CLINICAL STUDY

High phenotypic intrafamilial variability in patients with Pendred syndrome and a novel duplication in the *SLC26A4* gene: clinical characterization and functional studies of the mutated *SLC26A4* protein

Laura Fugazzola^{1,3}, Valentina Cirello¹, Silvia Dossena⁶, Simona Rodighiero⁶, Marina Muzza¹, Pierangela Castorina⁴, Faustina Lalatta⁴, Umberto Ambrosetti^{2,5}, Paolo Beck-Peccoz^{1,3}, Guido Bottà⁶ and Markus Paulmichl⁶

Departments of ¹Medical Sciences and ²Otorhinolaryngology and Ophthalmology, University of Milan, 20122 Milan, Italy, ³Endocrine and Diabetes Unit, ⁴Medical Genetic Unit and ⁵Audiology Unit, Fondazione Policlinico IRCCS, 20122 Milan, Italy, ⁶Department of Biomolecular Sciences and Biotechnology and CIMAINA, University of Milan, 20133 Milan, Italy

(Correspondence should be addressed to L Fugazzola; Email: l.fugazzola@policlinico.mi.it)

Abstract

Objective: Pendred syndrome (PS) is characterized by the association of sensorineural hearing loss (SNHL) and a partial iodide organification defect at the thyroid level. It is caused by mutations in the *SLC26A4* gene. The encoded transmembrane protein, called pendrin, has been found to be able to transport chloride and other anions.

Design: The aim of the present study was to characterize a family with PS, which shows a strong intrafamilial phenotypic variability, including kidney atrophy in one member. The age of disease-onset was significantly different in all three affected siblings, ranging from 2 to 21 years for thyroid alterations and from 1.5 to 11 years for SNHL.

Methods: Clinical and genetic studies were carried out in affected siblings. The functional activity of the novel duplication found was studied by a fluorimetric method in a human renal cell line (HEK293 Phoenix) in which the protein was overexpressed.

Results: All three siblings were found to be compound heterozygotes for the missense mutation (1226G>A, R409H) and for a novel 11 bp duplication (1561_1571CTTGGAATGGC, S523fsX548). The latter mutation creates a frameshift leading to the loss of the entire carboxy-terminus domain. Functional studies of this mutant demonstrated impaired transport of chloride and iodide when expressed in HEK 293 Phoenix cells, when compared with wild type pendrin.

Conclusions: A novel 11 bp duplication was found in a family with Pendred syndrome, showing a high intrafamilial phenotypic variability. An impaired transmembrane anionic transport of the mutated *SLC26A4* protein was demonstrated in functional studies using a heterologous cell system.

European Journal of Endocrinology 157 331–338

Introduction

In 1997, the *SLC26A4* (or PDS, OMIM 605646) gene was cloned and mapped on chromosome 7q22-31.1 (1). It codes for a protein called pendrin, which is mainly expressed not only in the thyroid, inner ear, and kidney, but also in the breast, endometrium, placenta, and testis (2–7). Pendrin is thought to be composed of 12 transmembrane domains with an intracellular N-terminus and C-terminus (8, 9). Mutations of this gene have been associated, in compound heterozygosity or in homozygosity, with Pendred syndrome (PS), an autosomal recessive disorder first described in 1896 and characterized by the association of sensorineural hearing loss (SNHL) and goiter (10). The phenotype of PS is highly variable with regard to deafness, goiter, and

thyroid function (12–15). Sensorineural deafness is congenital, and in most cases has a progressive course. However, in some patients it is already severe at birth. Deafness is associated with radiologically detectable structural malformations of the inner ear. In particular, typical malformations are both an enlarged vestibular aqueduct (EVA) and an enlarged endolymphatic duct and sac (EED and EES) (16, 17). SNHL is a constant feature of PS, whereas goiter is observed in about 50% of the affected individuals and can vary from a slight enlargement to a large multinodular goiter, with the time of onset ranging from the first months of life until puberty. The goiter is the consequence of a partial iodide organification defect, as demonstrated by the positive perchlorate discharge test. However, the defect is only partial and most patients are euthyroid or subclinically

hypothyroid (18). The clinical features of PS are the consequence of impairment in the function of pendrin. Indeed, in the inner ear pendrin is involved in the endolymph pH control and its dysfunction, owing to an altered endolymph composition, which results in an enlargement of the membranous labyrinth structures and the damage of the neuroepithelium secondary to osmotic and toxic mechanisms (3).

