Vitamin D status and parathyroid hormone in obese children before and after weight loss

Thomas Reinehr, Gideon de Sousa, Ute Alexy 1, M Kersting 1 and Werner Andler
Vestische Hospital for Children and Adolescents Datteln, University of Witten/Herdecke, Dr F Steiner Strasse 5, 45711 Datteln, Germany and 1 Research Institute of Child Nutrition Dortmund, Institute at the Rheinische Friedrich-Wilhelms-University Bonn, Dortmund, Germany
(Correspondence should be addressed to T Reinehr; Email: t.reinehr@kinderklinik-datteln.de)

Abstract
Objective: The roles of vitamin D and parathyroid hormone (PTH) are discussed controversially in obesity, and studies of these hormones in obese children are limited. Therefore, we studied the relationships between PTH, 1,25-dihydroxy-vitamin D (1,25-OH Vit D), 25-hydroxy-vitamin D (25-OH Vit D), weight status, and insulin sensitivity before and after weight loss in obese children.

Methods: Fasting serum PTH, 1,25-OH Vit D, 25-OH Vit D, inorganic phosphate, calcium, alkaline phosphatase (AP), insulin, glucose, and weight status (SDS–BMI and percentage body fat) were determined in 133 obese children (median age 12.1 years) and compared with 23 non-obese children. Furthermore, these parameters were analyzed in 67 obese children before and after participating in a 1-year obesity intervention program.

Results: Obese children had significantly (P<0.001) higher PTH and lower 25-OH Vit D concentrations compared with non-obese children, while calcium, phosphate, AP, and 1,25-OH Vit D did not differ significantly. Changes of PTH (r=0.23, P=0.031) and 25-OH Vit D (r=−0.27, P=0.013) correlated significantly with changes of SDS–BMI, but not with changes of insulin sensitivity (homeostasis model assessment; HOMA-B%). Reduction of overweight in 35 children led to a significant (P<0.01) decrease of PTH concentrations and an increase in 25-OH Vit D levels.

Conclusions: PTH levels were positively and 25-OH Vit D concentrations were negatively related to weight status. Since these alterations normalized after weight loss, these changes are consequences rather than causes of overweight. A relationship between PTH, vitamin D, and insulin sensitivity based on the HOMA index was not found in obese children. Further longitudinal clamp studies are necessary to study the relationship between vitamin D and insulin sensitivity.

Introduction
Vitamin D and parathyroid hormone (PTH) are well known for their essential role in bone metabolism and calcium homeostasis. The main sources of vitamin D are ergocalciferol and cholecalciferol, the former normally available in food and the latter produced in the skin by u.v. radiation of 7-dehydrocholesterol. Both of these compounds are hydroxylated in the liver to 25-hydroxyvitamin D (25-OH Vit D), which is the major circulating metabolite precursor to the hormonally active form, 1,25-dihydroxy-vitamin D (1,25-OH Vit D).

It has become increasingly clear that the vitamin D endocrine system is related to obesity in adults. Obesity has been found to be associated with lower levels of serum 25-OH Vit D(1–5) and higher levels of serum PTH(1, 5–8). A low vitamin D intake was associated with increased body mass index (BMI) (9). PTH has been postulated as an independent predictor of obesity (7). Overweight as a consequence of elevated serum PTH was explained by several mechanisms (10, 11): PTH stimulates the renal hydroxylation of 25-OH Vit D to its active form, 1,25-OH Vit D, which in turn elevates the calcium influx into adipocytes. In these cells, intracellular calcium enhances lipogenesis through the activation of fatty acid synthase and inhibits lipolysis via activation of phosphodiesterase 3B, which subsequently reduces catecholamine-induced lipolysis (11–13). Both these effects would promote lipid storage in fat tissue. Additionally, studies support a direct role for PTH in suppressing lipid oxidation in the muscle (14). However, these hypotheses are discussed controversially since in obese adults with weight loss, increasing and decreasing 25-OH Vit D as well as decreasing and increasing PTH concentrations have been reported (15–20), so that the question whether the alterations of these hormones are a consequence or cause of overweight remains open.

