CASE REPORT

Pharyngeal pituitary non-functioning adenoma with normal intra-sellar gland: massive tumor shrinkage on octreotide therapy

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Abstract

Objective: Functioning or non-functioning ectopic tumors may develop from pharyngeal pituitary remnants. They constitute <1% of all obstructive pharyngeal masses and they have a strong tendency to bleed. We report a case of a non-functioning ectopic pituitary adenoma of the rhino-pharynx studied over a long-term somatostatin analog treatment.

Patient and treatment: A 60-year-old woman presented with severe posterior epistaxis. She had complained of nasal obstruction for the past 2 years. Magnetic resonance imaging (MRI) and endoscopic examination revealed a 2 cm exophytic, bleeding mass in the cavum, which was judged inoperable, and a biopsy was performed. On immunostaining, tumor cells were positive for pancytokeratins MNF 116 and C11, epithelial membrane antigen, chromogranin and neuron-specific enolase (NSE), and negative for synaptophysin, desmin, actin, estrogen and progesterone receptors, all anterior pituitary hormones and human chorionic gonadotropin. Blood levels of the above hormones and tumor markers were normal, except for a moderate elevation of NSE (33.8 μg/l, normal value <12 μg/l). It was concluded that this was a non-functioning pituitary adenoma of the rhino-pharynx. MRI showed a normal intra-sellar pituitary gland, including the normal bright signal of the posterior lobe. Somatostatin receptor scintigraphy (SRS) disclosed intense tracer uptake in the tumor, indicating high somatostatin receptor content. There was also an intense uptake in the intra-sellar pituitary. Therapy with long-acting octreotide was started, 20 mg per month i.m.

Results: The patient has been on octreotide for the last 12 months. Nasal obstruction rapidly subsided and bleeding did not recur. Repeated endoscopic examinations showed rapid tumor reduction, the mass shrinkage being almost complete at 3 months. This was confirmed by MRI, while SRS showed markedly decreased uptake in the residual tumor and the intra-sellar pituitary, and NSE became normal.

Conclusion: Pharyngeal pituitary remnant adenomas are rare, but they must be considered in the differential diagnosis of bleeding or obstructive masses of the rhino-pharynx. In this case, the positive SRS influenced the choice of octreotide, as an alternative to surgery. As we show for the first time in this location, octreotide can exert prolonged and marked anti-tumoral effects in non-functioning adenoma.

European Journal of Endocrinology 148 357–364

Introduction

Ectopic pituitary tumors are a rare cause of pharyngeal obstructive or bleeding masses, but remnants of the Rathke pouch can be found along the cranio-pharyngeal migration route towards the sellar area in practically every individual, when carefully sought (1, 2). However, their function remains unknown (1, 4–6). Tumors developing from those remnants are usually referred to as ectopic pituitary tumors. Since the first description by Erdheim (3) there have been about 74 reported cases. Their localization and type of secretions are summarized in Table 1. They are found most often in the sphenoidal sinus and the supra-sellar region and less frequently in the rhino-pharynx with only a few cases reported in this localization. Non-functioning and adrenocorticotropin (ACTH)-secreting tumors are the most frequent types.

Therapy of those masses has been in general surgical, often followed by external radiotherapy. There have been
attempts at treatment of ectopic pharyngeal pituitary with dopamine agonists, which succeeded in lowering hormone secretion (7, 8), but to the best of our knowledge there has been no attempt at medical therapy with somatostatin analogs. It must be stressed that the endocrine nature of these tumors is usually recognized after immunohistochemical study of the excised tissue, and that hormonal secretion into blood has seldom been assessed. We report here the first case of a pharyngeal pituitary tumor which is being successfully treated with a long-acting preparation of octreotide.

Case report

A 60-year-old woman presented with a first episode of intense posterior nasal bleeding lasting for several days and requiring emergency care. During the previous 2 years she had noticed progressive nasal obstruction. There was no headache or other local symptoms. She was on metformin for type 2 diabetes and on an angiotensin-converting enzyme inhibitor for mild hypertension, both well controlled, and on no other drugs. Posterior rhinoscopy followed by panendoscopy using a rigid adult bronchoscope showed a fungating, hard mass easily bleeding on contact, which was judged inoperable, and a biopsy was performed.