At the thyroid level, the role of pendrin is not clear. Some groups, including ours, gave evidence that pendrin could function as a transporter of iodide at the apical pole of the thyroid cell and its impaired function could thus lead to the iodide organification defect observed in these patients (8, 9, 19–21). However, since the thyroid phenotype is often mild or absent, it has been hypothesized that in the absence of pendrin function, an iodide flux into the colloid space may still occur through other transport systems beside SCL26A4 or through the intervention of other genetic and environmental factors (22). Pendrin is also expressed in the kidney, where it plays a crucial role in bicarbonate secretion and in the acid–base balance besides chloride reabsorption (4, 23). It is reasonable to hypothesize that patients affected with PS may have disturbances of renal function, especially in the regulation of electrolytes and acid–base balance. However, no abnormalities of systemic pH or serum chloride levels have been reported in PS patients, possibly due to the redundancy of compensatory mechanisms in the kidney.

After cloning of the *SLC26A4* gene, more than 100 different mutations have been described (<http://www.medicine.uiowa.edu/pendredandbor>) in patients with classical PS or affected with a non-syndromic hearing loss named familial EVA syndrome or DFNB4 and characterized by the typical inner ear malformations without thyroid abnormalities (24). The vast majority of PDS mutations involve a single nucleotide (either missense, non-sense, or splice-site). In the present study, a novel 11 bp duplication of the *SLC26A4* gene is described and functionally characterized. Interestingly, this mutation was found in compound heterozygosity in a family with an extremely variable phenotype.

Materials and methods

Patients

Patients MM, MA, and MC were born to apparently non-related parents who have normal hearing, no signs of hypothyroidism and no goiter. Patient MC came to our attention for pre-conceptual genetic counseling.

Patient MM, female aged 45 years, had a diagnosis of SNHL at the age of 5 years. At that time, she had normal speech. The SNHL showed a progressive worsening in the following years up to a profound hearing deficit (80 dB). At 21 years of age, due to the appearance of fatigue,

thyroid function was explored and a mild hypothyroidism was diagnosed, with normal free triiodothyronine (FT3) levels, slightly decreased free thyroxine (FT4) levels (9.8 pmol/l), and thyrotropin (TSH) levels above the normal range (8.9 mU/l). Thyroid ultrasound performed at the same age showed a small multinodular goiter, and the patient began L-T4 replacement treatment. Moreover, at 30 years of age, a bilateral renal atrophy was documented and the patient was submitted to right kidney transplantation at age 41. She was also affected by secondary hyperparathyroidism and had normal IQ.

Patient MA, male aged 44, had a diagnosis of SNHL at the age of 18 months, in a prelingual period. Again, deafness showed a progressive worsening, up to a profound hearing deficit (80 dB). Due to the supposed diagnosis of PS in the sister, thyroid function was also evaluated in her two brothers. In patient MA, a hypothyroidism with low free thyroid hormones levels and TSH = 15 mU/l was diagnosed at the age of 2 years, and L-T4 replacement treatment was started. He had a low IQ and language disturbances.

Patient MC, male aged 42, experienced the sudden appearance of a profound SNHL (80 dB) at the age of 11 years. At that time, the patient had normal speech. At the age of 5 years and 4 months, the diagnosis of subclinical hypothyroidism with normal free thyroid hormones levels and elevated TSH (8.1 mU/l) was made and L-T4 replacement treatment was immediately started. Thyroid ultrasound showed a slightly enlarged gland. Patient MC had a normal IQ. None of the three patients reported symptoms suggestive of vestibular dysfunction.

Clinical analyses

Serum TSH, free T4, free T3, thyroglobulin, and anti-thyroglobulin/anti-thyroperoxidase antibodies levels were measured by an immunometric assay (AutoDelfia; Wallace, Turku, Finland). Normal value ranges are 0.25–4.2 mU/l for TSH, 9–20 pmol/l for FT4, and 3.8–8.0 pmol/l for FT3. In all patients, ultrasound (US) of the neck was carried out, and the thyroid volume (TV) was calculated by the ellipsoid formula.

A thyroid scintigraphy (TS) with ^{123}I and a perchlorate discharge test were performed. In particular, ^{123}I was administered intravenously and counts were obtained at 1 and 2 h. For the discharge test, 1 g of potassium perchlorate (KClO_4) was administered orally and the uptake was calculated after 1, 2, and 3 h respectively.

