A promoter polymorphism of the CYP27B1 gene is associated with Addison's disease, Hashimoto's thyroiditis, Graves' disease and type 1 diabetes mellitus in Germans

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Abstract

Background: CYP27B1 hydroxylase catalyzes the conversion of 25 hydroxyvitamin D3 (25OHD3) to 1,25(OH)2D3, the most active natural vitamin D metabolite, which plays a role in the regulation of immunity and cell proliferation. We therefore investigated two single nucleotide polymorphisms in the CYP27B1 hydroxylase gene for an association with Addison's disease, Hashimoto's thyroiditis, Graves' disease and type 1 diabetes mellitus.

Methods: Patients with Addison's disease (n=124), Hashimoto's thyroiditis (n=139), Graves' disease (n=334), type 1 diabetes mellitus (n=252) and healthy controls (n=320) were genotyped for the promoter (–1260) C/A polymorphism and for the intron 6 (+2838) C/T polymorphism of the CYP27B1 gene. Patients and controls were compared using genotype-wise and allele-wise X2 testing.

Results: A significant association was found between allelic variation of the promoter (−1260) C/A polymorphism and Addison's disease, Hashimoto's thyroiditis, Graves' disease and type 1 diabetes mellitus (P=0.0062, P=0.0173, P=0.0094 and P=0.0028 respectively). Significant differences were also observed for the intron 6 (+2838) C/T polymorphism (P=0.0058) in Hashimoto's thyroiditis but not for the other autoimmune endocrine diseases.

Conclusions: The CYP27B1 promoter (−1260) C/A polymorphism appears to be associated with endocrine autoimmune diseases but the CYP27B1 intron 6 (+2838) C/T polymorphism appears to be associated only with Hashimoto's thyroiditis. These results imply a regulatory difference of the CYP27B1 hydroxylase to predispose to endocrine autoimmunity.

Introduction

CYP27B1 (25-hydroxyvitamin D3-1α-hydroxylase) is a mitochondrial P450 enzyme (1), which catalyzes the conversion of 25-hydroxyvitamin D3 to 1,25(OH)2D3, the most active natural vitamin D metabolite. It is the key enzyme determining the rate of 1,25(OH)2D3 production. Renal (2) and extrarenal tissues express CYP27B1 suggesting endocrine as well as para- and autocrine functions of this enzyme (3). Mutations within the CYP27B1 gene, which impair CYP27B1 hydroxylase activity and cause vitamin D-dependent rickets, have been described (4). The CYP27B1 gene is located on chromosome 12q13.1-13.3, ten megabases centromeric of the vitamin D receptor (VDR) locus (5).

The secosteroid 1,25(OH)2D3 acts via the nuclear VDR, and effectively prevents the development of autoimmune diabetes mellitus (6) and autoimmune thyroiditis in animal models (7). In addition, disorders such as experimentally induced autoimmune encephalitis can be favorably influenced by administering 1,25(OH)2D3 (8).

This secosteroid exerts its immunomodulatory actions by inhibiting human leukocyte antigen (HLA) class II expression on endocrine cells (9), T cell proliferation and secretion of inflammatory cytokines that are thought to act as mediators in autoimmune tissue destruction (10). Additionally, 1,25(OH)2D3 inhibits differentiation and maturation of cultured human monocyte-derived dendritic cells into potent antigen presenting cells (11). Incubation of bone marrow cells
with an analog of 1,25(OH)₂D₃ resulted in a population of immature dendritic cells unable to produce high levels of interleukin-12 (12).

Allelic variations within the VDR gene have been implicated in mediating susceptibility to endocrine autoimmune disease (13–15). In different populations a genetic association of the HLA system with these autoimmune diseases has been found (16–19). In addition, polymorphisms within the cytotoxic T lymphocyte antigen 4 (CTLA-4) gene as well as variations within the VDR gene have been shown to confer susceptibility to these diseases (20–22).

Given the shared genetic susceptibility among endocrine autoimmune disorders and the association of VDR variants with autoimmune disease, we investigated the distribution of two polymorphisms, one in the promoter region and another in intron 6 of the CYP27B1 hydroxylase gene in patients with Addison’s disease, thyroid autoimmune disease and type 1 diabetes mellitus compared with healthy controls. The promoter region polymorphism was selected because of the possible role in the expression of the gene. The intron 6 polymorphism was studied in order to confirm our findings of an earlier publication (23).

Subjects and methods
Subjects
All patients were recruited from the endocrine outpatient clinics at the University Hospitals of Frankfurt am Main, Freiburg and Düsseldorf, Germany.

Addison’s disease was diagnosed by primary adrenocortical insufficiency without evidence of tuberculosis or adrenoleukodystrophy. Adrenal autoantibodies were detected with indirect immunofluorescence on cryostat sections; these were confirmed to be directed against 21-hydroxylase in a subgroup of patients by radioimmunoassay.

