A novel germline mutation in the TSH receptor gene causes non-autoimmune autosomal dominant hyperthyroidism

Luisella Alberti 1, Maria Carla Proverbio 3, Sabine Costagliola 4, Giovanna Weber 3, Paolo Beck-Peccoz 2, Giuseppe Chiumello 3 and Luca Persani 1

Institute of Endocrine Sciences, University of Milan, 1Istituto Auxologico Italiano IRCCS and 2Ospedale Maggiore IRCCS, Milan, Italy, 3Department of Pediatrics, University of Milan, Scientific Institute H.S. Raffaele, Milan, Italy and 4Institut de Recherche Interdisciplinaire, University of Brussels, Belgium

(Correspondence should be addressed to Luca Persani, Laboratorio di Ricerche Endocrinologiche, Istituto Auxologico Italiano IRCCS, Via Ariosto 13, 20145 Milano, Italy; Email: persani@auxologico.it)

Abstract

Objective: Clinical and genetic investigations were undertaken in a case of familial hyperthyroidism, with onset of thyrotoxic symptoms varying between childhood/adolescence.

Methods: Automatic sequence analysis was carried out of the TSH receptor (TSHR) gene. Functional studies were undertaken of mutant TSHR in transient expression experiments in COS-7 cells including the evaluation of cAMP accumulation and of protein expression by flow cytometry, as well as the calculation of specific constitutive activity (SCA).

Results: In four affected cases, the age of onset of thyrotoxic manifestations of non-autoimmune origin varied between 5 and 18 years. The disease transmission was typically autosomal dominant. TSHR gene sequence revealed the presence of a germline heterozygous substitution at codon 597 leading to the novel mutation V597F. This residue is located in the 5th transmembrane domain of the receptor protein in a critical region for membrane targeting and signal transduction. Functional studies of the V597F mutant indicate an 11-fold increase in SCA, associated with a reduction in receptor protein expression on the cytoplasmic membrane.

Conclusions: Description was made of a family with non-autoimmune autosomal dominant hyperthyroidism carrying a novel mutation of TSHR leading to the increment in specific constitutive activity. Factors that may influence the clinical expression of TSHR germline mutations are discussed.

European Journal of Endocrinology 145 249–254

Introduction

Autoimmunity (Graves’ disease) is the major cause of thyrotoxicosis in early infancy and childhood (1). Germline gain-of-function mutations of thyrotropin (TSH) receptor (TSHR) have been found to cause non-autoimmune hyperthyroidism with neonatal to juvenile onset (2). The sporadic cases so far reported presented already at birth or in infancy with severe thyrotoxic manifestations and goiter (3–9), whereas a delayed and progressive onset of hyperthyroidism with milder clinical manifestations was generally observed in the cases with inherited mutation of the TSHR gene (10–13). The reasons for such a variable age of onset and degree of disease manifestation may be attributed either to the type of the mutation or to other endogenous or exogenous factors. Typically, the disease is persistent with immediate relapses after withdrawal of anti-thyroid drugs, and sporadic cases are frequently resistant to medical treatment. Therefore, total thyroidectomy is the treatment of choice in these situations.

We report a new family with non-autoimmune autosomal dominant hyperthyroidism due to a novel germline mutation of the thyrotropin receptor gene.

