Circulating and cerebrospinal fluid ghrelin and leptin: potential role in altered body weight in Huntington’s disease

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Abstract

Objective: In addition to neurological impairment, weight loss is a prominent characteristic of Huntington’s disease (HD). Neuropathologically, the disease affects the caudate nucleus and the cerebral cortex, and also the hypothalamus. The recently discovered orexigenic hormone of gastric origin, ghrelin and the adipocyte hormone leptin, are two peripherally produced hormones exerting opposite effects on specific populations of hypothalamic neurons that play a key role in regulating energy intake and energy output. The aim of this study was to investigate the possible involvement of cerebrospinal fluid (CSF) and circulating ghrelin and leptin in the regulation of energy balance in patients with HD.

Methods: Twenty healthy normal-weight subjects undergoing orthopedic surgery, and fifteen patients with genetically verified HD, were enrolled in this study. The unified Huntington’s disease rating scale (UHDRS) was used to assess clinical course of the disease. Blood samples for hormonal measurements were obtained by venipuncture and in-parallel CSF samples for leptin/ghrelin determination were obtained by lumbar puncture.

Results: Patients with HD had increased concentrations of ghrelin in plasma compared with healthy subjects (4523.7 ± 563.9 vs 2781.1 ± 306.2 pg/ml, P < 0.01). On the other hand, patients with HD had decreased concentrations of leptin in plasma compared with healthy subjects (4.8 ± 1.6 vs 10.9 ± 2.4 ng/ml, P < 0.01). The concentrations of CSF ghrelin and CSF leptin were equivalent to values in healthy subjects. No correlation was found between disease duration and other clinical features of HD and plasma or CSF leptin/ghrelin levels. In patients with HD, baseline levels of GH, IGF-I, insulin and glucose did not differ from those in healthy subjects.

Conclusion: High circulating ghrelin and low leptin levels in patients with HD suggest a state of negative energy balance. Early nutritional support of patients with HD is advocated since patients with HD and higher body mass index at presentation have slower progression of the disease.

Introduction

A robust physiological system acts to maintain relative constancy of weight. The available data suggest that the concentration of afferent signals is sensed by groups of neurons in the hypothalamus and other brain regions and the neural circuits regulating weight are likely to be complex (1).

Huntington’s disease (HD) is a genetic, autosomal dominant, degenerative brain disorder, which neuropathologically strikes the caudate nucleus, cerebral cortex and hypothalamus. Weight loss is a prominent characteristic of HD and the basis of wasting is unclear. Selective neuronal loss in the hypothalamic lateral tuberal nucleus in HD patients has been shown (2). Recently, the protein product of the HD gene, huntingtin, was shown to be increased in the arcuate nucleus of postpartum female animals, yet its function is currently unknown (3). Weight loss in HD occurs in spite of an adequate caloric intake (4). Higher body mass index (BMI) at presentation is associated with slower disease progression. These features have been observed in transgenic mice (5).

Both acute and chronic alterations in energy balance or nutritional status may be relayed to the hypothalamic arcuate nucleus by neuronal pathways or via alterations in the levels of circulating hormones. Ghrelin, the recently discovered orexigenic hormone of gastric origin, may provide an endocrine link between the stomach and the central circuits involved with the...
regulation of energy intake and growth hormone (GH) release (6–10). Plasma levels of ghrelin are lower in obese subjects compared with lean subjects (11). In contrast, states such as fasting and anorexia nervosa are characterized by increased plasma ghrelin levels (12). Leptin, the adipocyte hormone, is believed to act tonically as an afferent signal from adipose tissue to the brain, in particular the hypothalamus, as part of a negative feedback loop regulating the size of energy stores and energy balance (13, 14). Body fat significantly contributes to leptin levels and thus plasma leptin levels are elevated in obesity and conversely decreased in anorexia nervosa (15, 16).