The patient presented no endocrine abnormalities and was not receiving estrogen replacement therapy. After the histological diagnosis was carried out, a long-acting preparation of octreotide (Sandostatin LAR; Novartis, Basle, Switzerland) was administered at a dose of 20 mg i.m. every 4 weeks.

Materials and methods

MRI was performed by using a 1.5 T MR unit (Magnetom Symphony; Siemens Medical System, Iselin, NJ, USA) with a quadrature head coil. The following MRI imaging sequences were used: T1 sagittal, transverse, T2 fatsat transverse, three-plane spin echo fatsat T1-weighted after gadolinium injection. Somatostatin receptor scintigraphy (SRS) was performed after injection of 190 MBq $^{111}$In-DTPA-o-Phe-1-octreotide (OctreoScan; Mallinckrodt Medical, Petten, The Netherlands). Whole-body scans, planar images (500 000 counts, matrix 256 x 256) and single photon emission computed tomography (SPECT, matrix 128 x 128, zoom 1.2, 360°, 120 projections, 45 s per frame) of the head were obtained 3 and 24 h after injection using 20% windows centered over both $^{111}$In energy peaks (172 and 245 keV). SPECT images were reconstructed with a Butterworth filter (cut-off 0.7, order 5.0) and transverse, coronal and sagittal sections were used for final diagnosis.

The biopsy specimens obtained during panendoscopy were fixed in 10% buffered formalin, embedded in paraffin, and stained with hematoxylin and eosin (H&E), periodic acid-Schiff, and Alcian blue at pH 2.5. Immunohistochemical staining was performed. To this end, 5 μm paraffin sections were rehydrated and subjected to microwave treatment for antigen retrieval. The peroxidase anti-peroxidase technique was chosen as the detection system. The antigens tested were the following (dilution and origin of antibodies in brackets) – two cytokeratins, MNF 116 (1:200, Dako, Zug, Switzerland) and C11 (1:10, Novocastra, Nunningen, Switzerland), epithelial membrane antigen (EMA), a marker of eccrine differentiation (1:100, Dako), gross cystic disease fluid protein 15, a marker of eccrine differentiation (1:250, University Hospital and Lausanne Medical School), α-2 ZN-glycoprotein (1:500, Nordic, Tilburg, Holland), S100 protein (1:2000, Dako), non-specific enolase (NSE) (1:900, Dako), glial fibrillary acidic protein (1:100, Dako), chromogranin A and B (1:100, Dako), synaptophysin (1:25, Dako), CD 34 (1:80, Immunotech, Nyon, Switzerland), desmin (1:40, Dako), actin (not diluted, Enzodiagnostics, Dossenheim, Germany), estrogen receptors (1:20, Dako), progesterone receptors (1:100, Dako), thyrotropin (TSH) (1:3000, Dako), growth hormone (GH) (1:2000, Dako), luteinizing hormone (LH) (1:3000, Dako), human chorionic gonadotropin (1:1500, Dako), ACTH (1:2000, Dako) and prolactin (PRL) (1:2500, Dako).

Table 1 Localization and secretion in 74 cases of ectopic pituitary adenoma.

<table>
<thead>
<tr>
<th></th>
<th>NSE</th>
<th>GH</th>
<th>LH</th>
<th>FSH</th>
<th>ACTH</th>
<th>PRL</th>
<th>ACTH and TSH</th>
<th>GH and PRL</th>
<th>TSH</th>
<th>CP</th>
<th>References</th>
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<tr>
<td>Total (74)</td>
<td>24</td>
<td>5</td>
<td>1</td>
<td>3</td>
<td>17</td>
<td>15</td>
<td>3</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>Sphenoidal sinus</td>
<td>6</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>5</td>
<td>6</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>17–30</td>
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<tr>
<td>Suprasellar</td>
<td>2</td>
<td>2</td>
<td>–</td>
<td>5</td>
<td>2</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>31–45</td>
<td></td>
</tr>
<tr>
<td>Nasopharynx</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>3</td>
<td>2</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>46–49</td>
<td></td>
</tr>
<tr>
<td>Cavernous sinus</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>57, 58, 61, 62</td>
<td></td>
</tr>
<tr>
<td>Clivus</td>
<td>3</td>
<td>–</td>
<td>–</td>
<td>1</td>
<td>3</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>59, 60</td>
<td></td>
</tr>
<tr>
<td>Third ventricle</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>1</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>1</td>
<td>55, 56</td>
</tr>
<tr>
<td>Pituitary stalk</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>50</td>
<td></td>
</tr>
<tr>
<td>Pharynx</td>
<td>2</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>1</td>
<td>2</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>2</td>
<td>51–54</td>
</tr>
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</table>