In the three patients, the presence of alterations of the cochlea and of the VA was tested by high-resolution computed tomography (CT) of temporal bones in the coronal and axial planes (1 mm contiguous sections). The VA was considered enlarged when its diameter at the midpoint between the common crus and the external aperture was 1.5 mm or more on thin CT sections (16). High-resolution fast-spin echo (FSE) T2-weighted magnetic resonance imaging (MRI) was carried out (in axial

and coronal planes) to study the membranous labyrinth and in particular the endolymphatic duct and sac (ED and ES). ES is rarely seen in normal subjects, despite high-quality FSE MR images, and it is considered enlarged when the diameter exceeds 2.5 mm (17).

Molecular studies

DNA was extracted by standard methods from the whole blood of the three patients and their parents. In patient MC, all 20 coding SLC26A4 exons were amplified using primers flanking each exon, as previously described (1). The experimental studies were approved by the ethics committees of each Institution. Informed consent was obtained for all screened subjects.

Functional analysis

In order to evaluate the functional activity of the novel mutation found in the present family, the properties of pendrin transport were studied by a fluorimetric method in a human renal cell line (HEK293 Phoenix) in which the protein was overexpressed (21). To this purpose, a plasmid containing the mutated SLC26A4 sequence was generated by site-directed mutagenesis (Quik Change site-directed mutagenesis kit, Stratagene, La Jolla, CA, USA) starting from the SLC26A4_{WT} cDNA previously cloned and characterized (25). The following primers were used: forward: 5'-CCTTCTTGAATG-GACTTGGAATGGCCTTGGAAGCATCCCTAGC-3', and reverse: 5'-GCTAGGGATGCTTCCAAGGCCATTCCAAG-TCCATTCCAAGAAGG-3'.

Cell culture and transient transfection HEK 293 Phoenix cells were grown as previously described (21). For *in vivo* fluorometric chloride/iodide measurements, HEK 293 Phoenix cells were transiently transfected with wild type pendrin (pendrin_{WT}) or mutated pendrin showing the duplication (pendrin_{S523fsX548}), cloned into the pIRES2-EYFP vectors; cells transfected with the empty plasmid (pIRES2-EYFP) were used as negative control. The vector used in this study allows to coexpress at the same time pendrin or its mutant as well as EYFP, by transfecting only one plasmid. The bicistronic mRNA will produce two separated proteins in the same cell; in this way, the functional integrity of the protein under study will be maintained, and the transfection efficiency will be easily determined, as the transfected cells will show yellow-green fluorescence. The day before transfection, cells were seeded in poly-L-lysine-treated glass coverslip (Ø 40 mm), placed into Petri dishes (Ø 60 mm), and grown to 60–80% confluency. HEK 293 Phoenix cells were transfected by calcium–phosphate coprecipitation method: for each coverslip, 6 µg plasmid dissolved in 162 µl water were

mixed with 18 µl buffer A (2.5 M CaCl₂, pH 5.8) and 180 µl buffer B (140 mM NaCl, 1.5 mM Na₂HPO₄, 50 mM HEPES, pH 7.05 adjusted with NaOH). After incubation for 15–20 min at room temperature, the transfection mixture was spread over the cells. The experiments were performed 48 h after transfection.