Recently ghrelin has been measured in human cerebrospinal fluid (CSF) and a negative association between fasting CSF ghrelin levels and BMI was found (17). Leptin is transported across the blood–brain barrier (BBB) and has been measured in CSF in healthy subjects, and in patients with anorexia nervosa or obesity (18, 19). A significant positive correlation between leptin concentrations in the CSF and plasma in healthy normal-weight subjects was found. CSF leptin levels in anorexia nervosa correlated with nutritional status. There is indirect evidence for impaired transport of leptin across the BBB in obesity (18).

In order to investigate the possible involvement of ghrelin and leptin in the regulation of metabolic balance, in this study we measured plasma and CSF ghrelin and leptin in a group of healthy subjects at normal weight and in patients with HD.

Materials and methods

Study subjects

After providing informed consent, 20 healthy normal-weight subjects and 15 patients with HD were studied. The protocol was approved by the University Hospital Ethics Committee. Fifteen HD patients (nine female and six male; age 48.9±3.2 years) admitted at the Institute of Neurology participated in the study. They had been previously genetically diagnosed, were stable, medication free and had no other concurrent medical illness; they consumed an average hospital diet. The patient’s signs and symptoms were rated using the unified Huntington’s disease rating scale (UHDRS), disability scale (DS) and functional disability (FDS) as part of their ongoing clinical care and GH sensitivity of 0.011 μg/l, and coefficients of variation of 6.3% (0.4 μg/l), 5.3% (10.2 μg/l) and 4.2% (43.4 μg/l).

Insulin levels were measured using RIA kits (INEP, Zemun, Yugoslavia) with the limit of detection being 0.60 mU/l. Within-assay coefficient of variation was between 2.5 and 5.2%. Interassay variations were 7.7–10.7%.

Insulin-like growth factor-I (IGF-I) levels were measured using enzyme-labeled chemiluminescent immunometric assay (Immulite 2000 IGF-I, DPC, LA, USA) with the limit of detection being 20 ng/ml. Within-assay coefficient of variation was between 2.3 and 3.9%. Interassay variations were 3.7–8.1%.

Blood glucose levels were measured using Beckman Glucose Analyzer 2 (Fullerton, CA, USA).

Statistical analysis

Group values are reported as the mean±S.E. Leptic values were non-Gaussian distributed and thus were lognormally transformed before analysis. Student’s paired t-test was used. Relationships between CSF leptin/ghrelin and plasma leptin/ghrelin and body weight were assessed by Pearson correlation. Correlations of disease duration, DS, FDS and UHDRS with plasma leptin/ghrelin and CSF leptin/ghrelin were analysed by Spearman correlation. All analyses were performed using the SPSS package. A P value of less than 0.05 was considered to be statistically significant.

Hormone measurements

Leptin levels were measured using RIA Linco kits (St Charles, MO, USA) with the limit of detection being 0.5 ng/ml. Within-assay coefficient of variation was between 4.6 and 6%. Interassay variations were 6.8–9.5%. All assays were run in duplicate.

Ghrelin (total) was subsequently measured using Linco RIA with the limit detection being 100 pg/ml. Within-assay coefficient of variation was between 5.6 and 8%. Interassay variations were 7.8–9.8%. All assays were also run in duplicate. For hormone assays CSF samples were concentrated fourfold in a vacuum lyophilizer, and ghrelin and leptin were subsequently measured.

GH levels were measured using time-resolved fluorometric assay (Wallac, Turku, Finland) with a GH sensitivity of 0.011 μg/l, and coefficients of variation of 6.3% (0.4 μg/l), 5.3% (10.2 μg/l) and 4.2% (43.4 μg/l).

