Genetic variation in the human vitamin D receptor is associated with muscle strength, fat mass and body weight in Swedish women

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Abstract

Objective: Bone mineral density (BMD) is under strong genetic control and a number of candidate genes have been associated with BMD. Both muscle strength and body weight are considered to be important predictors of BMD but far less is known about the genes affecting muscle strength and fat mass. The purpose of this study was to investigate the poly adenosine (A) repeat and the BsmI SNP in the vitamin D receptor (VDR) in relation to muscle strength and body composition in healthy women.

Design: A population-based study of 175 healthy women aged 20–39 years was used.

Methods: The polymorphic regions in the VDR gene (the poly A repeat and the BsmI SNP) were amplified by PCR. Body mass measurements (fat mass, lean mass, body weight and body mass index) and muscle strength (quadriceps, hamstring and grip strength) were evaluated.

Results: Individuals with shorter poly A repeat, ss and/or absence of the linked BsmI restriction site (BB) have higher hamstring strength (ss vs LL, P = 0.02), body weight (ss vs LL, P = 0.049) and fat mass (ss vs LL, P = 0.04) compared with women with a longer poly A repeat (LL) and/or the presence of the linked BsmI restriction site (bb).

Conclusions: Genetic variation in the VDR is correlated with muscle strength, fat mass and body weight in premenopausal women. Further functional studies on the poly A microsatellite are needed to elucidate whether this is the functionally relevant locus or if the polymorphism is in linkage disequilibrium with a functional variant in a closely situated gene further downstream of the VDR 3'UTR.

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Introduction

Body composition, including bone mineral content, fat mass and lean mass, is known to be under strong genetic control with a suggested heritability of fat mass and lean mass of 52–57% (1). Several candidate genes have been identified that may contribute to bone mineral density (BMD), far less is known about genes affecting muscle strength and fat mass. The gene for the vitamin D receptor (VDR) is one of the genes that has been most extensively studied in relation to BMD, due to the important role of vitamin D in bone metabolism (reviewed in 2). A functional genetic polymorphism in the VDR could be involved in other tissues that respond to vitamin D, such as muscle cells and adipocytes. It has been demonstrated that skeletal muscle is a target organ for vitamin D, and that vitamin D metabolites directly affect muscle cell metabolism through various pathways (3). Also, vitamin D deficiency is associated with muscle weakness (3, 4). Moreover, it has been demonstrated that vitamin D stimulates the differentiation of preadipocytes to adipocytes in the OB 17 cell line (5). Some studies have also suggested a relationship between body size, as measured by body length and weight, and allelic variations of the VDR gene (6–8), indicating that VDR genotype may function as a determinant of body composition.

Thus far, the association of VDR polymorphisms and muscle strength in a population-based study has not been studied and only Geusens et al. (9) have reported a correlation between VDR alleles and muscle strength in a study of non-obese, elderly women. The association studies made on the VDR gene have mainly focused on the different single nucleotide polymorphisms (SNPs)
rather than on the poly A microsatellite at the VDR 3′UTR – a region that has been postulated to be important for mRNA stability (reviewed in 10).

This study examines the relationship between a poly adenosine (A)-repeat and a BsmI SNP in the gene for the VDR on variation in muscle strength, fat mass, lean mass, body weight and body mass index (BMI) in a Swedish female population-based cohort.

Subjects and methods

Subjects

The study cohort consisted of a total of 175 women, aged 20–39 years, randomly selected from the population registry of the county of Uppsala. In all, 347 women were randomly selected and invited by mail to participate. Of the invited women, 172 did not participate and were excluded, of these nine were pregnant, 26 had moved, 98 did not contact the research centre and were not reached despite repeated letters, and 39 were not interested in participating. In total 175 (50%) of the invited women attended the investigation and were included. Of these 175 individuals, 169 were examined for left quadriceps and hamstring strength, 172 for right quadriceps and hamstring strength, 171 for left handgrip strength and 172 for right handgrip strength. This resulted in a total of 169 women all examined for bilateral quadriceps strength, bilateral hamstring strength and bilateral handgrip strength. The main reason for the absence of muscle strength data was instrument failure. Blood samples for DNA extraction were obtained from 168 individuals and the samples were stored at −20°C until processing and analysis. Subjects answered a questionnaire regarding, for example, medications, menstrual status, oral contraceptives and number of children. To increase the possibility of having a genetically homogenous cohort individuals with non-Swedish origin were excluded, resulting in a total of 153 individuals included in the genetic association study. The study was approved by the local ethics committee (Dnr 991007).