CP = craniopharyngioma (references 63–66); FSH = follicle-stimulating hormone.
Results

During the initial evaluation, rhinoscopic examination showed an exophytic hard mass in the left cavum necessitating a pandoscopy for a sufficient biopsy specimen. MRI showed a 2 cm spherical mass, located in the left part of the cavum, homogeneous and displaying marked contrast enhancement (Fig. 1A and B). Intraseellar anterior and posterior pituitary lobes were normal, and there was no apparent anatomical connection between the sellar content and the pharyngeal mass. The endoscopic and MRI picture were consistent with a primary epithelial rhino-pharynx cancer. The first SRS performed at diagnosis showed an uptake of the tracer by the tumor on the SPECT images performed at 3 h, which increased and became intense 24 h after injection (Fig. 2A–C). We observed a similar uptake of the tracer by the normal pituitary. Whole-body scan and planar images revealed a physiological tracer distribution.

At histological examination, the growth pattern of the tumor was trabecular with sheets and cords separated by fibrous stroma. Tumor cells were round or polygonal and had round or oval nuclei and variable amounts of acidophilic cytoplasm. Nucleocytoplasmic atypia and mitoses were absent (Fig. 3A and B). There was no mucin secretion by the tumor cells. Tumor cells were diffusely positive for cytokeratin MNF 116, EMA and chromogranin (Fig. 3C), and focally positive for cytokeratin C11 and NSE. They were negative with all other antibodies tested including those against adeno-hypophyseal hormones. The immunohistochemical profile of this tumor and particularly the presence of intra-cyttoplasmic chromogranin-positive granules was consistent with a diagnosis of an ectopic, hormone-negative pituitary adenoma.

Plasma anterior pituitary hormones were normal and NSE was moderately elevated, whereas other tumor markers were normal (Table 2).

Long-acting octreotide treatment was well tolerated, except for moderate diarrhea that subsided promptly after the adjunction of digestive enzymes. Bleeding did not recur and nasal obstruction progressively subsided. At 3 months of therapy, a repeated endoscopic study showed a striking shrinkage of the tumor that was
confirmed on MRI, where the tumor was barely distinguishable (Fig. 1C and D). A second SRS was performed 3 months after the beginning of therapy and 4 weeks after the last i.m. injection of long-acting octreotide. SPECT images of the head demonstrated a significant decrease of the tracer uptake by the tumor and the normal pituitary when compared with the initial scan (Fig. 2A, B, and C). Note the similar appearance and changes of the normal pituitary, which was clearly shown only on the diagnostic sagittal and coronal views (short arrows).

**Discussion**

Ectopic pituitary tumors constitute less than 1% of all obstructive pharyngeal masses, and although they have a particularly strong tendency to bleed, they are difficult to distinguish from more common causes on clinical grounds. They arise from ectopic pituitary remnants, which persist throughout the fetal and adult life, undergoing no involution. They can be demonstrated in aged normal individuals (9, 10). The described cases of NSE was progressively reduced to normal and insulin-like growth factor (IGF-I) showed a reduction of 31% but remained within the normal limits for age (Table 2).
adenomatous transformation of those remnants have always been associated with a normal intra-sellar pituitary disclosing no anatomical continuity with the ectopic pituitary tumor (11). The neuroendocrine nature of these masses, although difficult to establish with certainty, may prove particularly favorable for the treatment, as illustrated by our case. The clinical and MRI differential diagnoses included primary rhino-pharyngeal tumor, chordoma, chondroma, chondrosarcoma, craniopharyngioma or metastatic cancer, whereas an ectopic pituitary tumor was not taken into account initially. However, the neuroendocrine nature of this tumor was underlined by the presence of intra-cytoplasmic chromogranin-positive granules in tumor cells. This was corroborated by the high somatostatin receptor content disclosed by SRS.

