Cut-off limits of the GH response to GHRH plus arginine test and IGF-I levels for the diagnosis of GH deficiency in late adolescents and young adults


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Abstract

Objective: To define the appropriate diagnostic cut-off limits for the GH response to GHRH + arginine (ARG) test and IGF-I levels, using receiver operating characteristics (ROC) curve analysis, in late adolescents and young adults.

Design and methods: We studied 152 patients with childhood-onset organic hypothalamic–pituitary disease (85 males, age (mean ± S.E.M.): 19.2 ± 0.2 years) and 201 normal adolescents as controls (96 males, age: 20.7 ± 0.2 years). Patients were divided into three subgroups on the basis of the number of the other pituitary hormone deficits, excluding GH deficiency (GHD): subgroup A consisted of 35 panhypopituitary patients (17 males, age: 21.2 ± 0.4 years), subgroup B consisted of 18 patients with only one or with no more than two pituitary hormone deficits (7 males, age: 20.2 ± 0.9 years); and subgroup C consisted of 99 patients without any known hormonal pituitary deficits (60 males, age: 18.2 ± 0.2 years). Both patients and controls were lean (body mass index, BMI < 25 kg/m²).

Patients in subgroup A were assumed to be GHD, whereas in patients belonging to subgroups B and C the presence of GHD had to be verified.

Results: For the GHRH + ARG test, the best pair of highest sensitivity (Se; 100%) and specificity (Sp; 97%) was found choosing a peak GH of 19.0 µg/l. For IGF-I levels, the best pair of highest Se (96.6%) and Sp (74.6%) was found using a cut-off point of 160 µg/l (SDS: −1.3). Assuming 19.0 µg/l to be the cut-off point established for GHRH + ARG test, 72.2% of patients in subgroup B and 39.4% in subgroup C were defined as GHD. In patients belonging to group B and C with a peak GH response < 19 µg/l to the test, IGF-I levels were lower than 160 µg/l (or less than 1.3 SDS) in 68.7 and 41.6% of patients respectively predicting severe GHD in 85.7% of panhypopituitary patients (subgroup A).

Conclusions: In late adolescent and early adulthood patients, a GH cut-off limit using the GHRH + ARG test lower than 19.0 µg/l is able to discriminate patients with a suspicion of GHD and does not vary from infancy to early adulthood.

Introduction

The clinical features of adult growth hormone deficiency (GHD) are recognizable but not distinctive, so clinical suspicion must be confirmed by biochemical tests. Within an appropriate clinical context, GHD in adults has to be shown by a single provocative testing, provided that it is reproducible with clear normative limits. The insulin tolerance test (ITT) has been indicated as the test of choice while the GH-releasing hormone plus arginine (GHRH + ARG) test is considered the best alternative (1–3).

The diagnosis of GHD in late adolescents and young adults, defined as the ‘transition period’ from 15 to 25 years, represents a major clinical challenge and raises questions about the most appropriate method and cut-off value for diagnosing GHD after completion of growth and puberty (4).

Though the evaluation of spontaneous GH secretion, IGF-I and IGF-binding protein-3 levels could be useful in
diagnosing GHD in childhood (5) this is not the case in adulthood due to remarkable overlap between GHD and normal subjects (1–3). Thus, the most appropriate reevaluation of somatotroph function in adolescence or young adulthood is likely to be represented by measurement of a serum IGF-I concentration and/or a GH stimulation test (4). In fact, low IGF-I levels can be definitive evidence of persistent severe GHD in subjects with genetic GHD or panhypopituitarism (4, 6, 7), although normal IGF-I levels never rule out severe GHD (1, 7–10).