Fluorometric analyses A fluorimetric method previously validated by our group was used to evaluate the chloride/iodide transport of SLC26A4 harboring the novel 11 bp duplication found in the family studied in the present work (26). Briefly, since the EYFP protein is a fluorescent dye sensitive to intracellular halide amount, fluorescence is measured before and after substituting extracellular chloride with iodide. Considering that iodide is a much better EYFP quencher than chloride, an increase of intracellular iodide should lead to a decrease of the EYFP fluorescence. In particular, to evaluate the pendrin-induced halide (chloride and iodide) transport, HEK 293 Phoenix cells transiently transfected with the vector containing either pIRES2-EYFP-pendrin_{WT} or the 11 bp duplication or with the empty plasmid, were continuously perfused in a laminar-flow chamber (FCS2 System, Biophtechs Inc., Beck Road Butler, PA, USA) with 'isotonic high Cl⁻' (2 mM KCl, 135 mM NaCl, 1 mM MgCl₂, 1 mM CaCl₂, 10 mM D-glucose, 20 mM HEPES, pH 7.4, 308 mOsm with mannitol) or 'isotonic high I⁻' (2 mM KCl, 135 mM NaI, 1 mM MgCl₂, 1 mM CaCl₂, 10 mM D-glucose, 20 mM HEPES, pH 7.4, 308 mOsm with mannitol) solutions. The used iodide concentrations, which are beyond physiological iodide concentrations, were chosen to enhance fluorescence sensitivity to ion exchange. The EYFP fluorescence measurements were performed using a Leica TCS SP2 AOBS confocal microscope (Leica Microsystem, Heidelberg, Germany) with a 515 nm Ar/ArKr laser beam for exciting the EYFP fluorescence. Traces representing fluorescence time course were obtained analyzing the regions of interest (ROI) within single cells. Measurements started after steady state conditions were reached. Maximal fluorescence variations (ΔF%) represent the maximal observed percentage difference with respect to the fluorescence intensity at the moment of the solution substitution. Statistical analysis was done by ANOVA–Bonferroni's multiple comparison test.

Results

Clinical data

All patients had a normal thyroid function under L-T4 replacement treatment (1.25, 1.5, and 1.6 µg/kg/day for

Figure 1 Radiological studies of the inner ear in patient MA. (A) CT scan of the petrous temporal bones (axial view). The vestibular aqueduct is bilaterally very enlarged (arrows). A cochlear dysplasia is also present. (B) MRI scan of the petrous temporal bones. The endolymphatic duct and sac, which are usually not visible in normal subjects, are greatly enlarged in both ears (arrows).

patient MM, MA, and MC, respectively). Anti-Tg and anti-TPO antibodies were negative in the three patients. Upon ultrasound examination, patient MM showed a small multinodular goiter (23 ml). Patient MA had a

normal TV (7.2 ml), without echographic pattern alterations and nodules, while patient MC had a large multinodular goiter (65.7 ml) with tracheal deviation. The perchlorate (KClO₄) test, performed by measuring radiolabeled iodide thyroid uptake before and after the oral administration of 1 g of KClO₄, was positive in all patients with discharge rates of 40–60% of basal radioiodine uptake.

In the three siblings, malformations of the inner ear were documented. At CT scan, the VA was bilaterally enlarged, with a diameter ranging from 2 to 4 mm. The ED and ES resulted as enlarged in all patients at MRI examination, with a ES diameter ranging from 4 to 11 mm. Patient MA also had a dysplastic cochlea (Mondini malformation), which was not found in patients MM and MC (Fig. 1).

Mutation analysis

SLC26A4 sequence analyses showed a compound heterozygous pattern in the three patients: a known common missense mutation in exon 10 (1226 GA) and a novel 11 bp duplication in exon 14 (1561_1571dupCTTGGAATGGC; Fig. 2). The first mutation leads to the substitution of arginine for histidine at codon 409 (R409H) and the second leads to a frameshift with the

Figure 2 The sequences corresponding to a wild type and a mutated allele are reported. The nucleotide and aminoacidic sequences are also shown.

creation of a stop codon at 548. According to the nomenclature used, the latter mutation is denoted as S523fsX548 (27). Both mutations were inherited from the non-consanguineous parents. Whereas the mother is heterozygous for the 11 bp duplication mutation (S523fsX548), the father is heterozygous for the missense mutation (R409H; Fig. 3).

Functional analyses

After expressing pendrin_{S523fsX548}, an impaired Cl^-/I^- exchange can be documented in HEK 293 Phoenix cells (Fig. 4A). The intracellular fluorescence after iodide substituting for chloride in the bath solution (135 mM iodide instead of chloride) is decreased if wild type pendrin (pendrin_{WT}) is expressed. Conversely, following the same protocol, after the expression of pendrin_{S523fsX548} no decrease in fluorescence can be observed. A similar result was obtained when the empty vector was used for transfection (in both latter cases a slight increase of fluorescence can be observed instead of a decrease; Fig. 4B)

Returning to the initial conditions (i.e. 135 mM chloride instead of iodide in the bathing solution) results in an increase in fluorescence (positive $\Delta F\%$) for pendrin_{WT} as a consequence of the diminished quenching of EYFP that follows the iodide efflux in

exchange with chloride. Control and pendrin_{S523fsX548} behave differently, continuing to show a slight increase in fluorescence as already observed when iodide substituted for chloride. The observed increase possibly reflects some osmotic perturbation induced by solution substitutions leading to an aspecific anion leakage and/or cell shrinkage.

