CLINICAL STUDY

Reduced microvascular perfusion and reactivity in adult GH deficient patients is restored by GH replacement

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Abstract

Background: An increased cardiovascular risk and mortality in hypopituitary patients receiving conventional hormonal treatment without GH replacement have been shown in several studies. Various atherogenic risk factors including endothelial dysfunction – an early event in the atherogenesis – are more expressed in adults with GH-deficiency (GHD). Changes in microcirculation and vascular reactivity could represent an early marker of developing vascular changes.

Objective: To evaluate the microcirculation and vascular reactivity in a GHD state before and during GH replacement.

Subjects, methods and design: Thirteen adult patients (ten men, mean age 40±9 years) with severe GHD were studied. The skin microvascular perfusion and reactivity were measured by laser-Doppler flowmetry on the forearm. Two dynamic tests for vascular perfusion and reactivity were used – post-occlusive reactive hyperemia (PORH) and thermal hyperemia (TH) at 44°C. Measurements were performed before and after 6 and 12 months on GH replacement with a dose of GH that normalized IGF-I serum levels. The parameters of tissue perfusion and vascular reactivity measured in GHD were compared with values during GH treatment and with the results of the control group.

Results: Peak flow during TH in GHD patients was significantly reduced before GH treatment when compared with healthy subjects (means±S.E.M., 68±6.6 vs 111±8.3 perfusion units (PU), P < 0.001) and normalized on GH treatment (109±12.7 PU). The velocity of perfusion increase during TH before treatment was significantly reduced in GHD as well (0.84±0.07 vs 1.53±0.19 PU/s, P < 0.03) and normalized on GH treatment (1.38±0.24 PU/s). The PORH was also significantly reduced in GHD compared with controls (PORHmax 414±63 vs 528±58%, P < 0.05) and during GH treatment was restored to values not different from controls (642±86%, P = NS).

Conclusions: Skin microcirculation and vascular reactivity measured by laser-Doppler flowmetry is significantly reduced in GHD adults and is restored during GH replacement therapy. Reduced tissue perfusion and reactivity probably reflect the endothelial dysfunction in the GHD state. Reduced nitric oxide production and bioavailability and also other factors like increased sympathetic activity and reduced conversion of thyroxine to triiodothyronine in the GHD state can contribute to changes in microcirculation. Restoration of vascular reactivity by GH replacement might have favorable clinical consequences on the increased vascular morbidity of GHD patients. Reduced skin microvascular perfusion and reactivity in GHD probably contribute to the impaired thermoregulation – a clinical symptom of GHD.

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Introduction

An increased morbidity and mortality have been detected in hypopituitary adults in several studies, with the standardized mortality rate in the range 1.2–2.17 (1–6). Explanation for this observation is not clear. Some of these studies documented an increased vascular mortality (1, 4, 5), although some did not (2, 3, 6). Changes in various atherogenic risk factors have been demonstrated in growth hormone (GH) deficiency (GHD) – lipids, coagulation factors, insulin resistance, blood pressure, enhanced oxidative stress, cardiac function, inflammatory markers interleukin-6 and C-reactive protein and also the endothelial dysfunction measured by flow-mediated dilatation (FMD) (7–9). Endothelial dysfunction is considered to be an early marker of atherogenesis. Endothelium is a very active secretory organ producing...
many substances influencing various processes. One of its key functions is the regulation of blood circulation. Changes in the endothelial function can be manifest for instance by changes in the endothelium-dependent vascular reactivity.

Measuring skin microvascular reactivity by laser-Doppler flowmetry (LDF) is a method for assessing dynamic parameters of tissue perfusion to the depth of approximately 1 mm. Perfusion can be thus observed in skin capillaries, arterioles, venules and in arteriovenous shunts. LDF is performed non-invasively, is easy for both patients and staff and gives reproducible results. LDF is used, for instance, in plastic surgery to assess the viability of transplanted grafts of skin, in diabetology for evaluation of microangiopathy, in angiology for the diagnosis of Raynaud’s syndrome and for evaluation of tissue perfusion in other fields of medicine. The use of LDF remains currently mostly in clinical research and the indications for daily practice are under evaluation. The most important factors influencing LDF measurements are the room and skin temperature, smoking, vasoactive medication, and any mental stress which could cause peripheral vasoconstriction.

The aim of the study was to evaluate the microcirculation and vascular reactivity in a GHD state before and during GH replacement. Changes in microcirculation and vascular reactivity could represent an early marker of developing vascular changes.

**Study protocol, patients and methods**

Thirteen adult patients (ten men, mean age 40±9 years, body mass index (BMI) 28.4±3.5 kg/m²) with severe GHD (GH < 3 μg/l during an insulin tolerance test) were studied. Two patients had childhood-onset GHD and the rest adult-onset GHD. The causes of GHD were non-functioning pituitary adenoma and its treatment (in seven patients), prolactinoma (two), idiopathic hypopituitarism (two), pituitary adenoma with acromegaly (one) and with Cushing’s disease (one) both inactive after treatment. All the patients were normotensive, non-diabetics, non-smokers and were not using drugs influencing vascular perfusion (except for adequate replacement of hypopituitarism). All the patients were on stable replacement therapy – six on gonadal, hydrocortisone, thyroid and desmopressin replacement, four patients on replacement of three axes, one of two axes, two of one axis.

