CLINICAL STUDY

Hyperleptinemia in women with Cushing's disease is driven by high-amplitude pulsatile, but orderly and eurhythmic, leptin secretion

Ronald Groote Veldman, Marijke Frölich, Steve M Pincus, Johannes D Veldhuis and Ferdinand Roelfsema

Abstract

The episodicity of 24 h leptin release was studied in seven women (mean age 39 years, range 22–56 years) with pituitary-dependent hypercortisolism and in seven age- and body mass index (BMI)-matched female controls. Pulsatile leptin release was quantified by model-free CLUSTER analysis and deconvolution, the orderliness of leptin patterns by the approximate entropy statistic (ApEn), and diurnal regulation in Cushing’s disease remains unknown. Such information would probably offer

Introduction

Leptin, the protein product of the ob gene, is produced and secreted primarily by adipose tissue and in smaller amounts by the placenta and central nervous system (1). This metabolic protein binds to specific receptors belonging to the cytokine receptor family (2), which are expressed in many tissues, including that of brain and hypothalamus, thereby conveying feedback information on the quantity of energy stored. Plasma leptin levels are positively correlated with the body mass index (BMI), the percentage of body fat (3), and adipocyte cell size (4). Leptin secretion is higher in premenopausal and postmenopausal females than in comparably aged males (5), at any given level of adiposity (6). This gender difference emerges after the onset of puberty (7). Accordingly, it could be hypothesized that gonadal steroids are involved in the regulation of leptin production. In addition, in vitro and in vivo studies in experimental animals and humans have demonstrated that acute administration of glucocorticoids stimulates leptin gene transcription and increases plasma leptin concentrations (8–11). In Cushing’s syndrome, mean plasma leptin levels, collected hourly over 24 h, were elevated twofold compared with values in BMI-matched controls (12), and ‘fasting leptin’ was increased fourfold compared with lean controls (13). However, the nature of leptin’s episodic release or diurnal regulation in Cushing’s disease remains unknown. Such information would probably offer
further insights into the putative relationship between sustained endogenous hypercortisolemia and altered control of leptin production in the adult with Cushing’s syndrome.

Subjects and methods

Subjects

Seven women with pituitary-dependent Cushing’s disease (mean age, 39 years; range, 22–56 years) and seven age-, BMI- and gender-matched healthy controls (mean age, 44 years; range, 32–58 years) were studied. The diagnosis was established by elevated 24 h urinary excretion of free cortisol, subnormal suppression of urinary cortisol excretion during a low-dose dexamethasone test, suppression of plasma cortisol concentration by 190 nmol/l or more during a 7 h i.v. infusion of dexamethasone at a dose of 1 mg/h (14), positive immunostaining of the adenoma, and cortisol-dependency for at least three months post-operatively. The mean urinary excretion of free cortisol was 864 nmol/24 h (range, 241–2040, normal range, 40–200). All patients and control subjects were clinically and biochemically euthyroid and were not using any medication. Premenopausal controls were studied in the early follicular phase of the menstrual cycle.

Methods

Patients and control subjects were admitted to the hospital on the day of the study. An indwelling i.v. cannula was inserted in a forearm vein 60 min before the start of sampling, and blood samples were withdrawn at 10 min intervals and for the next 24 h, starting at 0900 h. The subjects were free to ambulate, but not to sleep, during the day. Meals were served at 0800, 1230 and 1730 h. Lights were turned off starting at 0900 h. The subjects were free to ambulate, draw at 10 min intervals and for the next 24 h, the start of sampling, and blood samples were with-
cannula was inserted in a forearm vein 60 min before

Assays

Leptin was measured by radioimmunoassay (RIA) (Linco Research, St Charles, MO, USA), with a detection limit of 0.5 μg/l and an intra-assay coefficient of variation of 3.4–8.3% over the leptin concentration range of 4.9–25.6 μg/l; the corresponding interassay coefficient of variation was 3.0–6.2%. Cortisol was assayed by RIA (Sorin Biomedica, Milan, Italy) with a detection limit of sensitivity of 25 nmol/l. The intra- and interassay precision varied from 2 to 4%. Growth hormone (GH) was measured by time-resolved fluoro-immunoassay (Wallac, Turku, Finland) specific for the 22 kDa GH, which was used as the standard (Genotropin; Pharmacia Upjohn, Uppsala, Sweden) as calibrated against the World Health Organization First International Reference Preparation, 80/505 (to convert μg/l to mU/l, multiply by 2.6). The limit of detection was 0.0115 mU/l. The intra-assay coefficient of variation varied from 1.6 to 8.4% in the assay range from 0.26 to 47 mU/l and the interassay coefficient of variation was 2.0–9.9% in the same range. Insulin was assayed by RIA with a detection limit of 3.6 mU/l. The interassay coefficient of variation was 3.8–8.0% over the concentration range of 12.5–94.5 mU/l. All samples from one individual were run in the same batch.

