Serum adiponectin levels, insulin resistance, and lipid profile in children born small for gestational age are affected by the severity of growth retardation at birth

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Abstract

Objective: Insulin resistance has been linked to intrauterine growth retardation (IUGR); adiponectin is a protein with insulin-sensitizing properties. This study was designed to test whether being born small for gestational age (SGA) has an effect on blood levels of adiponectin and leptin, insulin resistance parameters, and lipid profile in pre-puberty, taking into consideration the severity of IUGR.

Methods: Serum levels of adiponectin, leptin, total cholesterol (t-CHOL), high density lipoprotein (HDL)-cholesterol, low density lipoprotein (LDL)-cholesterol, triglycerides, apolipoproteins A-1 (Apo A-1), Apo B and Apo E, lipoprotein(a) (Lp(a)), fasting glucose, and insulin (Ins), the homeostasis model assessment insulin resistance index (HOMA-IR) and anthropometric indices were evaluated in 70 children aged 6–8 years, born appropriate for gestational age (AGA; n = 35) and SGA (n = 35), matched for age, gender, height, and BMI. SGA children were divided into two subgroups according to the severity of IUGR: SGA < 3rd percentile (n = 20), and SGA 3rd–10th percentile (n = 15). They were also subdivided in two subgroups, those with (n = 25) and those without (n = 10) catch-up growth, considering their actual height corrected for mid-parental height.

Results: SGA children had higher Ins and HOMA-IR than AGA children (Ins, 42 ± 23 vs 32 ± 11 pmol/l; HOMA-IR, 1.30 ± 0.8 vs 0.92 ± 0.3; P < 0.05). No significant difference in serum leptin was found between the SGA and the AGA groups but adiponectin showed a trend to be higher in SGA children (13.6 ± 5.7 vs 10.8 ± 5.9 mg/ml respectively). SGA children without catch-up growth had higher adiponectin (15.6 ± 8.5 mg/ml, P < 0.05) than AGA children. Among the SGA children, the subgroup < 3rd percentile had higher Lp(a) than the subgroup 3rd–10th percentile (P < 0.05). An independent positive correlation between adiponectin and Lp(a) was observed in SGA children (R = 0.59, P < 0.01).

Conclusion: SGA children, although more insulin resistant, had similar or higher adiponectin levels than matched AGA children in pre-puberty. The severity of IUGR appears to affect their metabolic profile during childhood.

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Introduction

Adiponectin is a protein consisting of 247 amino acids, produced by the most abundant gene transcript-1 (apM1) expressed in human adipose tissue (1, 2). This protein has been implicated in the pathophysiology of obesity-related insulin resistance, glucose intolerance, insulin-mediated lipoprotein metabolism, atherosclerosis, and coronary heart disease (1). The physiological role of adiponectin is yet to be completely determined, but its reduced expression and hypoadiponectinemia have been associated with insulin resistance in animal and human studies (1).

Low birth weight has also been related with increased risk for developing the metabolic syndrome in adulthood (3, 4). Some researchers have proposed that this association is genetically mediated (5), while others have postulated that it isthe result of fetal adaptation to an adverse intrauterine environment (3); the environment of the embryos may program alterations in their metabolic profile that will be expressed later in life (3, 6).

Only limited and controversial information is available about circulating adiponectin levels in pre-pubertal children born small for gestational age (SGA) (7–9). The two studies that investigated adiponectin levels in pre-puberty (7, 8) enrolled short SGA-born children who
were being monitored in outpatient endocrinology departments for short stature. It is known that most SGA born children ‘catch-up’ with the normal growth range postnatally with only a minority (10–20%) remaining short in childhood and adult life (10). The present study was designed to enroll a representative sample of SGA children born in a regional hospital and not only those with short stature.

Associations between adiponectin and blood lipids have been demonstrated in adults and adolescents (11, 12), but limited data have been published on blood lipids and their relation to adiponectin in SGA born children. A few studies showed mild disturbances of lipids in this group (13, 14), while others failed to reveal any lipid abnormality (8, 15). Lipoprotein(a) (Lp(a)) is an independent predictor of coronary disease that has been measured only in one study in SGA children (16). Although levels of Lp(a) are to a high degree genetically determined, significant associations with other lipoproteins have been demonstrated in children (16, 17) and its relation to other metabolic parameters in SGA children would be of interest.