The skin microvascular perfusion and reactivity were measured by LDF on the forearm. Two dynamic tests for microvascular reactivity were used – postocclusive reactive hyperemia (PORH) and thermal hyperemia (TH) at 44 °C. Measurements were performed before and after 6 and 12 months on GH replacement with a stable dose of GH that normalized insulin-like growth factor-I (IGF-I) serum levels. The dose of GH was at the beginning titrated to get IGF-I into the normal range. Mean stable GH dose was 0.92±0.45 IU/day (mean±s.d., range 0.4–2.0). The parameters of tissue perfusion and vascular reactivity measured in GHD were compared with values during GH replacement and with the results of the control group consisting of 18 healthy subjects (nine men, mean age 46±9 years). There were no significant differences in the microcirculation parameters between sexes in the control group.

LDF measurements were performed using a Periflux 4001 apparatus (Perimed, Järfälla, Sweden) after overnight fasting on all persons between 0800 and 0900 h after at least 30 min of rest at room temperature (22 °C) in the sitting position. The LDF heating probe was fixed on the ventral part of the forearm. The temperature of the probe was set to 32 °C thermal stabilization of the skin and maintained by a heating unit (Peritemp 4005; Perimed). After 5 min of stabilization the basal flow was recorded. Occlusion of the brachial artery was then performed for 3 min by inflating the tourniquet of a sphygmomanometer to 20 mmHg above the systolic blood pressure. After fast decompression the flow changes were recorded to evaluate PORH. A 5 min stabilization period was followed by increasing the skin temperature to 44 °C by heating the probe and the course of TH was recorded.

The following parameters of microvascular reactivity were evaluated: PORH as the peak flow during hyperemia (PORHmax) expressed in perfusion units (PU), the time necessary to reach this peak (t1) and the mean velocity of the perfusion increase (PORHmax/t1) expressed as a ratio between perfusion increase from basal values and t1 (PU/s). PORH was also expressed as a ratio of peak flow and basal flow as percent of basal value. Similarly, the peak flow during TH (THmax) and the time necessary to reach this peak (t2) were used for calculation of the mean velocity of perfusion increase during TH (THmax/t2) and TH was also expressed as percentage of basal flow. The reproducibility of LDF measurements estimated by repeated measurements in healthy volunteers during 10 consecutive days was characterized by day-to-day variation of the variables below 10%.

The method used for biochemical measurement of IGF-I was IRMA after acid–ethanol extraction.

All the performed studies were in accordance with the Helsinki Declaration of 1975, as revised in 1983; the study was approved by the Faculty of Medicine I ethics committee and the studied subjects gave written informed consent.

**Statistics**

Descriptive data are expressed as means±s.d. and the results as means±s.e.m. The GHD patients were compared with the control group using an independent two-sample t-test. A paired t-test was used to analyze
changes within individuals as a result of GH therapy. Statistical significance was inferred at $P < 0.05$.

**Results**

Thirteen patients were studied before and after 6 months and 12 patients after 12 months of GH replacement therapy; one patient withdrew from the study after 6 months. Reduced pretreatment IGF-I levels normalized during GH treatment from $135 \pm 79 \mu g/l$ (mean $\pm$ S.E.M.) to $269 \pm 67 \mu g/l$ ($P < 0.01$) after 6 months and $272 \pm 78 \mu g/l$ after 12 months ($P < 0.01$). There were no significant changes in BMI or blood pressure values during treatment.

Parameters of forearm microcirculation measured by LDF in GHD patients before GH therapy were significantly reduced in comparison with control subjects and restored on GH treatment (Table 1, Figs 1 and 2).

**Discussion**

This is the first study demonstrating the reduced skin perfusion and impaired microvascular reactivity

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**Table 1** Microcirculation parameters of GHD patients before and during GH replacement and of the controls. Data are means $\pm$ S.E.M.

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<tr>
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<th>GHD patients</th>
<th>Controls</th>
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<tbody>
<tr>
<td></td>
<td>Before GH</td>
<td>6 months’ GH</td>
</tr>
<tr>
<td>$TH_{\text{max}}$ (PU)</td>
<td>68 $\pm$ 6.6$^a$</td>
<td>104 $\pm$ 15.4$^c$</td>
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<tr>
<td>$TH_{\text{max}}/t_2$ (PU/s)</td>
<td>0.84 $\pm$ 0.07$^c$</td>
<td>1.52 $\pm$ 0.32</td>
</tr>
<tr>
<td>$TH$ (%)</td>
<td>1060 $\pm$ 228$^b$</td>
<td>1805 $\pm$ 207$^{b,c}$</td>
</tr>
<tr>
<td>PORH$_{\text{max}}$ (PU)</td>
<td>34 $\pm$ 4.9$^b$</td>
<td>37 $\pm$ 7.0</td>
</tr>
<tr>
<td>PORH$_{\text{max}}/t_1$ (PU/s)</td>
<td>3.36 $\pm$ 0.78</td>
<td>3.19 $\pm$ 0.74</td>
</tr>
<tr>
<td>PORH (%)</td>
<td>414 $\pm$ 63$^c$</td>
<td>514 $\pm$ 53$^b$</td>
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Statistical significance of differences between patients and controls: $^aP < 0.001$, $^bP < 0.01$, $^cP < 0.05$ and between patients before and during treatment: $^dP < 0.001$, $^eP < 0.01$, $^fP < 0.05$. 