Muscle strength measurements

Handgrip strength was measured by a JAMAR hydraulic hand dynamometer (5030F1, Jackson, MI, USA) and expressed in kg. Measurements were made three times on both hands and the mean values from the right and left hand were used in the comparison. Isokinetic knee-flexion and extension strength was measured by a Cybex II dynamometer (Lumex, Corp., Bay Shore, NY, USA) with a modified level arm (Aero Technical Corp., Stockholm, Sweden and devised by Knutsson and Litton) and an examination table (Alfex). Muscle strength was expressed in Newton metres (Nm) and calibration of the Cybex II has been described in detail elsewhere (11). The subjects were strapped in a sitting position with adjustable belts around the thigh and chest. The axis of the level arm of the dynamometer was adjusted after the axis of the knee joint. The range of motion measured was between full extension (0°) to 90° of knee flexion and back to full extension. The angular velocity used was 90°/s. Three measurements were performed and the best values for flexion and extension were used to calculate the mean values for left and right leg. Moreover, mean values for left and right flexion strength and for left and right extension strength were used in the association study.

Measurements of body composition, height and weight

Fat mass and lean mass were measured by dual-energy X-ray absorptiometry (DXA), using a DPX-IQ scan (Lunar Corp., Madison, WI, USA) and expressed in kg. The stability of the DPX-IQ equipment was checked every morning using a phantom. The long-term precision for L2–L4 BMD, expressed as the coefficient of variance was determined by daily measurements of a lumbar spine phantom and was less than 0.5% during the study period. Height and weight were assessed at a clinical examination by Holtain stadiometer (Holtain Ltd, Crosswell, UK) and digital scale (EKS) prior to the DXA scanning.

DNA analysis

Genomic DNA was extracted from leukocytes from 3 ml EDTA blood using a Wizard Genomic DNA purification kit (Promega, Madison, WI, USA), typically yielding approximately 50–75 μg genomic DNA/sample.

The VDR poly A microsatellite in the 3′UTR and the BsmI SNP in intron 8 were amplified by PCR using primers and conditions as previously reported (12).

Statistical analysis

All statistical calculations were performed using STATISTIKA software (StatSoft Inc., Tulsa, OK, USA) by ANOVA comparing the means for the differences of the continuous variables between genotype groups. Multiple regression analysis was performed to correct for confounding factors, thus the genotype information was entered into the model along with age, fat mass and lean mass. The R^2 value is the coefficient of determination and explains the model’s relative contribution to the variation of muscle strength. Hardy–Weinberg equilibrium at the different loci was tested by exact tests and the VDR polymorphisms were analysed for linkage disequilibrium by the Fisher’s test, using the GENEPOP web version 3.1c (http://wbiomed.curtin.edu.au/genepop).

Results

The number of poly A repeats in the VDR gene varied between 11 and 23 (Fig. 1), as determined after
sequencing of representative gene fragments (data not shown). The genotypes with 18 repeats were denoted short (s) and genotypes with ≥18 repeats were denoted long (L). This cut-off corresponds to the naturally occurring bimodal distribution of the genotypes in our study population (Fig. 1). The genotypes and frequencies for the BsmI SNP were: 14% BB (absence of the BsmI restriction site on both alleles), 48% Bb (heterozygous for the restriction site) and 38% bb (presence of the restriction site on both alleles). For the poly A microsatellite, 14% were homozygous for the short allele (ss), 45% were heterozygous (sL) and 41% were homozygous for the long allele (LL). The frequency distribution of the poly A and BsmI genotypes were similar to frequencies reported in other Caucasian study groups (13) and both polymorphisms were in Hardy–Weinberg equilibrium. Furthermore, a strong linkage was found between the s and B alleles (Chi² = 291, P < 0.0001) (Table 1). Clinical characteristics of the study population are presented in Table 2.