SGA children are usually investigated as a group without taking into consideration the severity of their intrauterine growth retardation (IUGR). Pooling the findings may hide differences of clinical significance between more and less severely growth retarded children. This study was designed to investigate adiponectin, insulin sensitivity, leptin, the lipid profile, and anthropometric markers in children born appropriate for gestational age (AGA) and SGA children, taking into consideration also the severity of IUGR.

Subjects and methods

The study included 70 pre-pubertal children aged 6–8 years, born SGA (n=35) or AGA (n=35) during an 18-month period (January 1996–June 1997) at the University Hospital of Ioannina, Greece, and living in the same geographical area in Northwestern Greece. This hospital hosts the majority (85%) of births in a region. The sensitivity of the adiponectin assay was 0.40 μg/ml. The intra- and interassay coefficients of variation (CVs) were <10 and <15% respectively. Serum leptin levels were measured by the Elisa method using the kit of Phoenix Pharmaceuticals Inc, Belmont, CA, USA (EK-ADI-01). The sensitivity of the adiponectin assay was 0.40 μg/ml. The intra- and interassay coefficients of variation (CVs) were <10 and <15% respectively. Serum leptin levels were measured by the Elisa method using the kit of BioVendor Laboratory Medicine Inc, Modrice, Czech Republic. The sensitivity of the leptin assay was 0.50 ng/ml. The intra- and interassay CV values were 5 and 10% respectively. Serum total cholesterol (t-CHOL), triglycerides (TRG), HDL-cholesterol (HDL-C), Lp(a) and apolipoproteins A-1 (Apo A-1), B (Apo B), and E (Apo E) were determined using standardized techniques described previously (20). Serum LDL-cholesterol (LDL-C) was calculated using the Friedewald–Freedrickson formula: LDL-C = t-CHOL − (HDL-C + TRG/2.2) (provided that triglycerides levels were lower than 400 mg/dl) (21). Fasting plasma insulin level (Ins) was determined using an immunoenzymatic method (analyzer AXXSYM, Abbott) and fasting glucose (Glc) concentration by the glucose oxidase method. Fasting
Adiponectin in children born growth retarded

Ins, the fasting Glc/Ins ratio and the homeostasis assessment model for insulin resistance (HOMA-IR = fasting Ins (mU/l) × fasting Glc (mmol/l)/22.5) were chosen as measures of insulin sensitivity (22).

**Statistical analysis**

A sample size of 70 children was calculated to be adequate for detecting a difference of 25% in serum adiponectin between the AGA and SGA groups with a power > 80% at a significance level of 5%. The calculation of the sample size was based on the adiponectin values (mean and s.d.) of the SGA study group, and on already published adiponectin values in children (23). The data were analyzed by one-way ANOVA, using Fisher’s protected least significant difference test for comparing the means of the study groups pairwise. The Mann–Whitney U-test was used for comparison between abnormally distributed data. Results are reported as the mean ± S.D. and ranges (minimum and maximum values). Multiple regression and forward stepwise regression analyses were also used to correlate adiponectin values and the other tested variables. After ascertaining that some variables were not normally distributed, logarithmic transformation was made. Statistical analyses were performed using the Stat View software package of SAS Institute, Inc., Cary, NC. Differences were considered statistically significant at P-value < 0.05.

**Results**

On retrospective analysis of the hospital neonatal case notes, 68 children were identified as born SGA during the 18-month period. Forty-five of the sixty-eight were selected to participate in the study based on birth date, with every third SGA child being excluded chronologically. A further ten children were eliminated; for two the parents did not give consent, for three their contact details were not found and five were excluded because of BMI > 85th percentile. The remaining 35 children were enrolled in the study. The perinatal characteristics of the non-participants (birth weight, gestational age, sex, mode of delivery) and of their mothers (age, parity, weight gain during pregnancy, pathology of gestation, socioeconomic status) did not differ from those of the children who participated to the study. The 35 matched controls were identified from among 150 AGA children born in the hospital during the same period who were initially contacted. Risk factors for IUGR were detected in 19 of the 35 SGA born children, with hypertensive disease of pregnancy being the most common (11/19 cases) following by placental insufficiency (4/19) and low maternal weight gain during pregnancy (4/19).