Ectopic pituitary adenoma should be distinguished from pituitary adenoma extending from the sella turcica by radiological examinations. This tumor must also be differentiated from olfactory neuroblastoma, which presents a fibrillary stroma and does not express keratin, unlike pituitary adenoma. Although pituitary adenoma may show some nuclear atypia, which were absent in our case, they do not have the degree of pleomorphism, mitotic activity and necrosis that characterizes small cell neuroendocrine carcinoma and sinonasal undifferentiated carcinoma.

Surgical ablation of the mass in our patient was judged potentially mutilating. Therefore, somatostatin receptor-based therapy was envisioned as an alternative to surgery. A suppressive effect of somatostatin analogs can be anticipated when the presence of somatostatin receptors (essentially subtypes SSTR2 and SSTR5) is
shown in a tumor mass (12). Although we did not measure in vitro receptors in a tissue specimen, we assumed they were abundant in the tumor according to the results of SRS, and indeed, somatostatin analog treatment resulted in an impressive tumor mass reduction together with normalization of NSE, the unique tumor marker observed in this case. This is consistent with the well-known effects of somatostatin analogs on pituitary GH-secreting adenomas and on gastro–entero–pancreatic neuroendocrine tumors (13–15). It should be noted that treatment with somatostatin analogs usually does not result in an anti-tumoral effect on non-functioning intra-sellar pituitary adenomas (16), contrary to the non-functioning pharyngeal pituitary adenoma discussed here.

In conclusion, to the best of our knowledge this is the first case of a pharyngeal pituitary adenoma in which a high somatostatin receptor content was demonstrated by SRS and which was successfully treated with a somatostatin analog, thus avoiding surgery. However, the duration of the ongoing anti-tumoral effect cannot be predicted, and the question remains open whether long-term octreotide therapy would modify the operability, should tumor regrowth occur.

References


Table 2 Plasma hormones and neuroendocrine tumor markers before and after octreotide treatment.

<table>
<thead>
<tr>
<th>Time on octreotide</th>
<th>Pituitary hormones</th>
<th>Tumor markers</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Before</td>
<td>Two months</td>
</tr>
<tr>
<td>LH (UI/l)</td>
<td>49.9</td>
<td>–</td>
</tr>
<tr>
<td>FSH (UI/l)</td>
<td>59</td>
<td>–</td>
</tr>
<tr>
<td>Free-α subunit (UI/l)</td>
<td>0.55</td>
<td>–</td>
</tr>
<tr>
<td>PRL (μg/l)</td>
<td>5</td>
<td>–</td>
</tr>
<tr>
<td>ACTH (ng/l)</td>
<td>13</td>
<td>–</td>
</tr>
<tr>
<td>Cortisol (nmol/l)</td>
<td>188</td>
<td>–</td>
</tr>
<tr>
<td>IGF-I (μg/l)</td>
<td>98</td>
<td>68</td>
</tr>
<tr>
<td>TSH (mU/l)</td>
<td>1.79</td>
<td>2.96</td>
</tr>
<tr>
<td>FT4 (pmol/l)</td>
<td>14</td>
<td>12</td>
</tr>
<tr>
<td>NSE (μg/l)</td>
<td>33.8</td>
<td>22.1</td>
</tr>
<tr>
<td>PP (pmol/l)</td>
<td>47</td>
<td>–</td>
</tr>
<tr>
<td>GRF (pmol/l)</td>
<td>47</td>
<td>–</td>
</tr>
<tr>
<td>GAWK (pmol/l)</td>
<td>36</td>
<td>–</td>
</tr>
</tbody>
</table>

* Postmenopausal values. ** p.m. values.

FSH = follicle-stimulating hormone; PP = pancreatic polypeptide; GRF = growth hormone-releasing factor; GAWK = GAWK, a sequence of chromogranin B; FT4 = free thyroxine.
References:


Received 25 October 2002
Accepted 4 December 2002