Oestradiol and progesterone change β<sub>3</sub>-adrenergic receptor affinity and density in brown adipocytes

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Abstract

Objective: To check if the oestradiol- and progesterone-driven reduction in noradrenaline responsiveness of brown adipocytes is due to a reduction in either the density or the affinity of β<sub>3</sub>-adrenoceptors (β<sub>3</sub>-AR). β<sub>1</sub>/β<sub>2</sub>-AR were also studied.

Design: Four groups of animals were considered. (i) control rats at thermoneutrality, (ii) cold-acclimated rats, to determine β-AR under continuous sympathetic stimulation, which is known to decrease noradrenaline responsiveness, (iii) oestradiol- and (iv) progesterone-treated cold-acclimated rats to determine hormonal effects on β-AR populations in thermogenically active brown adipocytes.

Methods: Oestradiol and progesterone were chronically elevated by means of s.c. Silastic implants. Densities and affinities of β-AR populations were determined by binding studies using [3H]CGP-12177 as radioligand.

Results: Two populations of low and high binding affinities (K<sub>d</sub> 1.6 and 27.3 nmol/l) corresponding to β<sub>3</sub>- and β<sub>1</sub>/β<sub>2</sub>-AR respectively were found at thermoneutrality. β<sub>3</sub>-AR density was higher than that of β<sub>1</sub>/β<sub>2</sub>-AR (B<sub>max</sub> 419 and 143 fmol/mg protein respectively). Cold-acclimated rats showed a reduction of β<sub>3</sub>-AR binding capacity (B<sub>max</sub> 308 fmol/mg protein). Oestradiol and progesterone reduced the density of β<sub>3</sub>-AR to 167 and 185 fmol/mg protein, while increasing their affinity for [3H]CGP-12177 (K<sub>d</sub> 9.5 and 4.0 nmol/l vs 16 nmol/l in cold-acclimated untreated rats). The density of β<sub>1</sub>/β<sub>2</sub>-AR was also reduced after oestradiol treatment (B<sub>max</sub> 51 fmol/mg protein).

Conclusions: Both oestradiol and progesterone reduce the density of β<sub>3</sub>-AR in brown adipose tissue (BAT) while increasing their affinity for [3H]CGP-12177. Oestradiol also reduces the density of β<sub>1</sub>/β<sub>2</sub>-AR whereas cold-acclimation reduces the density of β<sub>3</sub>-AR.
Oestradiol and progesterone were administered by implanting Silastic capsules (1.5 and 3.2 mm internal and external diameter respectively) filled with the corresponding hormone under ether anaesthesia (17β-oestradiol, one capsule 0.5 cm long containing about 5 mg; progesterone, two capsules, 4 cm long, each containing about 25 mg).

Animals were killed by decapitation. Interscapular BAT (IBAT) was removed and cleaned of adherent tissues in 250 mmol/l sucrose, 10 mmol/l Tris–HCl and 1 mmol/l EDTA adjusted to pH 7.4 (STE) at 4 °C. Purified plasma membranes were obtained as described by Muzzin et al. (7). Briefly, about 1.25 g IBAT (pooled from four to ten rats) were minced with scissors into pieces of about 1 mm³. After homogenizing (Potter-Elvehjem, Cole Palmer International, USA) the pooled tissue at 1700 r.p.m. for 1 min, nuclei and cellular debris were removed by a low speed centrifugation (1075 g for 10 min). The supernatant was centrifuged at 34 500 g for 35 min and the pellet resuspended in STE, homogenized and transferred to a centrifuge tube filled with 18.5% metrizamide. After centrifugation at 156 000 g for 60 min in a floating rotor, membranes remained over the metrizamide layer. They were collected and either used immediately or stored frozen at −80 °C.