Materials and methods

Case report

The propositus is a female born after normal gestation. Her family originates from the Romagna region in Italy. She was seen at 5 years of age for high stature accompanied in the last months by progressive worsening of symptoms such as hyperactivity, irritability, and palpitations. Physical examination showed mild signs of thyrotoxicosis with a small goiter: her heart rate was 120 beats/min, blood pressure was 130/70 mmHg. Her body weight was 34 kg, and her height was above the 97th centile: 129 cm. She had no sign of sexual maturation and no skin lesions. The biochemical investigations showed TSH: <0.01 mU/l (normal value (n.v.) 0.24–4.0); free thyroxine (FT₄):
45.6 pmol/l (n.v. 9–18); free tri-iodothyronine (FT₃): 15.6 pmol/l (n.v. 4–8) in the absence of anti-thyroid auto-antibodies, including anti-thyroidperoxidase, anti-thyroglobulin, as well as anti-TSH receptor (TRAb). Serum levels of β-chorionic gonadotropin and 17β-estradiol were below the detection limits. Ultrasound showed a moderate and diffuse thyroid enlargement (5.8 ml) with normal ecogenicity. Thyroid scintiscan showed a diffuse 99Tc uptake. Ovary ultrasound was normal, as was skull magnetic resonance imaging. Orbital ultrasound did not show any sign of associated ophthalmopathy. Her bone age was 7 years (2 years in advance of chronological age, but 1.5 years retarded for statural age). Dual-energy X-ray absorptiometry (DXA) showed reduced bone density for age. Serum growth hormone (GH) levels were 0.1 ng/ml in 3 different determinations, but insulin-like growth factor-I (IGF-I) was high (325 ng/ml; n.v. for age: 14–194; n.v. for bone age: 24–220). The family history revealed that her father, her paternal aunt and elder half-sister (the child of her father’s first marriage) underwent total thyroidectomy for non-autoimmune hyperthyroidism and large goiter at 18, 16 and 9 years respectively (Fig. 1).

After informed consent, blood was obtained from the propositus and all the available family members for genetic analyses.

The patient was initially treated with methimazole (MMI) 0.3 mg/kg body weight (bw)/day, but no clinical nor biochemical response was seen after 40 days (TSH <0.01 mU/l; FT₄: 41.4 pmol/l; FT₃: 17.1 pmol/l). The dose of MMI was then augmented to 0.6 mg/kg bw/day. After 75 days, a clinical improvement with normalization of the heart rate (80 beats/min) and circulating free thyroid hormones was seen (TSH <0.01 mU/l; FT₄: 15.8 pmol/l; FT₃: 7.3 pmol/l). This was accompanied by the normalization of circulating IGF-I (157 ng/ml) after 3 months.

**DNA analyses**

Genomic DNA was extracted from peripheral whole blood of the proband and of her father, mother, grandmother, and elder half-sister (see Fig. 1), using a commercial kit (Nucleon BACC2, Amersham Pharmacia Biotech Italia, Cologno Monzese, Italy) according to the manufacturer’s instructions. The fragments containing the whole length of exon 10 of the TSHR gene were amplified by PCR as previously described (14). The purified PCR products were directly sequenced in both forward and reverse directions using the Big Dye Terminator kit and resolved by capillary electrophoresis on an ABI 310 Automated Sequencer (PE Biosystems, Foster City, CA, USA). The sequence was confirmed using two different PCR products.

**Cloning and mutagenesis of TSHR**

cDNA encoding the human TSHR was obtained by RT-PCR from normal thyroid tissue using the following primers, 1 forward: 5'GAGGATGGAGAAATAGCCCCGAG-3' and 10 reverse: 5'GTGTCATGGGATTGGAATGC-3' (the substituted base is in bold and italic). Two full-length TSHR clones carrying the mutations were entirely sequenced to verify possible differences other than the missense mutation itself.

**Cell culture and transient transfection**

COS-7 cells were grown in Dulbecco’s modified Eagle’s medium (DMEM, Gibco BRL, Life Technologies Italia, San Giuliano Milanese, Italy) containing 10% fetal calf serum (FCS) and antibiotics. For transient transfection, cells were seeded at a density of 300 000 cells/3 cm dish and transected with 0.5 µg of specific DNA by the DEAE-Dextran method, followed by a DMSO shock
Cells were then grown in DMEM/10% FCS for 48 h before cAMP determination and flow cytometry experiment.