Discussion

Mutations in SLC26A4 appear to be responsible for a number of overlapping clinical conditions ranging from nonsyndromic hearing loss (familial EVA syndrome or DFNB4) with widened VAs to typical PS with evident thyroid signs. The majority of SLC26A4 variants are missense substitutions, while a smaller subset consists of nonsense mutations with or without frameshift, and there are several mutations affecting splice donor or acceptor sites.

The present study reports the functional characterization of a novel 11 bp duplication (resulting in S523fsX548), which was found in compound heterozygosity with a missense mutation leading to substitution of R409H. This duplication creates a frameshift with stop codon at 548 and the deriving truncated protein lacks the entire carboxy-terminus

Figure 3 Clinical and molecular features of the present Pendred syndrome family. Affected members are indicated as black symbols. The age of onset of thyroid and inner ear alterations are indicated. Electropherogram corresponds to SLC26A4 11 bp duplication identified in compound heterozygosity in the three affected siblings and in heterozygosity in the unaffected mother.

Figure 4 HEK 293 Phoenix cells expressing pendrin_{S523fsX548} have an impaired I⁻ transport. (A) Relative fluorescence intensity values were obtained analyzing the ROI within single cells transfected with pIRES2-EYFP-pendrin_{WT} ($n=7$), pIRES2-EYFP-pendrin_{S523fsX548} ($n=8$), or the empty ($n=16$) plasmids and exposed to the 'high Cl⁻' or the 'high I⁻' solutions. Cells transfected with pendrin_{S523fsX548} serve as the control cells. At the top of each column, the respective changes of the extracellular solutions (isotonic high Cl⁻ or isotonic high I⁻) are indicated. (B) Maximal fluorescence % variation after the ionic substitution is calculated as the average of different ROI inside the cells (pendrin_{WT} $n=7$; pendrin_{S523fsX548} $n=8$, empty vector $n=16$). In every panel, the maximal fluorescence variation of the pendrin_{WT} is significantly different from the pendrin_{S523fsX548} and the empty vector (calculated by ANOVA–Bonferroni's multiple comparison test).

domain. Indeed, functional studies demonstrated an impaired transport of chloride and iodide in HEK 293 Phoenix cells expressing S523fsX548 when compared with pendrin_{WT}. In particular, after exchanging extracellular chloride for the same amount of iodide and vice-versa, in cells expressing pendrin_{WT} the EYFP fluorescence variations were rapid and reversible, contrary to what was observed for both pendrin_{S523fsX548} and control. These data are consistent with the phenotype observed in the respective patients. The missense mutation R409H has been associated with PS in several cohorts and has been shown to impair pendrin function (28–31), in agreement with the phenotype resulting from the compound heterozygosis described in the affected members.

The present family is of particular interest, since it displays a high phenotypic variability. Indeed, despite the presence of major inner ear malformations responsible for a profound SNHL, the age of onset and the progression of deafness were significantly different in the three patients tested. Patient MM had the first diagnosis of SNHL at age 5 with a progressive worsening

in the following years up to a profound deficit, and patient MA was diagnosed with SNHL at 18 months and also showed a progressive worsening up to a profound hearing deficit. In contrast, patient MC experienced the sudden appearance of a profound SNHL at the age of 11. All three siblings had a comparable degree of EVA, excluding the hypothesis that the severity of the inner ear malformation could be a major determinant of the observed variability. Similarly, the diagnosis of hypothyroidism was made during childhood in the two male patients and at age 21 for their sister. The respective TVs range from normal (MA) to large multinodular goiters (MC). Interestingly, patient MM suffers from a renal disease and has been submitted to kidney transplantation and patient MA has a low IQ and language disturbances despite early start of replacement therapy at the diagnosis of subclinical hypothyroidism.

