we could not completely reject the possible contribution of some of them. For instance, cortisol has been reported to be stimulated by DDAVP (16). However, the DDAVP effect on cortisol levels lasts less than an hour thus making this explanation less relevant (16). We even observed a
tendency of reduction of the cortisol levels over time following DDAVP injection, even though statistically this was non-significant. This might reflect the dilution effect of the free water retained after DDAVP injection. DDAVP induces also production of catecholamines that, in turn, might result in increased IGFBP-1 serum levels (17). However, it has previously been reported that the effect of catecholamines on IGFBP-1 lasts < 120 min after inoculation (4). Furthermore, DDAVP in doses similar to those used in the present study did not influence epinephrine but only norepinephrine level (17), which has only a marginal effect on IGFBP-1 (4).

We could not detect any difference in the blood levels of glucagon or insulin after injection of DDAVP compared with saline. Even though there is a clear evidence that vasopressin is able to stimulate the secretion of both hormones through V1 receptors (18), it is not surprising that we did not register it, taking into account that the effect is transitional and lasts for less than 60 min.

Injection of DDAVP did not change the concentrations of total IGF-I or glucose. Although IGFBP-1 is an acute regulator of IGF-I bioavailability (1), short-term changes in its blood levels are not always followed by changes in glucose or in total or free IGF-I levels (19).

In conclusion, we demonstrated that DDAVP increases the blood levels of IGFBP-1 suggesting a novel mechanism to explain the high levels of IGFBP-1 observed in critically ill patients or patients with diabetes mellitus. Further investigations are needed to confirm the clinical relevance of our finding.

Acknowledgements

We thank nurses Constanta Navodaru, Eugenia Mirea, and Micsunica Ilie for their care of the patients, Inga-Lena Wivall for exceptional technical assistance and Agneta Hilding for statistical advice. We also thank the Family Stefan Persson Foundation, von Kantzow Foundation, Thuring Foundation, Tore Nilson Foundation, and Elin Danielsson for their financial support.

References


Received 11 January 2008
Accepted 17 January 2008