Effects of acute infusion of erythropoietin on paradoxical responses of growth hormone to thyrotropin-releasing hormone in acromegalic patients

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Abstract
Objective: Our aim has been to evaluate the effects of i.v. infusion of recombinant human erythropoietin (rhEPO) on the responses of growth hormone (GH), prolactin (PRL) and thyrotropin (TSH) to thyrotropin-releasing hormone (TRH) stimulation in acromegalic patients.

Methods: We studied 16 patients (8 females, aged 29–68 years) with active acromegaly and 12 control subjects (7 females, 24–65 years). All participants were tested with TRH (400 μg i.v. as bolus) and with TRH plus rhEPO (40 U/kg at a constant infusion rate for 30 min, starting 15 min before TRH injection) on different days. Blood samples were obtained between −30 and 120 min for GH and PRL determinations, and between −30 and 90 min for TSH determinations. Hormone responses were studied by a time-averaged (area under the secretory curve (AUC)) and time-independent (peak values) analysis.

Results: Twelve patients exhibited a paradoxical GH reaction after TRH administration with great interindividual variability in GH levels. When patients were stimulated with rhEPO plus TRH there were no changes in the variability of GH responses or in the peak and AUC for GH secretion. Infusion with rhEPO did not induce any significant change in GH secretion in normal subjects. Baseline and TRH-stimulated PRL concentrations in patients did not differ from those values found in controls. When TRH was injected during the rhEPO infusion, a significant (P<0.05) increase in PRL concentrations at 15–120 min was found in acromegalic patients. Accordingly, the PRL peak and the AUC for PRL secretion were significantly increased in patients. Infusion with rhEPO had no effect on TRH-induced PRL release in control subjects. Baseline TSH concentrations, as well as the TSH peak and the AUC after TRH, were significantly lower in patients than in controls. Infusion with rhEPO modified neither the peak TSH reached nor the AUC for TSH secretion after TRH injection in acromegalic patients and in healthy volunteers.

Conclusion: Results in patients with acromegaly suggest that (i) the paradoxical GH response to TRH is not modified by rhEPO infusion, (ii) rhEPO has no effect on TRH-induced TSH release, and (iii) acute rhEPO administration increases the TRH-induced PRL release in acromegalic patients.
hormone (GHRH) after chronic rhEPO administration has been found in uremic patients undergoing hemodialysis (9) or continuous ambulatory peritoneal dialysis (10). Potentiation of GH responses to GHRH has also been obtained in a group of hemodialyzed patients after acute administration of an i.v. rhEPO infusion (11). Furthermore, the paradoxical response of GH to TRH that is frequently associated with uremia has been reported to be abolished after correction of the anemia with chronic rhEPO therapy (12), and acutely administered rhEPO has been shown to inhibit this abnormal GH release in patients with chronic renal failure (13).

Together, these data suggest that rhEPO might exert some effects at hypothalamic or pituitary level, and support the hypothesis that this peptide can modulate the response of somatotrope cells through a mechanism other than increased oxygen supply. Based on this background, the present study was performed with the aim of evaluating the influence of acutely administered rhEPO on the responses of GH to TRH stimulation in a group of patients with active acromegaly. We have also assessed the responses of prolactin (PRL) and thyroid-stimulating hormone (TSH) after stimulation with TRH plus rhEPO in patients with active acromegaly.