Analytical techniques

Analysis of leptin secretion. Leptin secretion was evaluated using a pulse detection algorithm. The statistical significance of the difference in the mean leptin concentration between groups was tested using the Student’s t-test (22). Statistical analysis was performed using SPSS for Windows (Release 8.0; SPSS, Chicago, IL, USA). Results are expressed as the means ± S.E.M. The Student’s paired, two-tailed t-test was used to compare leptin concentrations between groups. Differences were considered significant for P < 0.05. Linear regression
analysis was performed to analyse the relationships between leptin and insulin or cortisol production. Data were transformed logarithmically for comparisons of derived values.

Results

Figure 1 shows the individual 24 h plasma leptin concentration profiles of two patients and their respective controls. The BMI was 26.4 ± 1.3 kg/m² in patients and 24.5 ± 1.6 kg/m² in controls ($P = 0.167$). Mean and integrated plasma leptin concentrations were more than twofold higher in patients than in controls (see Table 1). This increase was accounted for by an elevation in both the interpeak nadir concentration and incremental peak height (see Fig. 2).

The mean calculated leptin secretion rate was $0.772 ± 0.135 \mu g/l$ per min in patients and $0.333 ± 0.052 \mu g/l$ per min in controls ($P = 0.004$). Nadir

![Graphs showing individual 24 h plasma leptin concentration profiles of patients and controls.](image)

Table 1 Cluster analysis of leptin secretion in patients with Cushing's disease and in controls. Values are expressed as means ± S.E.M. Blood samples were withdrawn at 10 min intervals for 24 h in seven women with Cushing's disease and seven control women matched for age and body mass index. Significance was determined using the paired two-tailed Student's t-test.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Patients</th>
<th>Controls</th>
<th>$P$ value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Integrated concentration ($\mu g/l/24$ h)</td>
<td>45 700 ± 7900</td>
<td>19 700 ± 3100</td>
<td>0.007</td>
</tr>
<tr>
<td>No. of peaks</td>
<td>7.1 ± 1.5</td>
<td>6.0 ± 0.5</td>
<td>NS*</td>
</tr>
<tr>
<td>Peak width (min)</td>
<td>131 ± 35</td>
<td>154 ± 27</td>
<td>NS*</td>
</tr>
<tr>
<td>Peak area ($\mu g/l/min$)</td>
<td>658 ± 106</td>
<td>392 ± 61</td>
<td>NS*</td>
</tr>
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</table>

* NS, not significant.
secretion rates were 0.682 ± 0.119 μg/l per min in patients and 0.292 ± 0.045 μg/l per min in controls (P = 0.004). Table 2 gives the 24 h secretion rates for leptin, cortisol and GH, and shows significantly higher secretion rates for leptin and cortisol, but not for GH, in patients compared with matched controls. The statistical distributions of the 10 min sample secretion rates were similar in patients and controls. The skewness was 0.55 ± 0.12 in patients vs 0.52 ± 0.07 in controls (P = 0.885), with similar kurtosis estimates of, respectively, 0.30 ± 0.32 vs 0.11 ± 0.16 in patients and controls (P = 0.64). Thus, mean and nadir leptin secretion rates were elevated in Cushing’s disease, without any distortion of their relationships. Cosinor analysis disclosed significant differences in the 24 h rhythmic mean plasma leptin concentration (patients, 31.8 ± 5.5 μg/l; controls, 13.7 ± 2.2 μg/l; P = 0.007). The fractional amplitude of the daily leptin rhythm was similar in the two groups (patients, 0.17 ± 0.03; controls, 0.23 ± 0.04; P = 0.331). In addition, patients and controls exhibited comparable acrophases (patients, 2314 h ± 76 min; controls, 0058 h ± 18 min; P = 0.181).