The anthropometric indices of the subjects at birth and at the time of the study are depicted in Table 1. At the time of the study there were no significant differences in mean age, BW, BMI, and BMI SDS between the AGA and SGA groups, and all the children were at Tanner stage I. No statistical differences were found between the groups in SFT, but the subgroup SGA < 3rd percentile had significantly lower WC than the control AGA group (P < 0.05).

Table 2 shows the serum levels of adiponectin and leptin, and the insulin resistance parameters in the two groups of children at the pre-pubertal stage. No significant differences in levels of leptin were found between the SGA and the AGA groups, but adiponectin

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**Table 1** Anthropomorphic variables at birth and at the time of study in appropriate for gestational age (AGA) and small for gestational age (SGA) groups of pre-pubertal children.

<table>
<thead>
<tr>
<th>Variable</th>
<th>AGA (n=35)</th>
<th>SGA (n=35)</th>
<th>SGA 3rd–10th percentile (n=15)</th>
<th>SGA &lt;3rd percentile (n=20)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>At birth</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gestational age (weeks)</td>
<td>37.9 ± 1.4</td>
<td>37.0 ± 1.3</td>
<td>36.8 ± 1.4</td>
<td>37.1 ± 1.7*</td>
<td>NS</td>
</tr>
<tr>
<td>Gender (male)</td>
<td>20</td>
<td>20</td>
<td>9</td>
<td>11</td>
<td>NS</td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td>3385 ± 380</td>
<td>1885 ± 497*</td>
<td>1990 ± 448*</td>
<td>1755 ± 426*</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>Birth length (cm)</td>
<td>51.0 ± 2</td>
<td>44.5 ± 4.7*</td>
<td>45.4 ± 4.3*</td>
<td>42.7 ± 5.1*</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>Head circumference (cm)</td>
<td>34.8 ± 1.3</td>
<td>31 ± 2.8*</td>
<td>31 ± 2.7*</td>
<td>31 ± 2.9*</td>
<td>&lt;0.001*</td>
</tr>
<tr>
<td>At the time of study</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>6.89 ± 1.4</td>
<td>6.92 ± 1.2</td>
<td>6.94 ± 1.7</td>
<td>6.89 ± 2.2</td>
<td>NS</td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>26.1 ± 6.2</td>
<td>25.6 ± 6.1</td>
<td>25.8 ± 6.6</td>
<td>24.6 ± 6.7</td>
<td>NS</td>
</tr>
<tr>
<td>Body height (cm)</td>
<td>119 ± 8.9</td>
<td>118 ± 9.3</td>
<td>119 ± 8.9</td>
<td>117 ± 8.1</td>
<td>NS</td>
</tr>
<tr>
<td>Head circumference (cm)</td>
<td>50.9 ± 1.2</td>
<td>50.4 ± 2.1</td>
<td>50.4 ± 1.9</td>
<td>50.6 ± 2.2</td>
<td>NS</td>
</tr>
<tr>
<td>Waist circumference (cm)</td>
<td>60.9 ± 7.7</td>
<td>59.1 ± 9.6</td>
<td>59.7 ± 10.2</td>
<td>58.4 ± 8.8*</td>
<td>&lt;0.05*</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>18.5 ± 2.1</td>
<td>17.7 ± 2.6</td>
<td>18.2 ± 2.5</td>
<td>17.5 ± 2.9</td>
<td>NS</td>
</tr>
<tr>
<td>BMI-SDS</td>
<td>0.18 ± 0.48</td>
<td>0.17 ± 0.52</td>
<td>0.18 ± 0.52</td>
<td>0.13 ± 0.60</td>
<td>NS</td>
</tr>
<tr>
<td>SFT-triceps (mm)</td>
<td>10.1 ± 2.7</td>
<td>9.0 ± 3.4</td>
<td>9.3 ± 3.5</td>
<td>8.9 ± 3.7</td>
<td>NS</td>
</tr>
<tr>
<td>SFT-subcapular (mm)</td>
<td>8.8 ± 2.6</td>
<td>8.6 ± 2.9</td>
<td>9.1 ± 2.8</td>
<td>8.4 ± 3.1</td>
<td>NS</td>
</tr>
<tr>
<td>SFT-suprailiac (mm)</td>
<td>9.8 ± 2.7</td>
<td>9.4 ± 3.4</td>
<td>9.7 ± 3.9</td>
<td>8.8 ± 3.0</td>
<td>NS</td>
</tr>
</tbody>
</table>