The Consensus Statement about the management of the GH-treated adolescent in the transition to adult care stated that 3 μg/l as the cut-off limit in response to ITT used to define severe GHD in adulthood, is too conservative in the transition period (4). Thus, a cut-off limit of the GH response < 5 μg/l to whatsoever provocative test, which is in between the adult and childhood cut-off limits (< 3 and < 7 μg/l respectively), was proposed as a criterion for severe GHD in the transition period (4). However, this assumption was not clearly evidence based. In fact, in pediatric age, the most pronounced GH response to a stimulation test occurs in late puberty with GH levels inevitably exceeding 5 μg/l (11). Recently, we defined a peak GH response < 6.1 μg/l as the cut-off limit for ITT in late adolescents and young adults using the receiver operating characteristics (ROC) curve analysis with a 96% sensitivity (Se) and a 100% specificity (Sp) (11). This cut-off limit for ITT is higher than that recommended by the Consensus Statement and would therefore avoid false-negative diagnostic responses (11).

GHRH + ARG test demonstrated excellent Se and Sp both in childhood and in adulthood assuming appropriate cut-off limits (12–15). However, for the adolescence and young adults phase (defined as the age period from 15 to 25 years), the reliability of the GHRH + ARG test in confirming childhood onset GHD has not been assessed yet (16–18).

Based on these foregoing matters, the aim of this study was to define the appropriate diagnostic cut-off limits of GH response to the GHRH + ARG test as well as of IGF-I levels and their Se and Sp for the diagnostic accuracy of GHD in late adolescents and young adults, using an appropriate methodology for the ROC curve analysis.

### Subjects

We evaluated the total IGF-I levels and GH response to the GHRH-ARG test in 152 lean patients (85 males and 67 females, age (mean ± s.e.m.): 19.2 ± 0.2 years (range: 15–25 years), BMI (mean ± s.e.m.): 20.9 ± 0.2 kg/m²), all with structural hypothalamic–pituitary abnormalities and childhood-onset organic hypothalamic–pituitary disease (Table 1). All the patients considered in this study were diagnosed as GH deficient during childhood. The diagnosis of GHD in childhood had been based on clinical and auxological criteria and evidence of a GH response < 10 μg/l to at least two 'classical' provocative tests (19).

The most common disorders were congenital pituitary diseases in 126 patients (82.8%; i.e., pituitary hypoplasia, septo-optic dysplasia, ecticopic posterior pituitary) and acquired pituitary disease in 26 (17.2%) (i.e., hypothalamic–pituitary adenomas, craniopharyngiomas, and other peripituitary tumors requiring neurosurgery and/or radiotherapy and post-traumatic hypopituitarism; Table 2). The patients were subdivided into three subgroups on the basis of the number of the other pituitary hormone deficits, excluding GHD. The first subgroup consisted of 35 panhypopituitary patients (subgroup A: 17 males and 18 females, age (mean ± s.e.m.): 21.2 ± 0.4 years). They all had secondary hypothyroidism, hypoadrenalism, and hypogonadism; 12 out of 35 patients had also diabetes insipidus. The second subgroup consisted of 18 patients with only one or with no more than two pituitary hormone deficits (subgroup B: 7 males and 11 females, age (mean ± s.e.m.): 20.2 ± 0.9 years). Three patients (2.5%) had secondary hypothyroidism, two (1.7%) secondary hypoadrenalism, six (5.1%) secondary hypogonadism, two (3.5%) had both secondary hypothyroidism and hypoadrenalism, and two (1.7%) had both secondary hypothyroidism and hypogonadism. The third subgroup consisted of 99 patients without any current known hormonal pituitary deficits (subgroup C: 60 males and 39 females, age (mean ± s.e.m.): 18.2 ± 0.2 years). All patients were studied while on optimized replacement therapy for other pituitary deficits including cortisone acetate, thyroid hormone, transdermal.