For radioligand binding studies, membranes (30 µg protein) were incubated at 37 °C for 30 min in 50 mmol/l Tris–HCl and 10 mmol/l MgCl₂ at pH 7.4 containing GTP 50 µmol/l with increasing concentrations of [³H]CGP-12177 (44 Ci/mmol, Amersham Pharmacia, UK) in the presence or absence of (−)-propranolol. Once incubated, membranes were collected using a Brandel M-24 harvester (Brandel, MD, USA) equipped with Whatman GF/C paper. Radioactivity was measured after incubating every sample at 60 °C for 20 min in 10 ml scintillation mixture containing 500 µl Soluene-350 (Packard, Groningen, The Netherlands). Specific binding was defined as the difference between the total binding obtained in the absence of competing ligand and the non-specific binding obtained in the presence of (−)-propranolol.

Three different plasma membrane pools were obtained from each experimental group. Each pool (four to ten rats) was only used to make one binding study. Each binding study was made up of a series of 9–12 tubes in duplicate. The quantitative parameters K_d and B_max were determined by Scatchard analysis. One-way ANOVA was used to determine differences between groups, except for β₁ K_d comparisons, where Kruskal–Wallis analyses were made due to the absence of a Gaussian distribution. The Newman–Keuls test was used as post hoc analysis. P < 0.05 was considered statistically significant.

Results

Muzzin et al. (8) showed that [³H]CGP-12177 binds two populations of β-AR on rat IBAT plasma membranes, a high affinity binding site (0.64 ± 0.017 nmol/l) made up of β₁/β₂-AR subtypes, representing 30% of the total binding (330 ± 44 fmol/mg protein), and a low affinity binding site (31 ± 9 nmol/l) made up of β₃-AR, representing 70% of the total binding (770 ± 85 fmol/mg protein). The identity of the low affinity binding site as β₃-AR was confirmed in the same work by using CHO-K1 β₃-transfected cells, which also gave a similar low affinity binding site (44 ± 4 nmol/l) for [³H]CGP-12177.

Using [³H]CGP-12177 as AR radioligand as described by Muzzin et al. (8) we also found two
populations of binding sites in control rats (Fig. 1A), a high affinity one having a $K_d$ value of $1.6 \pm 0.6$ nmol/l and a $B_{\text{max}}$ value of $143.3 \pm 37.9$ fmol/mg protein, and a low affinity one having a $K_d$ value of $27.3 \pm 6.9$ nmol/l and a $B_{\text{max}}$ value of $419.4 \pm 11.2$ fmol/mg protein (Table 1, first row). Since these values were in excellent agreement with those found by Muzzin et al., we considered the high and the low binding affinity sites in our study as the $\beta_1/\beta_2$-AR subtypes and the $\beta_3$-AR subtype respectively.

Both $\beta_1/\beta_2$-AR and $\beta_3$-AR were also found in BAT membranes after cold-acclimation (Fig. 1B). However, IBAT membranes of cold-acclimated rats had a lower density of $\beta_3$-AR than those of control rats ($38.6 \pm 38.9$ vs $143.3 \pm 7.9$ fmol/mg protein respectively) (Table 1, second row). They also showed a reduction in the number of $\beta_1/\beta_2$-AR ($95.0 \pm 1.0$ vs $143.3 \pm 37.9$ fmol/mg protein respectively) but the difference did not attain statistical significance (Table 1, second row). No effects were detected in binding affinity.

Figure 2 shows that oestradiol and progesterone modified both the binding affinity and the capacity of $\beta_3$-AR. Thus the $B_{\text{max}}$ changed from a value of $308.6 \pm 38.9$ fmol/mg protein in untreated rats to $167.7 \pm 15.4$ and $185.6 \pm 20.1$ fmol/mg protein after oestradiol and progesterone treatment respectively (Table 1). Simultaneously, the $K_d$ changed from a value of $16.0 \pm 1.5$ nmol/l in untreated rats to $9.5 \pm 2.0$ and $4.0 \pm 0.9$ nmol/l after oestradiol or progesterone treatment respectively (Table 1).

Only oestradiol produced a change in the $\beta_1/\beta_2$-AR, which reduced their number from a $B_{\text{max}}$ of $95.0 \pm 1.0$ fmol/mg protein in untreated rats to $51.0 \pm 14.1$ in oestradiol-treated animals, leaving unchanged their affinity for $[^3\text{H}]$CGP-12177. Progesterone had no effect on either binding affinity or capacity of $\beta_1/\beta_2$-AR (Table 1).