Values are means ± s.d. Significances from pairwise comparisons for all groups are shown in the right column and they are as follows: *P < 0.001 all SGA groups vs AGA for characteristics at birth, †P < 0.05 SGA < 3rd vs AGA for WC. SFT, skinfold thickness.
showed a trend to be higher in the SGA than in the AGA children (P = 0.08). In the subgroup of SGA < 3rd percentile levels of adiponectin remained higher than AGA group after adjustment for differences in WC (P < 0.05). Concerning the insulin resistance parameters, the SGA group had significantly higher values of fasting Ins (P < 0.05) and HOMA-IR (P < 0.05) than the AGA group. The subgroup SGA < 3rd percentile had lower fasting Glc values than the AGA group (P < 0.05).

As shown in Table 3, no significant differences were observed in the serum t-CHOL, LDL-C, TRG, Apo A-1, Apo B, Apo E, Lp(a), and Apo B/Apo A-1 ratio between the SGA group and the AGA controls. However, within the SGA group, Lp(a) levels were significantly higher in the subgroup SGA < 3rd percentile than in the subgroup SGA 3rd–10th percentile (P < 0.05).

Further analysis was made according to catch-up growth between SGA subgroups A (n = 10) and B (n = 25). The two groups did not differ for age or sex. No differences were found in insulin resistance indices between these two subgroups. The mean values in subgroups A and B respectively were for: Ins (pmol/l) 43.6 ± 21 and 39 ± 16; Glc/Ins, 16 ± 6.5 and 19.6 ± 11 HOMA-IR 1.33 ± 0.77 and 1.15 ± 0.67. Adiponectin levels (μg/ml) in subgroups A and B respectively were 15.6 ± 8.5 and 12.4 ± 6.6, and leptin levels (ng/ml) were 7.2 ± 5.1 and 12.9 ± 12, and the differences were not significant. However, when compared with the AGA group, adiponectin values were significantly higher in subgroup A (SGA without catch-up growth; P < 0.05) but not in subgroup B (SGA with catch-up growth). No significant differences were found in the levels of blood lipids between the two subgroups of SGA children.

**Multiple regression analysis**

Adiponectin was regarded as dependent factor and anthropometrics at birth and at the time of the study, insulin resistance indices, leptin, and levels of lipids were analyzed as independent variables. All the factors were first examined by simple regression and those which showed a significant relation to adiponectin were subsequently entered to a multiple regression model. These factors were BW, BH, BMI, and Lp(a) levels in the SGA group and BMI and Glc/Ins ratio in the AGA group. A positive independent association was observed only between adiponectin and Lp(a) levels in SGA children (t = 2.92, R = 0.59, P < 0.01). No independent relation was found in the AGA children between adiponectin and any of the other factors. Similar results in multiple and stepwise regressions were obtained when the factors were examined irrespective of their relation to adiponectin in the simple regression model. To avoid the ‘colinearity’ effect, factors that showed a close relation to each other (i.e., BMI and BW. Ins and Glc/Ins etc.) were entered separately into the model.

**Discussion**

In this study, higher insulin levels and HOMA-IR values were observed in the SGA group than in the matched AGA group. Several studies in adults and adolescents have shown an inverse relationship between adiponectin levels and insulin resistance (24–26). According to these earlier studies (24–26), the higher insulin and HOMA-IR levels observed in the children born SGA would be expected to be accompanied by reduced adiponectin levels compared with the controls. However, this was not so, and the adiponectin levels of the children born SGA were similar to or even higher than those of the AGA group.

Information on adiponectin levels in pre-pubertal children born SGA is limited (7, 8). Lopez-Bermejo et al. reported no differences in adiponectin levels between children born AGA and SGA attending an Outpatient Endocrinology Clinic for short stature (8). However, in agreement with the results of the present study, the adiponectin levels were shown to be higher in the SGA children after adjusting for sex, age, and BMI and also after accounting for insulin resistance in a general linear model of comparison (8).