Intrafamilial and interfamilial phenotypic variability, mostly in terms of presence/absence and severity of thyroid and inner ear malformations, have been reported by other groups (12–14, 32). In the family tested in the present work, the extreme variability can not only be found in the degree of hearing impairment, but also in the time of onset for the thyroid alterations. Moreover, the evidence of a renal dysfunction in one patient is particularly interesting because this alteration in PS is seldom seen, however expected, since the SLC26A4 transporter is involved in kidney acid–base regulation and blood volume maintenance (33–35). However, no evidence exists about a direct cause–effect relationship between impaired pendrin function and kidney atrophy, and this finding could be a mere coincidence. The reason why the three tested family members suffering from PS show this pronounced phenotypic variability is unknown. The degree of iodide deficiency has been supposed to be a modifying factor in PS (36), but other environmental or genetic factors are also likely to be involved. These include the action of modifier genes that indeed have been demonstrated to play an important role in disease variability in other autosomal disorders (37).

In conclusion, the identification of novel complex mutation expands the spectrum of mutations in the SLC26A4 gene. The impairment of chloride and iodide transport recorded for the mutant further supports the hypothesis that pendrin could play a crucial role in iodide transport from the thyroid cell to the colloid. The clinical studies on the present family confirm that the PS phenotype is extremely variable, both for the degree of impairment and the time of onset, the inner ear malformations being the only constant phenotypic feature.

References

- 1 Everett LA, Glaser B, Beck JC, Idol JR, Buchs A, Heyman M, Adawi F, Hazani E, Nassir E, Baxevanis AD, Sheffield VC & Green ED. Pendred syndrome is caused by mutations in a putative sulphate transporter gene (PDS). *Nature Genetics* 1997 **17** 411–422.