### Table 1 Characteristics of the population studied. Results are expressed as mean ± s.e.m.

<table>
<thead>
<tr>
<th>Patients</th>
<th>Control subjects</th>
<th>Subgroup A (panhypopituitary patients)</th>
<th>Subgroup B (1 or 2 pituitary hormone deficits)</th>
<th>Subgroup C (no pituitary hormone deficits known)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>201</td>
<td>35</td>
<td>18</td>
<td>99</td>
</tr>
<tr>
<td>Age (years)</td>
<td>20.7 ± 0.2</td>
<td>21.2 ± 0.4</td>
<td>20.2 ± 0.9</td>
<td>18.2 ± 0.2</td>
</tr>
<tr>
<td>Sex (male/female)</td>
<td>98/105</td>
<td>17/18</td>
<td>7/11</td>
<td>60/39</td>
</tr>
</tbody>
</table>

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Table 2  Etiology of hypopituitarism in patients studied.

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>Group A (35 pt.)</th>
<th>Group B (18 pt.)</th>
<th>Group C (99 pt.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acquired diseases</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pituitary adenomas</td>
<td>5</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Craniohypophygiomas</td>
<td>5</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>Traumatic brain injuries (TBI)</td>
<td>2</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>Peripititary lesions</td>
<td>2</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Congenital diseases</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pituitary hypoplasia</td>
<td>4</td>
<td>2</td>
<td>76</td>
</tr>
<tr>
<td>Septo-optic dysplasia (SOD)</td>
<td>3</td>
<td>2</td>
<td>5</td>
</tr>
<tr>
<td>Empty sellae</td>
<td>14</td>
<td>4</td>
<td>16</td>
</tr>
</tbody>
</table>

gonadal steroids, i.m. testosterone, and desmopressin, where appropriate.

In all patients studied recombinant human GH (rhGH) treatment had been discontinued when patients reached their final height, as indicated by growth velocity below 2 cm/year in the last year of treatment. The reevaluation of the somatotroph function was evaluated by GHRH + ARG test and total IGF-I levels after at least 3 months of GH therapy (range: 3–24 months).

As a control group, we studied 201 healthy lean late adolescents and young adults (96 males and 105 females; age (mean ± S.E.M.): 20.7 ± 0.2 years (range: 15–25 years), BMI (mean ± S.E.M.): 21.3 ± 0.2 kg/m²).

Patients and controls were enrolled in a multi-center study (six pediatric endocrinology centers and two adult endocrinology centers).

All subjects were measured for height and weight and all of them were lean (BMI < 25 kg/m²).

The study had been approved by the local Ethical Committees and all the patients provided informed consent.

Methods

After an overnight fast, all subjects underwent both total IGF-I measurement and GHRH + ARG test (GHRH1-29; GEREF, Serono, Italy; 1 µg/kg i.v. at 0 min; ARG hydrochloride, 0.5 g/kg i.v. over 30 min from 0 to +30 min, up to a maximum of 30 g). Blood samples for GH evaluation were taken every 15 min from 0 to +90 min.

The measurement of both serum GH and total IGF-I concentrations was centralized and performed using the laboratory of the Division of Endocrinology and Metabolism, University of Turin.

Serum GH was assayed by immunoradiometric method assay (IRMA) method (HGH-CTK IRMA; Diasorin, Saluggia, Italy). All samples from an individual subject were analyzed together. The Se of the method was 0.15 µg/l. The inter- and intra-assay coefficients of variation were 3.5–4.4 and 5.1–7.5% respectively at GH levels of 1.98–41.92 and 2.99–42.45 µg/l respectively.

Serum total IGF-I was assayed by RIA method (SM-C-RIA-CT; Pantec, Turin, Italy) after acid–ethanol extraction to avoid interference by binding proteins. The Se of the method was 0.1 µg/l. The inter- and intra-assay coefficients of variation were 5.0–9.5 and 8.8–10.8% respectively at IGF-I levels of 79.41–712.3 and 79.6–766.4 µg/l respectively.