Vitamin D also acts as a necessary cofactor for insulin secretion (21–24). Vitamin D repletion improves insulin
sensitivity and insulin secretion in animal studies (25). Hypovitaminosis D has been proposed as a risk factor for reduced insulin secretion, impaired glucose tolerance, and type 2 diabetes in adults (23–29). However, some studies in obese adults demonstrated no relationship between vitamin D, PTH, and insulin sensitivity (30–32).

As data concerning the vitamin D endocrine system in obesity and its relation to insulin sensitivity are controversial, and studies in children are lacking, we studied obesity and its relation to insulin sensitivity before and after participating in a 1-year obesity intervention program and compared them with normal weight children. The aims of this study were 1) to analyze the alterations of these hormones in obese children, 2) to prove their reversibility in weight loss pointing towards a consequence of overweight rather than a cause, and 3) to study the relationship between insulin sensitivity and vitamin D status.

Methods

We examined anthropometrical markers (weight, height, and skinfold thickness) as well as fasting serum PTH, 1,25-OH Vit D, 25-OH Vit D, calcium, inorganic phosphate alkaline phosphatase (AP), glucose, and insulin concentrations in 133 obese Caucasian children and in 23 healthy Caucasian normal weight children. Additionally, these parameters were determined in a subgroup of 67 obese children before and after participating in the same 1-year outpatient obesity intervention program. The 66 obese children who did not participate in the intervention had failed to prove their motivation by performing a 3-day dietary record, filling out a questionnaire concerning their eating and exercise habits, and participating regularly in an exercise therapy for overweight children for a period of 8 weeks (33). All analyses were performed in spring. Children with endocrine disorders, premature adrenarche, or syndromal obesity were excluded from the study. All participants were non-smokers without any regular medication. Obesity was defined according to International Task Force of Obesity (34).

Blood sampling was performed in the fasting state at 0800 h. 1,25-OH Vit D was determined by highly specific RIA (immunoextraction followed by quantification by 125I RIA, Gamma B 1,25-OH Vit D RIA Immunodiagnostics System, Boldon, UK) with cross-reaction to 25-OH Vit D of 0.001%. The sensitivity of this assay was 3.4 pg/ml. 25-OH Vit D was determined by a high-specific chemiluminescence-immunassay (LIAISON 25-OH-Vit D Assay, Diasorin, Dietzenbach, Germany) with cross-reaction to 1,25-OH Vit D of 1.1%. The sensitivity of this assay was 2.0 ng/ml. The normal range of this test kit for the 25-OH Vit D concentrations was 5–64 ng/ml (35). Intact PTH was determined by a highly specific solid-phase two-site chemiluminescent enzyme-labeled immunometric assay using an Immulite analyzer (DPC Biermann, Bad Nauheim, Germany). Insulin concentrations were measured by microparticle-enhanced immunometric assay (Abbott). Glucose levels were determined by colorimetric test using a Vitros analyzer (Ortho Clinical Diagnostics, Neckargmuend, Germany). Calcium, Inorganic phosphate, and AP were measured by commercially available test kits (Calcium Cobas, Inorganic Phosphate Cobas, AP IFCC Cobas, Roche Diagnostics). Intra- and interassay coefficients of variation were <14% for PTH, 25-OH Vit D, and 1,25-OH Vit D and <5% for all other methods. Homeostasis model assessment (HOMA) was used to detect the degree of insulin resistance and insulin sensitivity (36): the resistance can be assessed from the fasting glucose and insulin concentrations by the formula: resistance (HOMA–IR)=(fasting insulin (mU/I)×fasting glucose (mmol/l))/22.5. Insulin sensitivity (HOMA-B%) was computed as (20×fasting insulin in mU/I)/(fasting glucose in mmol/l − 3.5).

Height was measured to the nearest centimeter using a rigid stadiometer. Weight was measured unclothed to the nearest 0.1 kg using a calibrated balance scale. Since BMI is not normally distributed in childhood, we used the LMS method to calculate SDS–BMI as a measurement for the degree of overweight. This method summarizes the data in terms of three smooth age-specific curves termed L (λ), M (μ), and S (σ) based on German population-specific data (37, 38). The M and S curves correspond to the median and coefficient of variation body mass index for German children at each age and gender, whereas the L curve allows for the substantial age-dependent skewness in the distribution of body mass index (37, 38). The assumption behind the LMS method is that after Box–Cox power transformation, the data at each age are distributed normally (37). Reduction of overweight was defined by a reduction in SDS–BMI, since BMI is age and gender dependent in childhood.