The regularity of leptin secretion, as measured by ApEn (1.20%), did not differ between patients and controls: 1.67 ± 0.03 vs 1.61 ± 0.05 respectively (P = 0.393). Synchrony quantification by Cross-ApEn of the cortisol–leptin time series revealed respective values of 2.02 ± 0.04 in patients and 1.88 ± 0.09 in controls (P = 0.223). In contrast, the ApEn (1.20%) for cortisol was elevated at 1.53 ± 0.09 in patients vs 0.93 ± 0.07 in controls (P < 0.0005). This contrast denotes a markedly disorderly cortisol release in patients.

Table 2 Daily secretion rates for leptin, GH and cortisol, and fasting plasma insulin concentrations in female patients with Cushing’s disease and in matched controls. Values are the means ± S.E.M. and are expressed as mass units per liter distribution volume. Blood samples were withdrawn at 10 min intervals for 24 h in seven women with Cushing’s disease and seven control women, matched for age and body mass index. The mean plasma insulin concentration was the mean of six early morning fasting samples. The significance was determined using the paired two-tailed Student’s t-Test.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Patients</th>
<th>Controls</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leptin secretion (μg/l)</td>
<td>1112 ± 194</td>
<td>480 ± 75</td>
<td>0.007</td>
</tr>
<tr>
<td>GH (mU/l)</td>
<td>515 ± 91</td>
<td>568 ± 108</td>
<td>0.747</td>
</tr>
<tr>
<td>Cortisol (μmol/l)</td>
<td>116 ± 12</td>
<td>51 ± 9</td>
<td>0.001</td>
</tr>
<tr>
<td>Insulin (mU/l)</td>
<td>26.9 ± 5.7</td>
<td>13.6 ± 1.6</td>
<td>0.035</td>
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Figure 2 Comparisons of mean 24 h plasma leptin concentrations (a), maximal leptin peak heights (b), interpeak nadir values (c) and incremental leptin peak elevations (d) in seven women with Cushing’s disease and seven BMI-matched controls. P values were determined as in Table 1.
Fasting plasma insulin concentrations (mean of six consecutive early morning samples) were 50% lower in controls than in patients (13.6 ± 1.6 mU/l vs 26.9 ± 5.7 mU/l respectively; \( P = 0.035 \)).

In patients, no significant correlations were found between the 24 h cortisol-production and leptin-pulse parameters, leptin secretion or leptin ApEn. Pooling of data from patients and controls revealed significant correlations between the 24 h leptin secretion rate and fasting plasma insulin concentrations \( (R^2 = 0.34, \ P = 0.028) \) and 24 h cortisol production \( (R^2 = 0.33, \ P = 0.033) \), but not 24 h GH production \( (R^2 = 0.074, \ P = 0.348) \).

**Discussion**

Women with pituitary-dependent Cushing’s disease exhibit a more than twofold elevation in the daily rate of leptin secretion compared with age-, gender- and BMI-matched controls, as assessed here by two independent analytical techniques. Discrete peak-detection analysis demonstrated proportionate increases in the interpeak nadir serum leptin concentration and incremental leptin peak amplitude. The number of plasma leptin concentration peaks per 24 h did not differ between groups, thus excluding an increase in pulse frequency as the driving mechanism for relative hyperleptinemia. The present analyses corroborate and extend the results reported by Leal-Cerro et al., who utilized hourly blood sampling (12). The latter study could not include detailed pulse and entropy evaluation, but also identified a nearly twofold elevation of mean plasma leptin in Cushing’s syndrome compared with eucortisolemia. As weight gain is a frequent clinical sign of Cushing’s disease (23), the increased plasma leptin concentration might be interpreted as a physiological adaptation that signals appetite restraint despite hypercortisolism. The observation that circulating leptin concentrations did not fall 10 days following correction of the hypercortisolemic state could further suggest that factors other than ongoing cortisol hypersecretion also play a significant role in leptin overproduction in established Cushing’s syndrome (13).