Statistical analysis

Results are expressed as mean ± S.E.M. of absolute values (µg/l). For IGF-I levels, age-adjusted SDS were also calculated. The normality of data distribution was confirmed by the Shapiro–Wilk test. Comparison of mean values between two different categories of the considered variables was performed using Student’s t-test or by one-way ANOVA among groups. Age differences among groups and age differences in peak GH to the GHRH + ARG test and in individual total IGF-I levels both in controls and in the subgroups of patients were evaluated by ANOVA. Correlations between IGF-I and peak GH response to the GHRH + ARG test and between age and peak GH response to GHRH + ARG or to IGF-I levels were analyzed with Pearson’s coefficient. A P value of < 0.05 was considered statistically significant. Receiver operating curve (ROC) analysis was used for the evaluation of the cut-off points (Medcalc 7.2). The lowest limit of normality for peak GH responses to the GHRH + ARG test for the diagnosis of probable permanent GHD during adolescence was defined as the value that provided the best pair of Se/Sp values (19). Se was calculated as the number of true-positive (TP) cases out of the sum of TP plus false-negative cases (TP/(TP + FN)) and Sp was calculated as the number of true-negative cases out of the sum of true-negative plus false-positive cases (TN/(TN + FP)). For the purpose of ROC curve analysis we assumed that the panhypopituitary patients (subgroup A) had GHD, according to other studies and the statement derived from either the Growth Hormone Research Society consensus or the recent Endocrine Society Clinical Practice Guidelines for adult GHD, suggesting increased likelihood of GHD and increased number of other pituitary hormone deficits (1, 20). The other two subgroups of patients (B and C: patients without or with no more than two pituitary hormonal deficits) included subjects with probably either normal or impaired GH secretion, in which the presence of GHD had to be verified.

ROC curves are constructed by plotting the Se on the ordinate as a function of the complement of the Sp for all the possible cut-off values of the diagnostic test. Each point of the ROC curves represents a Se/Sp pair corresponding to a particular decision threshold. The area under the ROC curve (ROC AUC) represents the probability of correctly distinguishing between affected and non-affected individuals. A perfect diagnostic test has a ROC curve that passes through the upper left-hand corner (AUC = 1), where the TP fraction is 1.0 or 100% (perfect Se) and the FP fraction is 0 (perfect Sp).
Tests with an AUC of <0.5 would not discriminate between affected and non-affected subjects (21).

The diagnostic cut-off points were calculated for both peak GH response to the GHRH+ARG test and individual IGF-I levels. To provide optimal separation of GHD and normal subjects, we applied two criteria: 1) to minimize misclassification of control subjects and GHD, balancing between high Se and high Sp and 2) to provide the highest pair of Se and Sp values for GHD.

For each of the cut-off points calculated by the ROC curve analysis, the accuracy value, defined as the number of true cases out of the total number of subjects studied, was also provided.

**Results**

**Peak GH response to the GHRH + ARG test**

No statistically significant differences were present in adult height, weight, or BMI among subgroups of patients or between those with congenital or acquired GHD.

No age or gender differences were found between controls and patients or among subgroups of patients.

Neither age difference nor correlation was found in mean peak GH response to the GHRH+ARG test and age among groups, both in controls and in subgroups of patients.

The mean peak GH in control subjects (69.3 ± 2.6 µg/l) was significantly higher than that in patients (P < 0.0001). Among the subgroups of patients, the mean peak GH response to the GHRH+ARG test was significantly different (P < 0.0001). In particular, in subgroup A peak GH (3.4 ± 0.8 µg/l) was lower than that found in subgroup B (18.1 ± 8.2 µg/l; P < 0.05) and both were significantly lower than that found in subgroup C (42.4 ± 3.6 µg/l; P < 0.001; Fig. 1, Table 3). Peak GH response to GHRH+ARG ranged between 0.1 and 19.0 µg/l in panhypopituitary patients (subgroup A) and between 0.1 and 150.8 µg/l in the other subgroups of patients (subgroups B and C), potentially affected by GHD, and between 9.7 and 235.0 µg/l in the controls (Fig. 1).

The GH peaks always occurred between 30 and 60 min.