Tricep and subscapularis skinfold thickness was measured by one investigator in duplicate using a caliper and averaged to calculate the percentage of body fat, using a skinfold thickness equation with the following formulas (39):

Boys, body fat %
= 0.783 × (subscapularis skinfold thickness + tricep skinfold thickness in mm) + 1.6

Girls, body fat %
= 0.546 × (subscapularis skinfold thickness + tricep skinfold thickness in mm) + 9.7

The pubic stage was determined by inspection and palpation according to Marshall and Tanner. Pubertal developmental stage was categorized into two groups.
Vitamin D in obese children

Results

The anthropometrical data of the 133 obese and 23 normal weight children are presented in Table 1. The 133 obese children demonstrated significantly increased PTH concentrations ($P<0.001$) and decreased 25-OH Vit D levels compared to the 23 normal weight children, while the children did not differ in their 1,25-OH Vit D concentrations (Fig. 1). Four of the obese and none of the non-obese children demonstrated 25-OH Vit D levels below the normal range for German children. Furthermore, the obese and normal weight children did not differ in their calcium (2.3 ± 0.1 vs 2.3 ± 0.1 mmol/l, $P=0.727$), inorganic phosphate (4.6 ± 0.6 vs 4.8 ± 0.3 mg/dl, $P=0.095$), or AP levels (212 ± 87 vs 212 ± 51 U/l, $P=0.989$).

At baseline, PTH correlated significantly positively to SDS–BMI ($r=0.25$, $P=0.008$) and percentage body fat ($r=0.25$, $P=0.017$), negatively to 25-OH Vit D ($r=−0.23$, $P=0.004$), but not significantly to 1,25-OH Vit D, age, insulin, HOMA-B%, or HOMA-IR. 25-OH Vit D correlated significantly negatively to SDS–BMI ($r=−0.34$, $P<0.001$) and percentage body fat ($r=−0.33$, $P=0.005$), but not significantly to 1,25-OH Vit D, age, insulin, HOMA-B%, or HOMA-IR. 1,25-OH Vit D correlated significantly to age ($r=0.18$, $P=0.013$), but not significantly to SDS–BMI, percentage body fat, insulin, HOMA-B%, or HOMA-IR. In multiple linear regression analyses adjusted for age, gender, pubertal stage and weight status (BMI), no significant correlation between insulin and PTH, 25-OH Vit D, and 1,25-OH Vit D could be observed.

The changes in weight status (SDS–BMI) of the 67 obese children between baseline and 1 year later correlated significantly to changes in PTH ($r=0.23$, $P=0.031$) and 25-OH Vit D ($r=−0.27$, $P=0.013$), but not significantly to changes in 1,25-OH Vit D ($r=−0.03$, $P=0.389$; Fig. 2). Changes in insulin, HOMA-IR, and HOMA-B% did not significantly correlate to the changes of PTH, 25-OH Vit D, and 1,25-OH Vit D in partial correlation adjusted for changes of weight status (SDS–BMI).

Table 1  Anthropometrical data of the obese and normal weight children.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Obese</th>
<th>Normal weight</th>
<th>$P$ value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>133</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>12.1 ± 2.4</td>
<td>11.9 ± 1.9</td>
<td>0.661</td>
</tr>
<tr>
<td>Gender</td>
<td>51% male</td>
<td>52% male</td>
<td>0.926</td>
</tr>
<tr>
<td>Pubertal stage</td>
<td>36% prepubertal</td>
<td>35% prepubertal</td>
<td>0.988</td>
</tr>
<tr>
<td>BMI</td>
<td>27.7 ± 3.8</td>
<td>17.9 ± 3.0</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>SDS–BMI</td>
<td>2.2 ± 0.4</td>
<td>−0.4 ± 1.0</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>
The changes in HOMA-IR, HOMA-B%, insulin, glucose, PTH, 25-Vit D, and 1,25-OH Vit D concentrations between baseline and 1 year later in the 35 obese children with reduction of overweight and the 32 obese children without reduction of overweight are shown in Table 2. The reduction of overweight led to a significant decrease of PTH levels and insulin resistance index (HOMA-IR), as well as an increase of 25-OH Vit D concentrations, while 1,25-OH Vit D levels did not change significantly. In the obese children without reduction of overweight, no significant changes occurred apart from a significant increase of insulin concentrations, insulin resistance index (HOMA-IR) and percentage body fat.