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**Figure 1** (a) The comparison of maximal flow during TH ($TH_{\text{max}}$) between controls (C) and GHD patients before (GH0) and during GH therapy (GH6 and GH12) (means $\pm$ S.E.M.). $P < 0.001$ between patients before GH and controls. (b) The comparison of the velocity of flow increase during TH ($TH_{\text{max}}/t_2$) between controls (C) and GHD patients before (GH0) and during GH therapy (GH6 and GH12). $P < 0.05$ between patients before GH and controls.
measured by LDF in GHD patients and its restoration on GH replacement. We are not aware of any other study using this technique in adult GHD patients to compare with. Several studies have proved an impaired endothelium-dependent FMD of the brachial artery in GHD patients while endothelium-independent vasodilatation was similar to that of controls (7, 8, 10). Results of FMD induced by shear stress are known to be dependent on endothelium-derived production of nitric oxide (NO) (11). A systemic NO production is reduced in the GHD state and it normalizes on GH replacement (12). Vasodilatation stimulated by IGF-I can be completely reversed by NO synthase inhibitor NG-monomethyl-L-arginine, indicating that the mediators of GH action on blood flow are IGF-I, NO synthase and NO (13). Similar to the effect of GH replacement on FMD measured on the brachial artery, we suggest a key role for the IGF-I–NO synthase–NO axis in the regulation of microcirculation as well. Mainly the shear stress during PORH test is considered to demonstrate the endothelium-dependent FMD. Not only production of NO but also NO bioavailability can modify the vasomotion. The elevated levels of free radicals in GHD can reduce NO bioavailability and vasoreactivity (8). Reduction of increased oxidative stress and restoration of NO bioavailability during GH treatment (8) is another plausible mechanism improving microcirculation.

However, other factors could also contribute to the changes in microvascular reactivity and blood flow in the GHD state. An increased sympathetic activity described in GHD (14) can enhance vasoconstriction and reduce microvascular perfusion in the GHD state. Reduction of an increased sympathetic activity during GH therapy can positively influence tissue perfusion (14). The other factor influencing the microcirculation is the thyroid hormone status. Skin perfusion measured on LDF is increased in hyperthyroid patients and decreased in hypothyroid patients (15). GH replacement stimulates peripheral conversion of thyroxine (T4) to the active hormone triiodothyronine (T3) in GHD adult patients (16). This shift to higher levels of T3 in tissues might also contribute to the increase in the microcirculation.

Adult GHD is associated with an abnormal body composition by reduced lean body mass, reduced total body water (TBW) and increased fat mass. GH replacement therapy increases TBW, plasma volume (PV) and total blood volume (TBV) (17) and this evokes speculations about possible relationships with changes in microcirculation. Mean changes in these parameters in the 3 month study of Christ et al. (17) were: TBW 2.5±0.53 kg (mean±S.E.M.), PV 350±117 ml and TBV 515±109 ml. Whether these changes in hydration contribute to the changes in microcirculation has not been studied yet. An acute 450 ml blood loss is not associated with the change in cutaneous microcirculation measured by LDF (18); however, the effect of long-term change in TBV and PV is not known.

The main increase in the parameters of tissue microcirculation in our study was seen in the data from the dynamic tests representing the vascular reactivity, while the basal microcirculation data were not significantly changed by GH therapy. The relationship of these parameters in TH and PORH tests is reflected by the relative expression of data as a ratio of peak flow and basal flow as a percent of basal values. Restoration of vascular reactivity by GH replacement we suppose to represent the improvement of the endothelial dysfunction, an early marker of atherosclerosis. This might have a favorable clinical effect on the increased vascular morbidity of GHD patients.

There is an interesting observation of Maison et al. (19), who found an impaired endothelium-dependent vasodilatation using the method of LDF in active acromegalic patients. Acromegalics are known to have an increased cardiovascular morbidity and mortality (20). It seems that both deficiency and excess of GH negatively influence endothelial function and hence contribute to cardiovascular risk factors in these states.

The observed reduced skin microvascular perfusion and reactivity in GHD patients might also contribute to the pathophysiology of the clinical symptoms of
GHD syndrome-impaired thermoregulation, reduced sweating and increased body heat storage (21).

We conclude that the tissue microvascular perfusion and reactivity are reduced in adult GHD syndrome and are restored on GH replacement. The stimulatory effect of GH replacement on the microcirculation is probably mediated by all the above mentioned mechanisms – improved endothelial function with increased NO production and bioavailability, changes in sympathetic tone and increased T₃ to T₄ conversion.

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References


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