- 2 Bidart JM, Mian C, Lazar V, Russo D, Filetti S, Caillou B & Schlumberger M. Expression of pendrin and the Pendred syndrome (PDS) gene in human thyroid tissue. *Journal of Clinical Endocrinology and Metabolism* 2000 **85** 2028–2033.
- 3 Everett LA, Morsli H, Wu DK & Green ED. Expression pattern of the mouse ortholog of the Pendred's syndrome gene (*Pds*) suggests a key role for pendrin in the inner ear. *PNAS* 1999 **96** 9727–9732.
- 4 Royaux IE, Wall SM, Karniski LP, Everett LA, Suzuki K, Knepper MA & Green ED. Pendrin, encoded by the Pendred syndrome gene, resides in the apical region of renal intercalated cells and mediates bicarbonate secretion. *PNAS* 2001 **98** 4221–4226.
- 5 Rillema JA & Hill MA. Prolactin regulation of the pendrin-iodide transporter in the mammary gland. *American Journal of Physiology, Endocrinology and Metabolism* 2003 **284** E25–E28.
- 6 Suzuki K, Royaux IE, Everett LA, Mori-Aoki A, Suzuki S, Nakamura K, Sakai T, Katoh R, Toda S, Green ED & Kohn LD. Expression of PDS/Pds, the Pendred syndrome gene, in endometrium. *Journal of Clinical Endocrinology and Metabolism* 2002 **87** 938.
- 7 Lacroix L, Mian C, Caillou B, Talbot M, Filetti S, Schlumberger M & Bidart JM. Na(+)/I(−) symporter and Pendred syndrome gene and protein expressions in human extra-thyroidal tissues. *European Journal of Endocrinology* 2001 **144** 297–302.
- 8 Royaux IE, Suzuki K, Mori A, Kato R, Everett LA, Kohn LD & Green ED. Pendrin, the protein encoded by the Pendred syndrome gene (PDS), is an apical porter of iodide in the thyroid and is regulated by thyroglobulin in FRTL-5 cells. *Endocrinology* 2000 **141** 839–845.
- 9 Gillam MP, Sidhaye AR, Lee EJ, Rutishauser J, Stephan CW & Kopp P. Functional characterization of pendrin in a polarized cell system. Evidence for pendrin-mediated apical iodide efflux. *Journal of Biological Chemistry* 2004 **279** 13004–13010.
- 10 Pendred V. Deaf-mutism and goiter. *Lancet* 1896 **11** 532.
- 11 Lopez-Bigas N, Rabionet R, de Cid R, Govea N, Gasparini P, Zelante L, Arbones ML & Estivill X. Splice-site mutation in the *PDS* gene may result in intrafamilial variability for deafness in Pendred syndrome. *Human Mutation* 1999 **14** 520–526.
- 12 Masmoudi S, Charfedine I, Hmani M, Grati M, Ghorbel AM, Elgaied-Boulila A, Drira M, Hardelin JP & Ayadi H. Pendred syndrome: phenotypic variability in two families carrying the same PDS missense mutation. *American Journal of Medical Genetics* 2000 **90** 38–44.
- 13 Campbell C, Cucci RA, Prasad S, Green GE, Edeal JB, Galer CE, Karniski LP, Sheffield VC & Smith RJ. Pendred syndrome, DFNB4, and PDS/SLC26A4 identification of eight novel mutations and possible genotype–phenotype correlations. *Human Mutation* 2001 **17** 403–411.
- 14 Napiontek U, Borck G, Muller-Forell W, Pfarr N, Bohnert A, Keilmann A & Pohlenz J. Intrafamilial variability of the deafness and goiter phenotype in Pendred syndrome caused by a T416P mutation in the *SLC26A4* gene. *Journal of Clinical Endocrinology and Metabolism* 2004 **89** 5347–5351.
- 15 Bos H, Feldmann D, Duval V, Messaz O, Denoyelle F, Loundon N, Sergout-Allaoui A, Houang M, Duriez F, Lacombe D, Delobel B, Lema J, Catros H, Journel H, Doruin-Garraud V, Obstoy MF, Toutain A, Odent S, Toubanc JE, Couderc R, Petit C, Garabedian EN & Marlin S. Screening of SLC26A4 (*PDS*) gene in Pendred's syndrome: a large spectrum of mutations in France and phenotypic heterogeneity. *Clinical Genetics* 2004 **66** 333–340.
- 16 Phelps PD, Coffey RA, Trembath RC, Luxon LM, Grossman AB, Britton KE, Kendall-Taylor P, Graham JM, Cadge BC, Stephens SGD, Pembrey ME & Reardon W. Radiological malformations of the ear in Pendred syndrome. *Clinical Radiology* 1998 **53** 268–273.
- 17 Fugazzola L, Mannavola D, Cerutti N, Maghnie M, Pagella F, Bianchi P, Weber G, Persani L & Beck-Peccoz P. Molecular analysis of the Pendred's syndrome gene and magnetic resonance imaging studies of the inner ear are essential for the diagnosis of true Pendred' syndrome. *Journal of Clinical Endocrinology and Metabolism* 2000 **85** 2469–2475.
- 18 Fugazzola L, Cerutti N, Mannavola D, Vannucchi G & Beck-Peccoz P. The role of pendrin in iodide regulation. *Experimental and Clinical Endocrinology and Diabetes* 2001 **109** 18–22.
- 19 Scott DA, Wang R, Kreman TM, Sheffield VC & Karnishki LP. The Pendred syndrome gene encodes a chloride–iodide transport protein. *Nature Genetics* 1999 **21** 440–443.
- 20 Wolff J. What is the role of Pendrin? *Thyroid* 2005 **15** 346–348.
- 21 Dossena S, Vezzoli V, Cerutti N, Bazzini C, Tosco M, Sironi C, Rodighiero S, Meyer G, Fasci U, Furst J, Ritter M, Fugazzola L, Persani L, Zorowka P, Storelli C, Beck-Peccoz P, Bottà G & Paulmichl M. Functional characterization of wild-type and a mutated form of SLC26A4 identified in a patient with Pendred syndrome. *Cellular Physiology and Biochemistry* 2006 **17** 245–256.
- 22 Taylor JP, Metcalfe RA, Watson PF, Weetman AP & Trembath RC. Mutations of the *PDS* gene, encoding pendrin, are associated with protein mislocalization and loss of iodide efflux: implications for thyroid dysfunction in Pendred syndrome. *Journal of Clinical Endocrinology and Metabolism* 2002 **87** 1778–1784.
- 23 Soleimani M, Greeley T, Petrovic S, Wang Z, Amlal H, Kopp P & Burnham CE. Pendrin: an apical Cl[−]/OH[−]/HCO₃[−] exchanger in the kidney cortex. *American Journal of Physiology, Renal Physiology* 2001 **280** F356–F364.
- 24 Li XC, Everett LA, Lalwani AK, Desmukh D, Friedman TB, Green ED & Wilcox ER. A mutation in PDS causes non-syndromic deafness. *Nature Genetics* 1998 **18** 215–217.
- 25 Dossena S, Maccagni A, Vezzoli V, Bazzini C, Garavaglia ML, Meyer G, Furst J, Ritter M, Fugazzola L, Persani L, Zorowka P, Storelli C, Beck-Peccoz P, Bottà G & Paulmichl M. The expression of wild-type pendrin (SLC26A4) in human embryonic kidney (HEK 293 Phoenix) cells leads to the activation of cationic currents. *European Journal of Endocrinology* 2005 **153** 693–699.
- 26 Dossena S, Rodighiero S, Vezzoli V, Bazzini C, Sironi C, Meyer G, Furst J, Ritter M, Garavaglia ML, Fugazzola L, Persani L, Zorowka P, Storelli C, Beck-Peccoz P, Bottà G & Paulmichl M. Fast fluorimetric method for measuring pendrin (SLC26A4) Cl/I transport activity. *Cellular Physiology and Biochemistry* 2006 **18** 67–74.
- 27 den Dunnen JT & Antonarakis E. Nomenclature for the description of human sequence variations. *Human Genetics* 2001 **109** 121–124.
- 28 Fugazzola L, Cerutti N, Mannavola D, Crinò A, Cassio A, Gasparoni P, Vannucchi G & Beck-Peccoz P. Differential diagnosis between Pendred's and 'pseudo-Pendred's' syndromes: clinical, radiological and molecular studies. *Pediatric Research* 2002 **51** 479–484.
- 29 Bogazzi F, Russo D, Raggi F, Ultimieri F, Berrettini S, Forli F, Grasso L, Ceccarelli C, Mariotti S, Pinchera A, Bartalena L & Martino E. Mutations in the SLC26A4 (pendrin) gene in patients with sensorineural deafness and enlarged vestibular aqueduct. *Journal of Endocrinological Investigation* 2004 **27** 430–435.
- 30 Van Hauwe P, Everett LA, Coucke P, Scott DA, Kraft ML, Ris-Stalpers C, Bolder C, Otten B, de Vijlder JJ, Dietrich NJ, Ramesh A, Srisailapathy SC, Parving A, Cremers CW, Willems PJ, Smith RJ, Green ED & Van Camp G. Two frequent missense mutations in Pendred syndrome. *Human Molecular Genetics* 1998 **7** 1099–1104.
- 31 Gillam MP, Bartolone L, Kopp P & Benvenega S. Molecular analysis of the *PDS* gene in a nonconsanguineous Sicilian family with Pendred's syndrome. *Thyroid* 2005 **15** 734–741.
- 32 Reardon W, Coffey R, Phelps PD, Luxon LM, Stephens D, Kendall-Taylor P, Britton KE, Grossman A & Trembath R. Pendred syndrome: 100 years of underascertainment? *Quarterly Journal of Medicine* 1997 **90** 443–447.
- 33 de Seigneux S, Malte H, Dimke H, Frokiaer J, Nielsen S & Frische S. Renal compensation to chronic hypoxic hypercapnia: down-regulation of pendrin and adaptation of the proximal tubule. *American Journal of Physiology, Renal Physiology* 2007 **292** F1256–F1266.
- 34 Pech V, Kim YH, Weinstein AM, Everett LA, Pham TD & Wall SM. Angiotensin II increases chloride absorption in the cortical