VDR genotype and muscle strength

A significant association was found between VDR genotype and muscle strength in the study population. Women carrying the poly A genotype ss and/or the linked BsmI genotype BB had higher hamstring strength as compared with individuals in the LL and bb genotype group (ss 62.8 ± 9.0 Nm vs LL 55.6 ± 10.2 Nm, P value 0.02; BB 62.1 ± 8.7 Nm vs bb 55.5 ± 10.2 Nm, P value 0.03; ANOVA and Fisher’s LSD) (Fig. 2). To investigate whether the VDR genotype effect on muscle strength was modified by confounding factors, age, fat mass and lean mass were entered into a multiple regression model with the genotype information (Table 3). The results demonstrated borderline significant correlations of the BsmI (P = 0.05) and poly A (P = 0.06) genotypes and hamstring strength when taking age, fat mass and lean mass into account. No significant associations were found between VDR genotypes and quadriceps strength and between VDR genotypes and handgrip strength.

VDR genotype and body composition

The clinical characteristics of the poly A and BsmI genotype groups are shown in Table 4. Body weight and fat mass were higher in women carrying the poly A genotype ss compared with individuals in the LL genotype group (ss 69.4 ± 10.4 kg vs LL 64.2 ± 8.6 kg, P value 0.049; ss 23.7 ± 7.8 kg vs LL 19.9 ± 6.9 kg, P value 0.04). Similar results were seen concerning the effect of VDR BsmI genotype on body weight and fat mass (BB 69.1 ± 10.5 kg vs bb 64.4 ± 8.7 kg, P value 0.08; BB 23.9 ± 7.9 kg vs bb 20.1 ± 7.0 kg, P value 0.049).

Table 1 Contingency table for genotypic disequilibrium. The linkage disequilibrium was tested by the Fisher’s test using the GENEPOP web version 3.1c.

<table>
<thead>
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<th>Bb</th>
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<td>73</td>
<td>59</td>
<td>153</td>
</tr>
</tbody>
</table>

Chi² = 291; P < 0.0001.
The same trend was observed regarding the effect of the VDR genotypes on BMI (ss 24.5 ± 3.9 vs LL 23.0 ± 2.9, P value 0.08; BB 24.5 ± 3.9 vs bb 23.0 ± 3.0, P value 0.09).

Discussion

In this report we have demonstrated associations of the VDR genotype with muscle strength in a female population-based study. Our results show that individuals in the ss genotype group had 11.5% higher muscle strength, as measured at the hamstrings, compared with women in the LL genotype group. Thus, the results showed a strong association of VDR genotype with hamstring strength, and a similar pattern for the effect on quadriceps. This is in line with the quadriceps being the major load-bearing muscle during normal activity, and which may therefore be more susceptible to environmental factors, i.e. exercise, resulting in a proportionately larger genetic component of hamstring strength than of quadriceps strength. Vitamin D deficiency is associated with muscle weakness and low muscle strength is an important risk factor for development of osteoporotic fractures (14). VDR has been found in human skeletal muscle cells and affects muscle cell metabolism by binding to vitamin D metabolites (14, 15). We have previously shown an association between VDR genotype and BMD, where individuals in the ss genotype group had significantly higher lumbar spine BMD compared with the LL genotype group (12). Taken together, this correlates well with muscle strength being under genetic control and associated with BMD.

To our knowledge, this is the first study that demonstrates a significant association of VDR alleles and muscle strength in a population-based cohort of premenopausal women. However, Geusens et al. (9) have reported a correlation of a VDR polymorphism and muscle strength in postmenopausal women but the significance was only found in the subgroup of non-obese (BMI < 30 kg/m²) women. In contrast, the effect of VDR polymorphisms on muscle strength in our study was shown in the entire cohort. However, exclusion of obese subjects (BMI > 30 kg/m², n = 6) showed stronger correlations of VDR polymorphisms with muscle strength (data not shown), which is in agreement with Geusens et al. (9).