**ROC curve analysis**

Regarding the ROC curve analysis for the GHRH+ARG test, the best pair of values with the highest Se (100%) and the highest Sp (97%) was found using a peak GH cut-off point of 19.0 µg/l, with an accuracy of 81.5% (Fig. 2: Table 4).

Applying the cut-off points to the GHRH+ARG test of 19.0 µg/l, we found that among the subgroup B 13 out of 18 patients (72.2%) resulted GHD, whereas in the subgroup C 39 out of 99 (39.4%) resulted as affected by a persistent form of GHD.

By adopting the current published GH cut-off points for the GHRH+ARG test (20 µg/l in pediatric population (20) and 9.0 µg/l in adult population (1–3)) none and 2 out of 35 (5.7%) of the panhypopituitary adolescent patients would be excluded from a correct diagnosis of GHD respectively.

**IGF-I levels**

Individual total IGF-I levels (mean ± S.E.M.) and IGF-I SDS (mean ± S.E.M.) were significantly higher in control subjects (310.1 ± 6.1 µg/l and -0.4 ± 0.1) than in patients (P < 0.0001). Among the subgroups of patients, IGF-I levels were significantly different (P < 0.0001). In particular, in subgroup A, IGF-I levels (86.7 ± 12.9 µg/l and -1.97 ± 0.1) were lower than those found in subgroup B (157.1 ± 34.8 µg/l and -1.4 ± 0.2; P < 0.05) and in subgroup C (225.5 ± 18.1 µg/l and -1.07 ± 0.1; P < 0.0001), whereas in subgroups B and C IGF-I levels were similar (Fig. 3, Table 3).

There were no age or gender differences in mean IGF-I levels in patients or control groups. No age or gender differences were found between control and patient groups or among subgroups of patients.

Neither age difference nor correlation were found in mean total individual IGF-I levels among control and patient subgroups.

**ROC curve analysis**

Regarding the ROC curve analysis for IGF-I levels, the best pair of values for highest Se (96.6%) and highest Sp (74.6%) was found using a peak GH cut-off point of 160 µg/l (IGF-I SDS: -1.3), with an accuracy of 70.8% (Fig. 4: Table 4).

Considering the whole population studied, individual total IGF-I levels were significantly and positively correlated (r = 0.4; P < 0.001) to individual peak GH responses to GHRH+ARG test.

In patients of subgroups B and C who were found to be GHD by adopting the cut-off response to the GHRH+ARG test of 19.0 µg/l, IGF-I levels were significantly lower than in those without GHD (182.3 ± 21.0 µg/l versus 374.5 ± 30.2 µg/l in subgroup B, P < 0.05 and 182.3 ± 21.0 µg/l versus 272.3 ± 33.4 µg/l in subgroup C, P < 0.05).
Moreover, IGF-I levels were <160 μg/l (or less than −1.3 SDS) in 68.7% and in 41.6% of patients who resulted as GHD (peak GH below 19.0 μg/l) belonging to subgroups B and C respectively.

On the other hand, IGF-I levels were below 160 μg/l in 30 out of 35 panhypopituitary patients (subgroup A), who had GHD demonstrated by GH peak response to GHRH+ARG test below 19.0 μg/l. Thus low IGF-I levels could predict severe GHD in 85.7% of this population.

Finally, in subgroups of patients in whom GHD was demonstrated, this pituitary deficit was isolated in 39 out of 87 (44.9%) cases and associated with other pituitary deficits in 48 out of 87 (55.1%) cases.

The cut-off points with the best pair of values for Se/Sp for the GHRH+ARG test and IGF-I levels, with confidence intervals at 95%, are shown in Table 4.

**Discussion**

The results of this study firstly provide cut-off limits for the GH response to the GHRH+ARG test and IGF-I levels for lean late adolescents and young adults who are suspected for GHD, either isolated or in association with other pituitary deficits.