**cAMP determination**

The culture medium was removed and replaced with Krebs-Ringer-HEPES (KRH) buffer, 0.5% BSA, for 30 min at 37 °C. Then cells were incubated at 37 °C for 60 min in fresh KRH buffer, 0.5% BSA, supplemented with 0.5 mmol/l isobutylmethylxanthine (IBMX) and the indicated concentrations of bovine (b) TSH (Sigma-Aldrich srl, Milan, Italy). The medium was removed and the reaction stopped by adding 0.1 mol/l HCl. Samples were dried and cAMP was measured using a commercial assay kit (RIANEN, NEN Life Science Products Inc., Boston, MA, USA). The results were expressed in pmoles/dish.

**Flow cytometry**

Flow cytometry experiments were performed as previously described (17). For permeabilization, cells were incubated with PBS-paraformaldehyde 2% in ice for 10 min, washed once with PBS-BSA 0.1% and treated with 2% saponine at room temperature for 30 min. Permeabilized and non-permeabilized cells were labeled with 3G4 monoclonal antibody recognizing a linear epitope, and with BA8 antibody recognizing a conformational epitope, both within the ligand-binding domain of TSHR (18).

**Computation of specific constitutive activity**

The fold cAMP constitutive activity above mock-transfected COS-7 cells was calculated for each receptor construct, and then normalized for their own level of expression, as previously described (19). Specific constitutive activity (SCA) of the mutant was then expressed as fold-increase over the wild-type (19).

**Statistical analysis**

Data were analyzed by Student’s t-test and analysis of variance (ANOVA), as appropriate. P values <0.05 were considered statistically significant. Results are expressed as means±standard deviation (s.d.).

**Results**

**DNA analysis**

Automatic sequencing of DNA obtained from peripheral whole blood of the propositus revealed a heterozygous G to T transversion at codon 597 in the TSHR gene leading to the missense substitution V597F (Fig. 2). This codon is located in the fifth transmembrane domain of the receptor. Both the father and the half-sister of the propositus were heterozygote for the same substitution, whereas the TSHR sequence was of the wild-type (WT) in the mother and the paternal grandmother (Fig. 1).

**Functional studies of mutant receptor**

Flow cytometry experiments, one of which is shown in Fig. 3, indicate that the synthesis of TSHR-WT and TSHR-V597F constructs are not significantly different as demonstrated in permeabilized cells, but the receptor carrying the activating mutation displays a reduced expression level on the cell surface. The entity of this reduction is 60% of that observed with normal receptor using the 3G4 antibody (Fig. 3A). Expression of the V597F mutant was even lower (25% vs wild-type) when the BA8 monoclonal antibody directed against a conformational epitope was used (Fig. 3B).

In order to evaluate the activity of the V597F mutant receptor, we measured basal and TSH-stimulated cAMP accumulation in transiently transfected COS-7 cells in several experiments (n = 7). As shown in Fig. 4, expression of the V597F mutant receptor always resulted in a marked increase in basal cAMP production when compared with the wild-type receptor or mock-transfected cells. The V597F mutant maintained the ability to respond to TSH stimulation, although the increment in cAMP production at 10 U/l bTSH is 2.4-fold lower than that of wild-type (Fig. 4). Maximal cAMP stimulation was reached at 10 U/l bTSH in both cases. On this basis, calculation of SCA (19) showed an 11.4±0.4- or a 4.6±0.2-fold increase over the wild-type receptor in intact and permeabilized cells respectively.