In addition, we found an association of the VDR genotype with body composition. Our results showed that individuals with short poly A repeat (ss) and/or absence of the linked BsmI restriction site on both alleles (BB)
have significantly higher body weight and fat mass. Previous studies have demonstrated a relationship between body size in children and allelic variants of the VDR gene, with which our findings agree. One study in 589 French children reported that at 2 years of age body weight was greater in girls with the BB genotype compared with individuals in the bb genotype group (6). Furthermore, a retrospective study in infant health records of 66 postmenopausal British women showed that those with the BB genotype had 7% higher weight than those with the bb genotype at 1 year of age (7). Another study of 309 unrelated French subjects with early onset type 2 diabetes suggests that variations in the VDR gene are associated with susceptibility to obesity (16). However, the pathophysiological mechanisms of these associations remain unexplained. A direct effect of vitamin D on adipocyte differentiation and metabolism is a possible mechanism, as VDR is expressed in preadipocytes (17). Dace et al. (5) showed that calcitriol stimulates the differentiation of preadipocytes to adipocytes in the OB 17 cell line, which provides arguments for a probable role of vitamin D in adipose differentiation by binding to the VDR.

The molecular mechanisms behind the genotype/phenotype associations observed for these polymorphisms in the VDR gene are still unclear. The BsmI SNPs, situated in intron 8, are non-coding and do not change the amino acid sequence of the VDR protein and, as far as we know, the BsmI SNP is non-functional. However, we and others have shown that the BsmI B allele is in linkage disequilibrium with the poly A s allele in the VDR 3′UTR. There are several studies trying to identify whether this poly A microsatellite affects mRNA stability (18), but results have been inconclusive (19). A possible hypothesis is that a long poly A repeat is less stable and translated less efficiently into VDR protein, resulting in a decreased response to vitamin D and thus an inhibition of muscle cell and adipocyte differentiations. This is in line with Carling et al. (20) who have showed that B alleles are associated with higher VDR mRNA levels in a study of patients with sporadic primary hyperparathyroidism. However, further functional studies on the poly A microsatellite are needed to elucidate whether this is the functionally relevant locus or if the polymorphism is in linkage disequilibrium with a functional variant in a closely situated gene further downstream of the VDR 3′UTR.

This is, to our knowledge, the first study that demonstrates a significant association of the VDR poly A microsatellite and the linked BsmI SNP to muscle strength and body composition in premenopausal women. In conclusion, further studies are required to elucidate the molecular mechanism behind the genotype/phenotype associations and to investigate whether the VDR 3′UTR poly A microsatellite has a functional role.

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References

8 Burger-Lux MJ, Heaney RP, Hayes J, Deluca HE, Johnson ML & Gong G. Vitamin D receptor gene polymorphism, bone mass,

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**Table 4 Clinical features of the study population in relation to VDR poly A and BsmI genotype. Values are means ± S.D.**

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>LL (n = 63)</th>
<th>SL (n = 68)</th>
<th>SS (n = 22)</th>
<th>P value</th>
<th>bb (n = 59)</th>
<th>Bb (n = 73)</th>
<th>BB (n = 21)</th>
<th>P value</th>
</tr>
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<tr>
<td>Weight (kg)</td>
<td>64.2 ± 8.6</td>
<td>66.4 ± 12.1</td>
<td>69.4 ± 10.4</td>
<td>&lt; 0.05*</td>
<td>64.4 ± 8.7</td>
<td>66.3 ± 11.9</td>
<td>69.1 ± 10.5</td>
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<td>Height (cm)</td>
<td>167.1 ± 4.8</td>
<td>168.5 ± 6.3</td>
<td>168.6 ± 6.5</td>
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<td>BMI (kg/m²)</td>
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<td>Fat mass (kg)</td>
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<td>20.9 ± 8.1</td>
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<td>&lt; 0.05*</td>
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1’ss genotype group vs LL genotype; *BB genotype group vs bb genotype. *P < 0.05.


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