Thirty-three patients (26.4%) suffered either from thyroid autoimmune disease or were thyroid autoantibody positive as part of a polyglandular syndrome type 2 and only two patients were also affected by type 1 diabetes mellitus, also known as Schmidt’s syndrome. The age of onset varied from 15 to 42 years and no neurological deficits could be detected.

Hashimoto’s thyroiditis was diagnosed by positive thyroglobulin (Tg) and/or thyroid peroxidase antibodies, reduced echogenicity on thyroid ultrasound, and normal or elevated thyrotropin (TSH) levels.

Graves’ disease diagnosis rested on autoimmune hyperthyroidism with TSH receptor antibodies and/or ophthalmopathy.

Type 1 diabetes mellitus was diagnosed according to World Health Organization criteria. The median age at diagnosis was 10.5 years (range 1–37 years).

Healthy controls (n = 320), which were collected at random from the population in Frankfurt am Main, Germany, had no family history of type 1 diabetes mellitus, Hashimoto’s thyroiditis, Graves’ disease or Addison’s disease. Although the adrenal function of the controls was not formally assessed, they had normal thyroid function and were thyroid autoantibody negative. All individuals were of Caucasian origin. The study protocol was approved by the Ethics Committee of the University Hospital, Frankfurt am Main and written informed consent was obtained from all patients and controls.

Genotype analysis
DNA was extracted from whole blood according to standard protocols. Patients with Addison’s disease (n = 125), Hashimoto’s thyroiditis (n = 121), Graves’ disease (n = 85), type 1 diabetes mellitus (n = 252) and healthy controls (n = 125) were studied for the CYP27B1 intron 6 (+2838) C/T polymorphism (Genbank accession no. AF072470). The CYP27B1 promoter (−1260) C/A polymorphism (Genbank AB006987) aligned with promoter sequence from Kong et al. (24) was analyzed in 124 patients with Addison’s disease, 139 with Hashimoto’s thyroiditis, 334 with Graves’ disease, 220 with type 1 diabetes mellitus and 320 healthy controls, using polymerase chain reaction followed, for the first polymorphism, by single strand conformation polymorphism (SSCP) analysis, as described previously (25). The amplified fragment of the CYP27B1 promoter (−1260) polymorphism was digested with the restriction enzyme TfiI (New England Bio Labs, Beverly, MA, USA) according to the manufacturer’s instructions. Digestion product was separated on 2.5% agarose gel. The gel was visualized by SYBR green staining and ultraviolet illumination. The two polymorphisms are separated by an interval of 4471 bp.

Statistical analysis
Observed and expected genotype frequencies were compared based on the Hardy–Weinberg equation as well as power calculations performed (based on the observed homozygote/heterozygote frequencies in patients and in controls) using BiAS statistical package version 7.01 (Epsilon, Weinheim, Germany). Patients and controls were compared using allele-wise and genotype-wise X² testing. Probabilities (P) were regarded as significant if P < 0.05 using BiAS software.

Results
The observed CYP27B1 intron 6 (+2838) C/T polymorphism and CYP27B1 promoter (−1260) C/A polymorphism genotype frequencies were in accordance with the Hardy–Weinberg equilibrium in the groups with autoimmune disease as well as in the control group (data not shown).
Analysis of the CYP27B1 intron 6 (+2838) C/T polymorphism (see Table 1) showed no differences in either of the genotypes between patients with Addison’s disease, Graves’ disease, type 1 diabetes mellitus and controls ($P = 0.2082$, $P = 0.0991$, $P = 0.9285$ respectively). However, significantly more patients with Hashimoto’s thyroiditis were homozygous for TT than healthy controls (64.5% vs 44.8%, $P = 0.0058$). Thus, the gene frequency of the allele T was higher in patients with Hashimoto’s thyroiditis than in the control group (79.8% vs 66.8%, $P = 0.0016$; see Table 1).

The gene typing of the CYP27B1 promoter (−1260) C/A polymorphism (Table 2) revealed that the CC genotype was significantly more frequent in patients with Addison’s disease, than in healthy controls (51.6% vs 39.7% respectively, $P = 0.0062$). Patients with Hashimoto’s thyroiditis and type 1 diabetes mellitus also had higher frequencies of the allele C than controls (73.74% and 74.10% vs 66.1%, $P = 0.0269$ and $P = 0.0062$ respectively; Table 2). However, a borderline association was found for the frequency of the allele C between Graves’ disease and controls ($P = 0.0581$), whereas it did not show any difference ($P = 0.3354$) for patients with Addison’s disease.

When all patients with an autoimmune disease were considered as one group and compared with the controls, the allele C was significantly more frequent in the group with endocrine autoimmune diseases than in controls (66.1% vs 72.15%, $P = 0.0051$) (data derived from Table 2).