- collecting duct in mice through a pendrin-dependent mechanism. *American Journal of Physiology. Renal Physiology* 2007 **292** F914–F920.
- 35 Soleimani M & Xu J. SLC26 chloride/base exchangers in the kidney in health and disease. *Seminars in Nephrology* 2006 **26** 375–385.
- 36 Kopp P, Karamanoglu Arseven O, Sabacan L, Kotlar T, Dupuis J, Cavaliere H, Santos CLS, Jameson JL & Medeiros-Neto G. Phenocopies for deafness and goiter development in a large inbred brazilian kindred with Pendred's syndrome associated with a novel mutation in the *PDS* gene. *Journal of Clinical Endocrinology and Metabolism* 1999 **84** 336–341.
- 37 Zielenski J, Corey M, Rozmahel R, Markiewicz D, Aznarez I, Casals T, Larriba S, Mercier B, Cutting GR, Krebsova A, Macek M Jr, Langfelder-Schwind E, Marshall BC, DeCelle-Germana J, Claustres M, Palacio A, Bal J, Nowakowska A, Ferec C, Estivill X, Durie P & Tsui LC. Detection of a cystic fibrosis modifier locus for meconium ileus on human chromosome 19q13. *Nature Genetics* 1999 **22** 128–129.

Received 20 April 2007

Accepted 26 June 2007