In contrast to the results of the present study, Cianfarani et al. found lower adiponectin levels in SGA children born AGA and SGA attending an Outpatient Endocrinology Clinic for short stature (8).
Table 3 Plasma levels of lipids and apolipoproteins at the time of examination of pre-pubertal children born appropriate for gestational age (AGA) and small for gestational age (SGA).

<table>
<thead>
<tr>
<th>Study-group</th>
<th>t-CHOL (mmol/l) mean (ranges)</th>
<th>LDL-C (mmol/l) mean (ranges)</th>
<th>Apo A1 (g/l) mean (ranges)</th>
<th>Apo B (g/l) mean (ranges)</th>
<th>Apo E (g/l) mean (ranges)</th>
<th>Lp (a) (mg/dl) mean (ranges)</th>
</tr>
</thead>
<tbody>
<tr>
<td>AGA (n=35)</td>
<td>4.4±0.08 (2.69–6.28)</td>
<td>2.78±0.57 (1.76–3.37)</td>
<td>1.76±0.59 (1.47–2.30)</td>
<td>0.73±0.14 (0.60–1.19)</td>
<td>0.59±0.13 (0.40–0.99)</td>
<td>1.57±0.19 (1.15–2.35)</td>
</tr>
<tr>
<td>SGA (n=35)</td>
<td>4.31±0.61 (2.69–6.28)</td>
<td>2.68±0.46 (1.76–3.37)</td>
<td>1.51±0.11 (1.15–2.35)</td>
<td>0.74±0.13 (0.60–1.19)</td>
<td>0.59±0.13 (0.40–0.99)</td>
<td>1.51±0.11 (1.15–2.35)</td>
</tr>
<tr>
<td>SGA 3rd–10th percentile</td>
<td>4.3±0.61 (2.69–6.28)</td>
<td>2.68±0.59 (1.76–3.37)</td>
<td>1.51±0.11 (1.15–2.35)</td>
<td>0.74±0.13 (0.60–1.19)</td>
<td>0.59±0.13 (0.40–0.99)</td>
<td>1.51±0.11 (1.15–2.35)</td>
</tr>
<tr>
<td>SGA &lt;3rd percentile</td>
<td>4.3±0.61 (2.69–6.28)</td>
<td>2.68±0.59 (1.76–3.37)</td>
<td>1.51±0.11 (1.15–2.35)</td>
<td>0.74±0.13 (0.60–1.19)</td>
<td>0.59±0.13 (0.40–0.99)</td>
<td>1.51±0.11 (1.15–2.35)</td>
</tr>
</tbody>
</table>

Significance from pairwise comparisons for all groups are as follows: * P < 0.05 SGA < 3rd percentile vs SGA 3rd–10th percentile for Lp(a). Median values for Lp(a): (AGA) 4.13 mg/dl, (SGA) 2.8 mg/dl, (SGA <3rd percentile) 0.52 mg/dl, (SGA 3rd–10th percentile) 0.31 mg/dl.

A recent large study in young adults (27) has shown that adiponectin levels are marginally higher in subjects born AGA compared with those born SGA (13.2 ± 6 vs 12.6 ± 7 μg/ml), with a U-shape relationship between adiponectin and insulin resistance indices in the SGA, but not in the AGA group. This observation implies that some young adults born SGA are insulin-resistant despite relatively high levels of adiponectin, a finding observed also in some of the children in the present study. The authors interpret this as a defect in the insulin sensitizing action of adiponectin in SGA born adults.

It could be speculated that there is a period during the development during which SGA children exhibit insulin resistance combined with relatively raised adiponectin levels. Increase in the adiponectin level may be seen as an adaptation mechanism at this time, before a decline of this protein takes place later in life. It is possible that other metabolic, genetic, or inflammatory factors may also be related with the relatively high adiponectin levels observed in the SGA children in this study.