Data about normative cut-off levels in late adolescence and young adulthood are scarce. This applies to GH/IGF-I axis whose function and response to stimulation during this period of life has never been systematically defined. Consequently, the diagnosis of GHD in late adolescence and young adulthood cannot rely on definite normative data. This is clearly indicated by the Consensus Statement on the management of the GH-treated adolescent in the transition to adult care that indicated the GH cut-off point for the diagnosis of GHD in a peak lower than 5 μg/l (4). Indeed, this cut-off point was not evidence based and simply indicated a value in between the cut-off points of 7 and 3 μg/l; these cut-offs are generally adopted for the diagnosis of GHD in childhood and adulthood respectively (19, 22, 23) and reflect the assumption that GH secretion declines through aging (23–26). Actually, although ITT was assumed by this Consensus as the test of choice, the recommendation generally referred to any provocative test (4).

It is widely accepted that the diagnostic cut-off levels have to be defined in an appropriate population for each provocative test (1, 19). In a previous study, we were first to define the appropriate diagnostic cut-off limits of GH response to the ITT test in late adolescents and young adults.

### Table 3

<table>
<thead>
<tr>
<th>Patients</th>
<th>Subgroup A (panhypopituitary patients)</th>
<th>Subgroup B (1 or 2 pituitary hormone deficits)</th>
<th>Subgroup C (no pituitary hormone deficits known)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GH (μg/l)</td>
<td>69.3±2.6*</td>
<td>18.1±8.2+†</td>
<td>42.4±3.6*</td>
</tr>
<tr>
<td>IGF-I (μg/l)</td>
<td>310.1±6.1*</td>
<td>86.7±12.9+‡</td>
<td>157.1±34.8*</td>
</tr>
<tr>
<td>IGF-I SDS</td>
<td>−0.4±0.1*</td>
<td>−1.97±0.1+‡</td>
<td>−1.4±0.2*</td>
</tr>
</tbody>
</table>

*P<0.0001 among groups and between control subjects and patients; †P<0.001 subgroups A and B versus subgroup C; ‡P<0.05 versus subgroup B; §P<0.0001 versus subgroup C.

**Figure 2** ROC curves for peak serum GH responses to the GHRH+ARG test in the lean adolescent and young adult population. The arrow indicates the location on the ROC curves of the diagnostic cut-off point that minimizes misclassification of panhypopituitary patients and control subjects. The cut-off point, sensitivity and specificity, with their confidence intervals (c.i.) at 95%, ROC AUC and accuracy are shown in the box.

### Table 4

<table>
<thead>
<tr>
<th>GHRH+ARG test</th>
<th>IGF-I levels</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cut-off point (μg/l)</td>
<td>19.0</td>
</tr>
<tr>
<td>Sensitivity (%; 95% c.i.)</td>
<td>100 (89.9; 100)</td>
</tr>
<tr>
<td>Specificity (%; 95% c.i.)</td>
<td>97 (93.6; 98.9)</td>
</tr>
<tr>
<td>Accuracy (%)</td>
<td>81.5</td>
</tr>
</tbody>
</table>

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young adults as GH peak below 6.1 μg/l with a Se of 96% and a Sp of 100% (11). Thus, whenever the test of choice for the diagnosis of GHD during this period of life is ITT, one should refer to this cut-off level. Some physicians concern remains about the poor reproducibility of ITT, questioning its Sp (27, 28). Moreover, hypoglycemia may be contraindicated in some clinical conditions and it is generally not well tolerated (22, 29), in particular in adolescents and young adults with a clinical history of organic pituitary disease.

In this study, we aimed to appropriately define using the ROC curve analysis the cut-off levels of the normal GH response to GHRH CARG that is considered as reliable as ITT for the diagnosis of adult GHD (1, 12, 22, 30). It is well known that GHRH CARG test demonstrated excellent Se and Sp both in childhood and in adulthood assuming appropriate cut-off limits (12, 14–16) but normative data in late adolescence and young adulthood were lacking. Although the use of the GHRH + ARG test is questioned regarding its ability to diagnose GHD in subjects in whom the defect is represented by hypothalamic impairment, namely GHRH deficiency (e.g., after hypothalamic–pituitary irradiation) (31), it is actually increasingly used and the results obtained with this test strongly correlate with those obtained by testing with ITT (14, 31).