Systemic leptin concentrations in healthy individuals typically correlate positively with BMI, percentage body fat and total body fat (6). Regional fat distribution has a predominant impact on circulating leptin levels, since subcutaneous (but not visceral) fat mass measured by computed tomography or magnetic resonance imaging predicts leptin concentrations (24, 25). Van Harmelen and co-workers demonstrated, by using direct biopsy measurements, that leptin secretion from subcutaneous fat was 2–3 times higher than that from omental fat, although both fat compartments contributed significantly to the circulating leptin level (26). Patients with Cushing’s disease have reduced lean body mass as well as increased fat mass compared with normal subjects. However, a recent detailed comparative study, by Wajchenberg et al., of patients with Cushing’s disease and BMI-matched healthy controls showed that total body fat is no higher, but that the proportion of visceral fat to subcutaneous truncal fat is increased by 60% (27). This topography corresponds to an apparent shift of about 1.6 kg from subcutaneous to visceral abdominal fat stores. Although we do not have body compositional data for our patients, assuming analogous distribution, we reason that preferential omental fat accumulation in Cushing’s syndrome is not an evident explanation for the hyperleptinemia observed in this hypercortisolemic disease.

In contrast to the positive association between fat stores and leptin production, lean body mass is a negative statistical determinant of plasma leptin levels (5, 28, 29). In this regard, Wajchenberg et al. quantified slightly lower absolute (but not fractional) lean body mass in Cushing’s disease compared with obese controls. Accordingly, the role of altered lean body mass in hyperleptinemia associated with Cushing’s syndrome is not yet clear.

GH secretion is negatively correlated with total body fat and, in particular, central obesity, as well as with leptin secretion. GH, but not insulin-like growth factor, directly inhibits leptin gene expression in visceral fat in the obese rat (30). Likewise, treatment with recombinant human GH reduces plasma leptin levels in GH-deficient patients in parallel with a fall in fat mass and a rise in lean body mass (31, 32). Thus, it could be inferred that diminished GH secretion in Cushing’s syndrome results in disinhibition of leptin secretion (33, 34). Indeed, patients with Cushing’s disease evince a negative relationship between GH and cortisol secretion rates (35). Nonetheless, overtly subnormal GH secretion was only present in Cushing’s patients with severe hypercortisolism. Thus, attenuation of GH secretion in Cushing’s disease may be a plausible, but not exclusive, factor in the pathophysiology of hyperleptinemia in this disorder.

Chronic hyperinsulinemia also appears to stimulate leptin production (36–41). Insulin can heighten leptin secretion by human adipocytes in vitro (36). These data are consistent with, but not proof of, a causal relationship between hyperinsulinism and increased leptin production, as observed here jointly in Cushing’s disease. Analogous reasoning has been applied to obese patients with polycystic ovary syndrome, although this relationship is not evident in all studies (42–44).

Rhythmic leptin secretion was similar in patients and controls. The genesis of the 24 h variations in leptin production is not well established, except that such rhythmicity is not caused exclusively or primarily by diurnal variations in energy intake (45). Inasmuch as fasting maintains the nyctohemeral rhythmicity of leptin synthesis and secretion as modulated by ambient putative neural signals, our results tend to indicate that the latter regulatory inputs to fat cells are probably unchanged in women with Cushing’s disease.
The ApEn statistic was used to monitor the minute-to-minute pattern reproducibility of leptin release. This function quantifies non-pulsatile and non-circadian regulation of hormone secretion (46). Leptin ApEn values were similar in patients and controls, which signifies comparable feedback and feedforward control of leptin secretion. In contrast, in neuroendocrine tumoral states, secretion is characterized by increased disorderliness (high ApEn), mirroring loss of coordinate feedback control (47–49). For example, in relation to adrenocorticotropin (ACTH) and cortisol secretion, Cushing’s disease is marked by disruption of orderly ACTH secretion, and by erosion of the joint synchrony between ACTH and cortisol release (50). The unexpected stability of joint cortisol–leptin and leptin–GH secretion in Cushing patients points to maintained fat-cell responsiveness, at least to these two opposing metabolotropic signals (cortisol to stimulate leptin production and GH to inhibit leptin production).

In obese patients, leptin gene expression is increased in subcutaneous fat (51). Whether a similar mechanism operates in Cushing’s patients is not yet known, but it would account for most of the foregoing amplitude-specific mechanisms driving leptin overproduction, while sparing frequency and regularity measures. Likewise, more subcutaneous fat cells in Cushing patients (52) would explicate many of the present findings, as well as the early observation (above) of delayed resolution of hyperleptinemia after surgical curing of Cushing’s syndrome.

References


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