At baseline, no significant differences were observed in age, gender, pubertal stage, SDS–BMI, and percentage body fat between the obese children with and without weight loss (Table 2). Furthermore, there were no significant differences regarding PTH, 25-OH Vit D, or 1,25-OH Vit D levels between the obese children with and those without reduction of overweight.

Figure 1 Parathyroid hormone (A), 25-hydroxy-vitamin D (B), and 1,25-dihydroxy-vitamin D (C) in 133 obese and 23 normal weight children.

Figure 2 Changes in parathyroid hormone ((A), r = 0.23, P = 0.031), 25-hydroxy-vitamin D ((B), r = -0.27, P = 0.013) and 1,25-dihydroxy-vitamin D ((C), r = -0.03, P = 0.389) in relation to change of weight status (SDS-BMI) in the course of 1 year.
Table 2 Changes in weight status, percentage body fat, insulin resistance index (homeostasis model assessment; HOMA-IR), insulin sensitivity (HOMA-B%), insulin, glucose, lipids, parathyroid hormone, 25-hydroxy-vitamin D (25-OH Vit D), and 1,25 OH- Vit D concentrations in 67 obese children with and without reduction of overweight over a 1-year period.

<table>
<thead>
<tr>
<th></th>
<th>Reduction of overweight</th>
<th>No reduction of overweight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>35</td>
<td>32</td>
</tr>
<tr>
<td>Age (years)</td>
<td>11.1 ± 2.3</td>
<td>11.0 ± 2.5</td>
</tr>
<tr>
<td>Gender</td>
<td>51% male</td>
<td>59% male</td>
</tr>
<tr>
<td>Pubertal stage</td>
<td>46% prepubertal</td>
<td>43% prepubertal</td>
</tr>
<tr>
<td>Change of SDS–BMI</td>
<td>−0.3 ± 0.1</td>
<td>+0.1 ± 0.2</td>
</tr>
<tr>
<td></td>
<td>At baseline</td>
<td>At baseline</td>
</tr>
<tr>
<td></td>
<td>1 year later</td>
<td>1 year later</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>27.3 ± 3.5</td>
<td>26.4 ± 3.0†</td>
</tr>
<tr>
<td>SDS–BMI</td>
<td>2.3 ± 0.4</td>
<td>2.0 ± 0.3†</td>
</tr>
<tr>
<td>Subscapularis ST (mm)</td>
<td>30 ± 7</td>
<td>26 ± 7†</td>
</tr>
<tr>
<td>Triceps ST (mm)</td>
<td>31 ± 6</td>
<td>26 ± 6†</td>
</tr>
<tr>
<td>Percentage body fat (%)</td>
<td>46 ± 12</td>
<td>40 ± 11†</td>
</tr>
<tr>
<td>Insulin (mU/l)</td>
<td>19 ± 14</td>
<td>13 ± 7*</td>
</tr>
<tr>
<td>Glucose (mg/dl)</td>
<td>88 ± 4</td>
<td>86 ± 6</td>
</tr>
<tr>
<td>HOMA-β%</td>
<td>226 ± 121</td>
<td>286 ± 108</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>4.0 ± 2.9</td>
<td>2.8 ± 1.4*</td>
</tr>
<tr>
<td>PTH (pg/ml)</td>
<td>29 ± 9</td>
<td>20 ± 6†</td>
</tr>
<tr>
<td>1,25-OH Vit D (pg/ml)</td>
<td>56 ± 14</td>
<td>55 ± 9</td>
</tr>
<tr>
<td>25-OH Vit D (ng/ml)</td>
<td>11 ± 4</td>
<td>16 ± 9†</td>
</tr>
<tr>
<td>Calcium (mmol/l)</td>
<td>2.3 ± 0.1</td>
<td>2.3 ± 0.1</td>
</tr>
<tr>
<td>Inorganic phosphate (mg/dl)</td>
<td>4.8 ± 0.5</td>
<td>4.7 ± 0.3</td>
</tr>
<tr>
<td>Alkaline phosphatase (U/l)</td>
<td>194 ± 49</td>
<td>194 ± 21</td>
</tr>
</tbody>
</table>