### Discussion

Muzzin et al. (8) showed that the low affinity $\beta_3$-AR can be quantified in plasma membranes from rat brown adipocytes and from CHO $\beta_3$-transfected cells with the radioligand $[^3\text{H}]$CGP-12177. The high affinity $\beta_1/\beta_2$-AR are also quantified. Using the same radioligand, we also found two different receptor populations with similar relative proportions to those reported by Muzzin et al. (8). Therefore, for our study we defined the low affinity binding site as $\beta_3$-AR and the high affinity site as $\beta_1/\beta_2$-AR. The receptor density for both $\beta_3$- and $\beta_1/\beta_2$-AR was slightly lower in this study than in the work of Muzzin et al. (8); these differences are most likely due to the strain of rat used (Wistar in our experiment but Sprague–Dawley in theirs) or sex (females in our experiments but males in theirs).

After rats had been acclimated to the cold for 15 days, we found a significant reduction in the density of $\beta_3$-AR, from $308.6 \pm 38.9$ fmol/mg protein in controls to $308.6 \pm 38.9$ fmol/mg protein in the cold-acclimated animals (Table 1). This result indicated that chronic noradrenaline stimulation diminishes the density of $\beta_3$-AR, which agrees not only with Revelli et al. (9), who found that the actual number of $\beta_3$-AR was diminished in rat BAT after 72 h of cold-exposure, but also explains the reduced response to both noradrenaline and to other adrenergic agonists of brown adipocytes isolated from cold-acclimated hamsters (10, 11) and rats (12). Therefore, our results indicate that chronic sympathetic stimulation desensitizes $\beta_3$-AR by reducing $\beta_3$-AR density, and this desensitization would explain the reduced responsiveness to noradrenaline in BAT of cold-acclimated rodents.

We found no significant change in $\beta_1/\beta_2$-AR density in BAT membranes after cold-acclimation of rats (Table 1). Therefore, our results suggest that chronic noradrenaline stimulation does not change the $\beta_1/\beta_2$-AR binding capacity of BAT membranes. It could be argued, however, that whereas $\beta_3$-AR are only present in mature brown adipocytes, $\beta_1/\beta_2$-AR are also present in preadipocytes (13) and that preadipocyte number might have increased during cold-acclimation, thus masking a reduction in binding density. However, Bukowiecki et al. (14) showed that in the rat at room temperature the ratio of mature adipocytes to preadipocytes is 40:2, and, most importantly, those proportions are unchanged after 15 days in the cold. It therefore seems more logical to conclude that chronic noradrenaline stimulation, as present in

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Table 1 Density and affinity of $\beta_3$- and $\beta_1/\beta_2$-AR in IBAT plasma membranes from rats at thermoneutrality and from cold-acclimated rats treated with either oestradiol or progesterone. Values are means ± s.e. of three different binding studies.

<table>
<thead>
<tr>
<th>Temperature</th>
<th>$\beta_3$-AR</th>
<th>$\beta_1/\beta_2$-AR</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$B_{\text{max}}$ (fmol/mg protein)</td>
<td>$K_d$ (nmol/l)</td>
</tr>
<tr>
<td>28 °C</td>
<td>419.4±11.2</td>
<td>27.3±6.9</td>
</tr>
<tr>
<td>6 °C</td>
<td>308.6±38.9*</td>
<td>16.0±1.5</td>
</tr>
<tr>
<td>6 °C+C+oestradiol</td>
<td>167.7±15.4**</td>
<td>9.5±2.0**</td>
</tr>
<tr>
<td>6 °C+C+progesterone</td>
<td>185.6±20.1**</td>
<td>4.0±0.9**</td>
</tr>
</tbody>
</table>

* $P < 0.05$ when compared with controls at 28 °C.
** $P < 0.05$ when compared with untreated cold-acclimated rats.
cold-acclimated rats, does not alter density or affinity of \( \beta_1/\beta_2 \)-AR.