Our present data show that a cut-off limit of 19.0 μg/l after the GHRH + ARG test in adolescence is reliable both as a screening and a diagnostic test in a population suspected for GHD in adolescents and young adults, because of its high Se (100%) and high Sp (97%). This is highly essential as the GHRH + ARG test should be used as a first-step diagnostic test rather than a screening test. Moreover, this cut-off overlaps with that considered for diagnosis of GHD in childhood (i.e., 20.0 μg/l) (22) and is about twofold higher than that recommended for the diagnosis of GHD in adults (i.e., 9.0 μg/l) (1, 12, 14, 15). Thus, the first consideration is that the evaluation of the maximal secretory capacity of somatotroph cells by GHRH CARG in late adolescents and young adults allows showing normative levels that are similar to those in children and still higher than in adults. Indeed, this evidence emphasizes the importance of considering cut-off levels as a function of the phase of life.

The definition of cut-off points by ROC curve analysis is the most appropriate in order to provide the best pair of values for Se/Sp (21). In fact, a high Se/Sp cut-off point (i.e., 19.0 μg/l) maximizes detection of adult GHD and minimizes misclassification of normal subjects as GH deficient. In our study, these cut-off limits were found to have a pair of Se/Sp values similar to the ideal of 95% that might be considered the most appropriate either to clearly reconfirm the diagnosis of pediatric GHD or to exclude GHD if the clinical suspicion is low. It remains that these cut-off points referred to the GH assay used in this study and the remarkable variability among different assays is well known (32). Again, it has to be noticed that these cut-off limits were established in lean subjects only; considering that GH secretion is a function of weight and adiposity (15, 33–35), cut-off limits appropriate for overweight and obese subjects remain to be defined.

Based on the definition of these cut-off values appropriate for late adolescents and young adults, our findings allow us to confirm that childhood-onset panhypopituitarism is always associated with severe GHD (16, 36, 37). High risk for severe GHD (72%) is also displayed by patients who had been treated with rhGH in childhood with only one or with no more than two other pituitary hormone deficits. However, also in the group of patients with suspected GHD, without any other current pituitary deficits, as high as 40% of the patients showed a persistent GHD needing rhGH substitutive therapy in adult age.
Moreover, we evaluated by ROC curve analysis the cut-off limit for total IGF-I levels for adolescent and young adults. The cut-off limit of 160 μg/l (or −1.3 SDS) would have identified more than 87% of panhypopituitary patients with severe GHD and almost 50% of the patients affected by one or two pituitary hormone defects, other than suspected GHD, and in the group of patients who were not known to be affected by pituitary hormone deficiencies other than GH during childhood, suggesting that low IGF-I levels are able to strongly predict severe GHD also in late adolescents and young adults. Furthermore, despite the lower diagnostic pair of Se/Sp of this parameter very low levels of total IGF-I can be considered definitive evidence of severe GHD in a remarkable percentage of patients (especially in panhypopituitary patients) who could therefore skip provocative tests for GH secretion (7). However, although even in patients with total anterior hypopituitarism total IGF-I levels are often normal, this finding cannot rule out severe GHD that therefore ought to be verified by provocative testing of GH secretion and it is always mandatory to verify the suspicion of isolated GHD. These findings agree with those recorded in a previous study performed in another cohort of patients (11).

In conclusion, even considering the need to refer to age-related hormonal cut-off limits specific for each provocative test, our findings further demonstrate and strengthen the reliability of GHRH+ARG as a provocative test in adolescent and young adult patients in order to clearly reevaluate the GH secretion in the transition period. Although the cut-off values in infancy and in adolescents and young adults are similar, the GHRH+ARG test has a great accuracy and diagnostic value.

Acknowledgements

This study was partially supported by FS MEM and University of Turin, Italy.

References

G Corneli and others