Data as median and interquartile range.
*P<0.05 baseline versus 1 year later; †P<0.01 baseline versus 1 year later; ST, skinfold thickness.

Fourteen (40%) children with reduction of overweight and 20 (63%) children without reduction of overweight did not perform a 3-day dietary record after the end of intervention. These children did not differ significantly from the children with complete dietary records with respect to age, gender, and degree of overweight at baseline. Furthermore, the children with or without complete dietary records did not differ significantly in their PTH, 25-OH Vit D, or 1,25-OH Vit D concentrations. In the children with complete dietary records, caloric intake and percentage fat content in diet decreased significantly, while percentage carbohydrate content in diet increased significantly regardless of weight loss (see Table 3). Calcium intake did not change significantly in the children with or without weight loss. Vitamin D intake decreased in the children without reduction of overweight, while vitamin D intake remained stable (P=0.244) in the children with reduction of overweight. At baseline, there were no significant differences in diet composition with respect to fat, carbohydrate and protein content, as well as in caloric, vitamin D, and calcium intake between the children with and without overweight reduction.

Discussion

This is the first study analyzing cross-sectional and longitudinal relationships between the vitamin D endocrine system, weight status, and insulin sensitivity in childhood. We were able to demonstrate that obese children had significantly higher PTH levels and lower 25-OH Vit D concentrations as compared with non-obese children. PTH levels decreased and 25-OH Vit D concentrations increased significantly in obese children, who achieved a reduction of overweight in the course of 1 year in contrast to obese children without reduction of overweight. Therefore, the alterations of these hormones are a consequence rather than a cause of overweight.

The low levels of 25-OH Vit D in obese children in our study are in concordance with most studies in adults (1–5, 15, 43). A good vitamin D status has been postulated to prevent obesity (11, 27, 29, 44). Since 25-OH Vit D concentrations increased after reduction of overweight, this finding points towards a consequence rather than a cause of overweight. Since the main source of vitamin D in Germany is alimentary intake rather than a cause of overweight. Since the main source of vitamin D in Germany is alimentary intake due to the geographical latitude, a change of vitamin D intake during the intervention may have caused the increase of 25-OH Vit D concentrations. However, the 3-day weighed dietary records did not demonstrate an increase in vitamin D consumption. The higher vitamin D levels in normal weight subjects seem to be more a surrogate parameter for healthy nutrition than a real causal factor in the prevention of obesity. Intervention studies in obese children with vitamin D supplementation are necessary to prove this hypothesis. Furthermore, the low levels of 25-OH Vit D may be attributed to several other factors such as decreased exposure to sunlight in obese subjects due to limited exposure to sunlight in obese subjects due to limited
mobility, clothing habits, or the excessive deposition of vitamin D in adipose tissue (1, 3, 4). The increase in serum 25-OH Vit D after sun exposure was 57% less in obese compared with non-obese subjects (45). Additionally, adipose tissue is a major storage site of vitamin D (46).

Even if obese children had 25-OH Vit D concentrations in mean below the 25-OH Vit D concentrations of healthy non-obese children and even if four obese children had 25-OH Vit D levels below the normal range, the clinical relevance of these findings is at least questionable. We found no alterations in calcium, inorganic phosphate, AP, and most importantly in 1,25-OH Vit D levels, the bioactive vitamin D metabolite.