The links between IUGR and later metabolic disturbances, such as increased insulin resistance observed in this study, have not been fully elucidated. It has been speculated that intrauterine malnutrition could lead to decreased β-cell mass, and insulin resistance is intended to divert inadequate glucose resources to essential organs (brain and heart) at the expense of muscle (3). Another hypothesis involves a primarily peripheral reduced insulin sensitivity in utero, which allows a redistribution of nutrients, leading to eventual β-cell exhaustion (28). According to the ‘catch-up growth’ hypothesis (29), the crucial time for the development of long-term consequences is early postnatal life. Recent studies have shown that catch-up growth, especially in children born with a small birth weight, may cause long-term metabolic disturbances through mechanisms as yet unknown (29–31). In order to investigate whether catch-up growth affects the metabolic profile in pre-pubertal SGA children, the SGA children in the present study were subdivided into two subgroups according to their corrected height. The children attending an outpatient clinic for short stature than in short or obese AGA children (7). There are certain differences between the two studies. In the present study, a representative sample of SGA children recruited from birth charts was examined, and not only short SGA children, who account for only 10–20% of the total SGA born children (10). The controls in the Cianfarani study were either short or obese children and were significantly older than the respective SGA children. They also had an age range much broader than the children in the present study. These differences may have led to different findings in adiponectin values between the two studies, and when the SGA children in the present study were classified according to catch-up growth, increased adiponectin was observed in SGA children without catch-up growth, in agreement with the findings of Cianfarani et al. in short SGA children.
without catch-up growth had higher adiponectin levels than AGA children, in accordance with the findings of previous studies (7, 8). Cianfarani et al. (7) estimated catch-up growth by calculating the corrected height, while Lopez-Bermejo et al. (8) by considering the BMI SDS, and both reported higher adiponectin levels in SGA groups not experiencing catch-up growth.

Reported data have shown that SGA children tend to develop lower lean body mass than those born AGA, which may contribute to differential metabolic functioning (32, 33). Although the two study groups of SGA and AGA children were matched for height, BW, and BMI and had similar SFT and WC, subtle differences in body fat content or fat distribution cannot be ruled out. The measurement of BMI, WC, and skinfolds are regarded as a reliable estimator of body composition although a dual-energy X-ray absorptiometry analysis is reported to be more precise (34). Minor differences in fat content or distribution between the two groups may have an impact on insulin resistance indices and adipokine levels. Irrespective of the factors that contributed to the higher insulin resistance indices observed in the pre-pubertal children born SGA, it appears that the somewhat higher circulating levels of adiponectin in this group cannot overcome the increased insulin resistance. The lower glucose levels observed in SGA < 3rd percentile subgroup may be the result of subgroup analysis and small sample size and not an indication of improved insulin sensitivity in this subgroup because all the insulin resistance parameters were similarly affected in both SGA subgroups.

The relationship between low birth weight and blood lipids has scarcely been investigated in childhood (13–16). Thenola et al. (14) reported that SGA children at the age of 12 years had the levels of total cholesterol in the high normal range, although no significant differences were found compared with AGA children. Cianfarani et al. observed that SGA children without catch-up growth had significantly higher cholesterol levels than AGA children, but in a study comprising only short children (13). Decci et al. (15) and Pulzer et al. (16) found no difference in lipid profile in 10- and 15-year-old children respectively, except in Lp(a), which was measured in the latter study and found to be higher in SGA than AGA adolescents. In the present study, no differences were found in lipid levels between the SGA children, either examined as a group or subdivided according to catch-up growth, and matched AGA controls. The previous studies are not directly comparable with this study, as the children differed in age and pubertal development from those of the present study.

It is known that adiponectin may modulate the plasma lipid profile in an antiatherogenic manner, as associations with high HDL cholesterol and low triglyceride concentrations have been shown in adults (1). The present study demonstrated a positive correlation between adiponectin and Lp(a) only in children born SGA (but not in the AGA) and no relations with the other lipids. It was also observed that the most severely growth-restricted SGA children (birth weight < 3rd percentile) had higher Lp(a) levels than those between 3rd and 10th. It is difficult to determine the clinical significance of these findings.

Although intrauterine growth is a continuous process and the SGA classification is somewhat artificial, it appears that children at the low end of the birth weight range are prone to various metabolic aberrations. The severity of IUGR may be a factor to be taken into consideration when adiponectin, glucose and lipid metabolism are investigated in SGA children. This study indicates that prepubertal SGA born children have levels of serum adiponectin similar to or even higher than AGA children matched for age, sex, and BMI, despite the fact that the former are more insulin resistant. Further studies are required to assess whether the differences between AGA and SGA children observed in the pre-pubertal phase represent a long-term phenomenon.

References
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