Statistical analysis

Group values are reported as the mean±S.E. Leptin values were non-Gaussian distributed and thus were lognormally transformed before analysis. Student’s paired t-test was used. Relationships between CSF leptin/ghrelin and plasma leptin/ghrelin and body weight were assessed by Pearson correlation. Correlations of disease duration, DS, FDS and UHDRS with plasma leptin/ghrelin and CSF leptin/ghrelin were analysed by Spearman correlation. All analyses were performed using the SPSS package. A P value of less than 0.05 was considered to be statistically significant.
Results

The clinical characteristics of 20 healthy subjects and 15 patients with HD are shown in Table 1. Plasma ghrelin concentrations in patients with HD were significantly elevated compared with those of healthy controls (Table 1, Fig. 1) (4523.7 ± 563.9 vs 2781.1 ± 306.2 pg/ml, \( P < 0.01 \)). There was no difference in CSF ghrelin levels in patients with HD and healthy subjects (Fig. 1).

Plasma leptin levels were significantly decreased in patients with HD in comparison with healthy subjects (Table 1, Fig. 2). Plasma leptin levels were significantly lower in male patients with HD in comparison with healthy males (1.2 ± 0.4 vs 7.8 ± 3.1 ng/ml, \( P < 0.05 \)) and in female patients with HD in comparison with healthy females (3.8 ± 1.5 vs 14.3 ± 4.9 ng/ml, \( P < 0.05 \)).

Table 1 Characteristics of healthy subjects and patients with HD.

<table>
<thead>
<tr>
<th></th>
<th>Healthy (n = 20)</th>
<th>HD (n = 15)</th>
<th>P</th>
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</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>46.2 ± 4.1</td>
<td>48.9 ± 3.2</td>
<td>n.s.</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.8 ± 0.8</td>
<td>20.2 ± 0.6</td>
<td>0.001</td>
</tr>
<tr>
<td>Ghrelin plasma (pg/ml)</td>
<td>2781.1 ± 306.2</td>
<td>4523.7 ± 563.9</td>
<td>0.007</td>
</tr>
<tr>
<td>Ghrelin CSF (pg/ml)</td>
<td>414.8 ± 15.6</td>
<td>453.0 ± 40.8</td>
<td>n.s.</td>
</tr>
<tr>
<td>Leptin plasma (ng/ml)</td>
<td>10.9 ± 2.4</td>
<td>4.8 ± 1.6</td>
<td>0.007</td>
</tr>
<tr>
<td>Leptin CSF (pg/ml)</td>
<td>301.4 ± 40.2</td>
<td>297.5 ± 47.6</td>
<td>n.s.</td>
</tr>
<tr>
<td>Insulin (mU/l)</td>
<td>15.8 ± 3.1</td>
<td>15.3 ± 2.5</td>
<td>n.s.</td>
</tr>
<tr>
<td>IGF-1 (ng/ml)</td>
<td>116.6 ± 11.0</td>
<td>85.0 ± 11.3</td>
<td>n.s.</td>
</tr>
<tr>
<td>Growth hormone (μg/l)</td>
<td>1.05 ± 0.58</td>
<td>1.23 ± 0.54</td>
<td>n.s.</td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>4.2 ± 0.4</td>
<td>3.9 ± 0.3</td>
<td>n.s.</td>
</tr>
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</table>

n.s., \( P > 0.05 \).

Plasma insulin, IGF-I, growth hormone and glucose levels in patients with HD were similar to those in healthy subjects (Table 1).

Discussion

The present study shows for the first time that circulating ghrelin levels are increased and leptin levels are decreased in patients with HD, suggesting a state of negative energy balance. The rise in circulating ghrelin and fall in plasma leptin are compensatory and serve to preserve body weight and maintain energy homeostasis. In conditions of negative energy balance in addition to suppression of leptin levels, the increase in ghrelin serves to counter further decreases in energy stores. It seems that in patients with HD, the hypothalamic circuits through which ghrelin acts are not sensitive to perturbations in energy balance or that the effects of ghrelin are overcome by other central system signals involved in the regulation of energy homeostasis (13). Thus elevated ghrelin concentrations in our study may reflect ghrelin resistance. The neuronal targets for both ghrelin and leptin in the hypothalamus are in the arcuate nucleus, and ventromedial and lateral hypothalamus, and these structures express high

Table 2 Clinical features of HD patients.