**Discussion**

We report a new case of non-autoimmune autosomal dominant hyperthyroidism. In this family, thyrotoxic manifestations and goiter had a childhood to juvenile onset, ranging from 5–18 years of age in the four
Figure 3 Representative flow cytometry experiment, using 3G4 (A) and BA8 (B) monoclonal antibodies (Ab), for the analysis of wild-type (wt) and V597F TSHR protein expression in intact and permeabilized cells. Results with mock transfected cells (pTarget) are also shown. These data demonstrate a reduced expression of V597F mutant TSHR protein on the membrane of intact COS-7 cells.
showed a dramatic increase in ligand-independent receptor activity associated with the severe impairment of membrane targeting, leading to a diminished response to TSH stimulation. The increase in specific constitutive activity of the V597L mutant was markedly higher than that observed in the case of V597F (28.9- vs 11.4-fold over wild-type in intact cells). As in other G protein-coupled receptors (20), the intramolecular interactions between hydrophobic residues in the fifth and sixth transmembrane domains would contribute to the structural inhibitory constraint keeping the TSHR in its basal state (21). V597 residue is one of the hydrophobic residues involved in this structural inhibitory constraint and mutations at this residue are expected to alter constitutive receptor activity. However, the differences in specific constitutive activity between V597L and V597F are difficult to explain. In fact, both leucine and phenylalanine have non-polar side chains like valine, and the bulkier side chain of phenylalanine would be expected to produce a more profound disruption of the structural inhibitory constraint.

Although caution is advised in interpreting data obtained in a heterologous system, the reported differences in the in vitro functional characteristics of the two mutants affecting the same V597 residue are mirrored by the clinical phenotype recorded in the affected patients. In fact, the V597L mutation was found in one sporadic case with infantile onset (at 9 months of age) of severe thyrotoxic features (8), whereas V597F is associated with a childhood to juvenile onset of thyrotoxic features. Germline mutations causing dramatic increases in TSHR constitutive activity causing severe thyrotoxicosis since early infancy (or even congenital) (3–9) may be associated with intellectual impairment and craniosynostosis in later years (22), and familial vertical transmission may become unlikely. Alternatively, this would not be prevented in the case of germline mutations with less dramatic effects on TSHR function (10–13). Consistently, the inherited TSHR mutations generally present later in life (typical childhood or juvenile onset) with mild to moderate thyrotoxic manifestations. Hyperstaturism was described in our proband and bone was severely affected, indicating that, despite mild and unrecognized clinical symptoms, tissue thyrotoxicosis had been long-standing. Increased growth velocity was, at least in part, mediated by the thyroid hormone-dependent induction of IGF-I secretion from the liver, perhaps through the stimulation of hepatic growth hormone receptor gene expression, as already seen in the chicken (23). Consistently, increased IGF-I secretion reverted to normal during MMI treatment.

Increased medical awareness and improvement in diagnostic testing (such as availability of ultrasensitive TSH assay) (24) throughout the last decades could have allowed the earlier diagnosis in the proband. Nevertheless, the clinical expression of the same alteration of the TSHR gene in the different members of one family may also be influenced by several

![Graph](image-url)
environmental (such as modifications in iodine intake) and/or endogenous (such as sex) factors. During the 1980s, when the father and aunt of our proband both underwent thyroidectomy and before the prophylaxis with iodized salt was well-established, the mean urinary excretion of iodine was as low as 58.1 μg/l in the Romagna region (25). Accordingly, the diagnosis of thyrotoxicosis was reached at 15 and 18 years in the older members of the family and at 5 years in the proband. Differently from what is usually seen in sporadic cases, hyperthyroidism could be controlled by high doses of methimazole. However, progressive thyroid growth is expected through the years and strict ultrasound monitoring is advocated during treatment in order to program thyroid surgery before the formation of a large goiter, as already seen in her affected relatives.

Acknowledgements

This work was partially supported by Funds of the Ricerca Corrente of Istituto Auxologico Italiano IRCCS (to L A and L P).

References


16 Parma J, Van Sande J, Swillens S, Tonacchera M, Dumont J & Vassart G. Somatic mutations causing constitutive activity of the thyrotropin receptor are the major cause of hyperfunctioning thyroid adenomas: identification of additional mutations activating both the cyclic adenosine 3',5'-monophosphate and inositol phosphate-Ca2+ cascades. Molecular Endocrinology 1995 9 725–733.


Received 23 January 2001
Accepted 30 May 2001

www.eje.org