PTH concentrations were elevated in obese children in our study in concordance with most studies in adults (1, 3–5, 7, 43). Since patients with primary or secondary hyperparathyroidism were heavier than those in the control group, it has been postulated that increased PTH levels contribute towards obesity (6, 47, 48). Conversely, PTH decreased after weight loss in our study in concordance with most studies in obese adults (19, 20, 49), suggesting that the elevated serum PTH is a result and not a cause of obesity. Furthermore, we observed no alteration in calcium, inorganic phosphate, AP, and most importantly in 1,25-OH Vit D levels, the bioactive vitamin D metabolite.

The conflicting findings of increasing 25-OH Vit D and decreasing PTH in comparison with studies in obese adults with weight loss could be explained by the fact that weight loss was achieved in these studies by gastric surgery (17, 18). It is well known that, for example, Roux-Y bypass is associated with alterations in the vitamin D endocrine system such as secondary hyperparathyroidism due to disturbed resorption of vitamin D (16, 18).

One further finding of the present study is that 25-OH Vit D and PTH serum levels were not a major determinant of insulin sensitivity in obese children in cross-sectional and, most importantly, in longitudinal analysis. This finding is in concordance with most studies in obese adults (21, 32, 50). The association between low circulating concentrations of vitamin D, high PTH levels, and the prevalence of diabetes and impaired glucose tolerance in adults was mainly derived from cross-sectional studies (23, 24, 26, 51), which are susceptible to confounder effects. Apart from a lack of power due to the moderate study sample size, the normal 1,25-OH Vit D concentrations in our obese children could explain the missing relation between Vit D and insulin sensitivity, since 1,25-OH Vit D is essential for normal insulin secretion (51). Moreover, the HOMA model is only an assessment of insulin resistance and β-cell function. Clamp studies are the gold standard to analyze insulin resistance and sensitivity (52). Since the HOMA model correlates to clamp studies, it is a suitable method to study insulin resistance in field studies (53, 54). However, it has been reported that in vitamin D-depleted subjects especially the late insulin secretion is altered (23), which cannot be calculated by the HOMA model. Therefore, a relationship between insulin sensitivity and vitamin D status cannot be excluded by our study.

Data as median and interquartile range.

<table>
<thead>
<tr>
<th>Reduction of overweight (n=21)</th>
<th>No reduction of overweight (n=12)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>At baseline 1 year later</td>
</tr>
<tr>
<td></td>
<td>At baseline 1 year later</td>
</tr>
<tr>
<td>Energy intake (kcal/day)</td>
<td>1448±350 1186±267†</td>
</tr>
<tr>
<td>Fat (E%)</td>
<td>35.3±4.9 29.3±8.4†</td>
</tr>
<tr>
<td>Carbohydrate (E%)</td>
<td>49.3±4.9 52.9±7.8*</td>
</tr>
<tr>
<td>Protein (E%)</td>
<td>15.4±1.7 17.7±2.2†</td>
</tr>
<tr>
<td>Vitamin D intake (µg/day)</td>
<td>0.976±0.621 0.809±0.424</td>
</tr>
<tr>
<td>Calcium intake (mg/day)</td>
<td>653±232 656±205</td>
</tr>
</tbody>
</table>

*P<0.05 baseline versus 1 year later; †P<0.01 baseline versus 1 year later.
cautiously and underreporting might also explain the lower vitamin D intake after intervention. Furthermore, dietary recording after the end of intervention was performed in a subgroup of our study sample representing a potentially important bias. Finally, we have only demonstrated a statistical association between serum vitamin D system and weight status, which does not necessarily imply a cause-and-effect relationship. Accordingly, these alterations may simply be a pathophysiologically unrelated marker of obesity or related to other factors influencing both weight status and the vitamin D endocrine system. For instance, a deranged renal handling of calcium has been reported in obesity leading to a negative calcium balance and thus elevated serum PTH levels (20). This increased excretion of calcium was in parallel with sodium excretion (58), and the elevation of the latter may be a result of a higher salt intake in obese subjects (59).

In summary, fasting PTH levels were increased and 25-OH Vit D concentrations were decreased in obese children. These alterations normalized after reduction of overweight pointing towards reversibility in weight loss. PTH, 1,25-OH Vit D, and 25-OH Vit D were not related to insulin sensitivity in both cross-sectional and longitudinal analyses. Further prospective research is required to determine the physiological role of vitamin D endocrine system in obese humans.

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