<table>
<thead>
<tr>
<th></th>
<th>HD patients (n = 15)</th>
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<tbody>
<tr>
<td>Duration of the disease (years)</td>
<td>6.0 ± 0.95</td>
</tr>
<tr>
<td>Disability scale (DS)</td>
<td>78.7 ± 3.8</td>
</tr>
<tr>
<td>Functional disability scale (FDS)</td>
<td>9.1 ± 0.9</td>
</tr>
<tr>
<td>Unified Huntington’s disease rating scale (UHDRS)</td>
<td>43.8 ± 5.5</td>
</tr>
</tbody>
</table>

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levels of leptin and ghrelin receptors (20, 21). Quite clearly ghrelin and leptin appear to impose opposite effects on arcuate neuropeptide Y (NPY) neurons. Ghrelin triggers the expression of mRNA for Agouti related peptide (AGRP) and NPY while leptin induces reduction of NPY mRNA expression (22, 23). The involvement of orexigenic and anorexigenic pathways in weight loss in HD has not been studied so far. What has been suggested is that the selective neuronal loss in the hypothalamic lateral tuberal nucleus may play a role in the weight loss observed in the early stages of HD (2, 24). High levels of glutamate receptors normally present in the lateral tuberal nucleus might render these neurons selectively susceptible to excitotoxic cell death. Thus severe cell loss in the lateral tuberal nucleus may underlie the catabolic state that frequently occurs in HD patients (25). Furthermore, astrocytes in the arcuate nucleus of postpartum lactating female rats have increased levels of the huntingtin protein, which might play a role in the processes that regulate neuroendocrine function (3).

Experimental studies assessing the permeability of the BBB to ghrelin indicate that saturable systems transport ghrelin from brain to blood and vice versa (26). A negative association between fasting CSF ghrelin levels and BMI in humans has been reported, with lowered CSF levels found in obese patients (17). In keeping with this we provide the first information on CSF ghrelin in patients with HD. CSF ghrelin levels tended to be higher among HD patients but did not reach statistical significance possibly due to the normal BMI of the patients and the relatively small sample size. The same is true for CSF leptin levels, which tended to be lower. One may speculate that with further significant weight loss CSF concentrations may change; this has been shown for CSF leptin levels in patients with anorexia nervosa (19). Our data show similar ghrelin levels in the CSF in healthy subjects and in patients with HD, despite higher circulating levels; this rules out the possibility of ghrelin resistance due to altered transport across the BBB. On the other hand, in order to explain why CSF and serum levels of ghrelin and leptin are dissociated in our study, one also has to consider the possibility of altered transport properties of the BBB in HD, as the leptin transport system seems to be regulated (27).

The transgenic model of HD, the R6/2 mouse, shows severe wasting beginning at 12 weeks of age and dies between 12 and 15 weeks. Enhanced accumulation of body fat accompanied by increased serum leptin in transgenic mice between 8 and 9 weeks was observed and this initial obesity may stem from a defect in fat breakdown by adipocytes (5). Even if adipocyte defects occur in HD victims and contribute to their defective weight regulation, the possibility remains that HD mutation affects other regulatory systems, including those in the hypothalamus involved in feeding and weight regulation.

We did not find any correlation between disease duration—and patient’s signs and symptoms rated using standard scales—and parameters of negative energy homeostasis (leptin/ghrelin). These findings are consistent with those presented by Djourousse et al. (24) who showed that even in the earliest stages of HD, patients weigh significantly less than healthy age-matched controls. The observed alterations in ghrelin and leptin levels in HD patients may reflect the nutritional status. Other metabolic adaptations (insulin, IGF-I, GH and glucose levels) to chronic energy deficits have not occurred.

In conclusion we have shown that patients with HD exhibit the hyperghrelinemia and hypoleptinemia that are markers of negative energy balance. Since patients with HD and higher BMI at presentation have slower progression of the disease, the possibility of early nutritional support should be considered.

Acknowledgements

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References


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