A power calculation was performed for each group and a power of 86%, 71%, 83% and 93% (Addison’s disease, Hashimoto’s thyroiditis, Graves’ disease and type 1 diabetes mellitus respectively) was obtained for the CYP27B1 promoter (−1260) C/A polymorphism and a power of 25%, 24%, 61% and 4% (Addison’s disease, Hashimoto’s thyroiditis, Graves’ disease and type 1 diabetes mellitus respectively) was obtained for the CYP27B1 intron 6 (+2838) C/T polymorphism.

**Discussion**

In the present study, we investigated two CYP27B1 hydroxylase gene polymorphisms in the susceptibility to Addison’s disease, Hashimoto’s thyroiditis, Graves’

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**Table 1** Distribution of the CYP27B1 intron 6 (2338 T/C) polymorphism in patients with Addison’s disease, Hashimoto’s thyroiditis, Graves’ disease, type 1 diabetes mellitus and healthy controls.

<table>
<thead>
<tr>
<th>Polymorphism</th>
<th>Controls</th>
<th>Addison’s disease</th>
<th>Hashimoto’s thyroiditis</th>
<th>Graves’ disease</th>
<th>Type 1 diabetes mellitus</th>
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<td></td>
<td>n</td>
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<td>n</td>
<td>%</td>
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</table>

*P*-values are given for the comparison of controls and the patients with the respective disease.

Polyorphism: $P_{Addison} = 0.2082$, $P_{Hashimoto} = 0.0068$, $P_{Graves} = 0.0991$, $P_{Diabetes} = 0.9285$.

Gene frequencies: $P_{Addison} = 0.1177$, $P_{Hashimoto} = 0.0016$, $P_{Graves} = 0.5592$, $P_{Diabetes} = 0.8834$.

**Table 2** Distribution of the CYP27B1 promoter (−1260 C/A) polymorphism in patients with Addison’s disease, Hashimoto’s thyroiditis, Graves’ disease, type 1 diabetes mellitus and healthy controls.

<table>
<thead>
<tr>
<th>Polymorphism</th>
<th>Controls</th>
<th>Addison’s disease</th>
<th>Hashimoto’s thyroiditis</th>
<th>Graves’ disease</th>
<th>Type 1 diabetes mellitus</th>
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<td>Gene frequencies</td>
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*P*-values are given for the comparison of controls and the patients with the respective disease.

Polyorphism: $P_{Addison} = 0.0062$, $P_{Hashimoto} = 0.0173$, $P_{Graves} = 0.0094$, $P_{Diabetes} = 0.0038$.

Gene frequencies: $P_{Addison} = 0.3354$, $P_{Hashimoto} = 0.0269$, $P_{Graves} = 0.0581$, $P_{Diabetes} = 0.0062$. 
disease and type 1 diabetes mellitus. In our population the allele C of the CYP27B1 promoter −1260 polymorphism was found significantly more often in the group with an autoimmune endocrine disease than in the control group. Based on these results, the CYP27B1 promoter −1260 C polymorphism is associated with susceptibility to each autoimmune endocrine disease in the German population. These findings complement our observations that VDR polymorphisms are associated with Addison’s disease, type 1 diabetes and autoimmune thyroid diseases (13–15, 26) and the alternate allele CYP27B1 (-1260) A might confer protection.

We previously reported that the CYP27B1 +2838 C/T polymorphism was not associated with susceptibility to thyroid autoimmune disease in our population (23), but the size of the group with Hashimoto’s thyroiditis in that publication was small. In the present investigation in a larger patient sample, we find this polymorphism to be associated with Hashimoto’s thyroiditis. These results illustrate the importance of larger samples sizes as well as small P values (27) to corroborate the role of markers in susceptibility, in this case, to autoimmune diseases.

Therefore, another component of the vitamin D system appears to be associated with autoimmune endocrine disease. Inflammatory cytokines induce the expression of CYP27B1 in murine macrophages and this induction in macrophages of diabetic NOD mice was shown to be defective (28) corresponding to an increased type 1 diabetes risk. CYP27B1 knockout mice displayed enlarged lymph nodes in the vicinity of the thyroid gland and a reduction in peripheral CD4+ and CD8+ T lymphocytes (29).

Since our findings were obtained in a large sample size, the CYP27B1 gene and its promoter (−1260) C/A polymorphism appears to be a candidate gene for genetic susceptibility to autoimmune endocrine diseases. Corresponding to these results are data from a family study of type 1 diabetes mellitus where we have observed an increased transmission rate of allele C of the (−1260) CYP27B1 promoter polymorphism, confirming its role as a susceptibility factor (unpublished data). Our present investigation extends these findings to a larger group of patients and to other autoimmune endocrine diseases. The different regulation of hydroxylase would affect the local abundance of 1,25(OH)2D3, and influence the microenvironment through antigen presenting, dendrite and regulatory immune cells.

Since the promoter variant is located in the promoter region of the key enzyme of vitamin D metabolism, it may affect enzyme transcription and thus the rate of final hydroxylation of 1,25(OH)2D3.

However, functional investigations on the effect of this polymorphism on macrophage and lymphocyte function in patients and controls are needed to put these findings into perspective.

References


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