To study the effect of oestradiol and progesterone on \( \beta \)-AR we chose cold-acclimated rats as the experimental model for three reasons. Firstly, oestradiol and progesterone inhibition of BAT thermogenesis was found in cold-acclimated rats (1, 2). Secondly, the direct effect of oestradiol and progesterone on brown adipocytes reducing their responsiveness to noradrenaline was also found in adipocytes isolated from rats at room temperature; this is below thermoneutrality (5, 6). Thirdly, cold-acclimation itself is known to lower noradrenaline sensitivity (10, 11), an effect this study has revealed to be mediated by a reduction in the density of \( \beta_3 \)-AR. We found that oestradiol and progesterone brought about a further reduction in \( \beta_3 \)-AR density, that went from 308 ± 38.9 fmol/mg protein in untreated rats down to 167 ± 15.4 fmol/mg protein in those receiving oestradiol and to 185 ± 20.1 fmol/mg protein in those receiving progesterone (Table 1). However, oestradiol and progesterone not only decreased \( \beta_3 \)-AR density but also increased the affinity of the \( \beta_3 \)-AR for \([3H]CGP-12177\) at the same time, as revealed by the diminished \( K_d \) found in treated animals (16.0 ± 1.5 nmol/l in untreated rats down to 9.5 ± 2.0 nmol/l in oestradiol-treated rats and to 4.0 ± 0.9 nmol/l in progesterone-treated animals). Therefore, female sex steroids had a dual effect on \( \beta_3 \)-AR; on the one hand, they diminished their density and, on the other hand, they increased their affinity for \([3H]CGP-12177\). It is tempting to speculate that this dual effect could be related to the dual mechanism of action of steroid hormones, and thus, besides the well-known genomic effects controlling protein synthesis, there are non-genomic actions of steroids that include allosteric interactions of the hormones with neurotransmitter receptors, cation exchangers, ion channels and G proteins (15–17).

According to this dual model, the reduced density of \( \beta_3 \)-AR that we found after either oestradiol or progesterone treatment could probably be the result of a genomic action, since cytoplasmic oestradiol receptors have been identified in BAT (18). However, it seems plausible that the increased affinity of \( \beta_3 \)-AR for CGP-12177 in treated animals could be due to a direct interaction of the steroids with the \( \beta_3 \)-AR. In our system both effects, despite being opposite, do not compensate for each other, since brown adipocytes isolated from rats receiving the steroids showed a

**Figure 2** Effect of oestradiol and progesterone on the specific binding of \([3H]CGP-12177\) to IBAT plasma membranes. Scatchard analysis of the data is represented in the corresponding insert. Each point represents the mean of three different binding studies. Standard errors have been omitted for clarity. (A) Cold-acclimated rats; (B) cold-acclimated oestradiol-treated rats; (C) cold-acclimated progesterone-treated rats.
reduced response to noradrenaline (5, 6), implying that the effect of reducing density predominates.

Oestradiol but not progesterone also reduced the density of β3/β2-AR from 95.0 ± 1.0 down to 51.0 ± 14.1 fmol/mg protein leaving unchanged their affinity for [3H]CGP-12177 (Table 1). Again, a genomic mechanism is suggested to explain this effect.

In summary, our study reveals that β3-AR density is reduced by chronic noradrenaline stimulation, and by oestradiol and progesterone. Both steroids also increase β3-AR affinity for [3H]CGP-12177. However, only oestradiol reduced β3/β2-AR density leaving the binding affinity unchanged.

Acknowledgements

Thanks are given to Professor Giacobino for receiving Aurelio Malo in his laboratory and for his suggestions when preparing the manuscript. Thanks are also given to Dr Muzzin for his expert advice and to Françoise Kühne for technical assistance. The collaboration of Dr Margarita Fernández was, as always, greatly appreciated. This work was supported by Grants PM98-0090 from Dirección General de Enseñanza Superior e Investigación Científica (DGESIC) and 08.6/0007/1998 from Comunidad Autonoma de Madrid (CAM) to MP.

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Received 16 June 2000
Accepted 15 March 2001

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