Subjects and methods

Patients

Sixteen patients with active acromegaly and 12 healthy volunteers, whose main clinical and analytical characteristics are shown in Table 1, were studied. The study was approved by the local ethical committee, and informed consent was obtained from all participants before testing. There were no significant differences between acromegalic patients and control subjects in terms of gender distribution, age, systolic and diastolic blood pressure, hemoglobin concentrations, hematocrit values, or plasma concentrations of urea, creatinine, glucose, cholesterol, triglycerides, albumin and free thyroxine. Patients showed a higher body mass index, higher levels of plasma phosphorus concentrations and lower levels of calcium and total protein than controls. Diagnosis of active acromegaly was based on clinical features and the presence of elevated random GH concentrations. At the time of the study all patients showed IGF-I concentrations higher than 450 µg/l and GH levels higher than 2 µg/l after a 75 g oral glucose load. On magnetic resonance imaging, nine of the patients had a macroadenoma (> 10 mm in diameter) and seven had an intrasellar microadenoma. All patients presented with pure GH-secreting tumors. Three of them had been treated by trans-sphenoidal incomplete surgical resection of the pituitary adenoma, performed at least 6 months before this study. One patient was treated by trans-sphenoidal surgery followed by external conventional radiotherapy seven years before the study. Bromocriptine was used in three patients and octreotide in one. Drug therapy was stopped at least three weeks before the study. Nine patients had received no therapy for acromegaly. Four patients had non-insulin-dependent diabetes mellitus controlled by diet (n = 2) or oral drugs (n = 2), and four patients had hypertension controlled with anti-hypertensive therapy. Suppressive doses of levothyroxine were employed in one patient who had a previous history of papillary thyroid carcinoma. This patient was excluded from TSH evaluation.

Study design

Endocrine tests were begun at 0900 h after an overnight fast, with the subjects recumbent. An indwelling catheter was placed in a forearm vein and kept patent with a slow infusion of 0.9% NaCl. Each subject received TRH (TRH Prem, Zyma-Frumtost, Gland, Switzerland), 400 µg i.v. as a bolus at time 0. Blood samples were collected at ±30, 0, 15, 30, 60, 90 and 120 min. In another experiment, 5 days apart, the subjects were tested with TRH plus rhEPO (Erantin, Boehringer Mannheim, Mannheim, Germany) at 40 IU/kg body weight, infused at a constant rate for 30 min, starting 15 min before TRH administration. The tests with TRH

Table 1 Clinical and analytical data on acromegalic patients and control subjects (means ± S.E.M.).

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Patients (n = 16)</th>
<th>Controls (n = 12)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>49.88 ± 2.95</td>
<td>44.67 ± 4.05</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>79.14 ± 2.73</td>
<td>67.53 ± 3.36</td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
<td>29.40 ± 1.06</td>
<td>24.11 ± 0.56</td>
</tr>
<tr>
<td>Systolic blood pressure (mmHg)</td>
<td>140.8 ± 4.2</td>
<td>127.9 ± 3.9</td>
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<tr>
<td>Diastolic blood pressure (mmHg)</td>
<td>85.3 ± 3.3</td>
<td>79.2 ± 1.4</td>
</tr>
<tr>
<td>Time of evolution (years)</td>
<td>5.38 ± 0.78</td>
<td></td>
</tr>
<tr>
<td>Diabetes mellitus (yes/no)</td>
<td>4/12</td>
<td>0/12</td>
</tr>
<tr>
<td>Hypertension (yes/no)</td>
<td>4/12</td>
<td>0/12</td>
</tr>
<tr>
<td>Tumoral status (macro/micro)</td>
<td>9/7</td>
<td>–</td>
</tr>
</tbody>
</table>

Comparisons between patients and controls (Mann–Whitney U test): *P < 0.05; **P < 0.01; ***P < 0.001.

* GH measurement after 75 g oral glucose load.

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alone and TRH plus rhEPO were performed in a randomized order. In all blood samples, plasma GH and PRL concentrations were assessed. TSH concentrations were measured between –30 and 90 min. Blood hemoglobin concentrations and hematocrit values, and serum concentrations of urea, creatinine, calcium, phosphorus, total protein, albumin, cholesterol, triglycerides and free thyroxine, were also determined at time 0 in one of the experiments.

**Hormone assays**

Blood samples were centrifuged immediately and the plasma stored at –20°C until assayed. Human plasma GH concentrations were determined by using an automated immunoenzymatic assay (AIA 1200, Tosoh, Tokyo, Japan). Maximal intra-assay and inter-assay coefficients of variation were 5.4 and 3.3% respectively. The sensitivity of the GH assay was 0.1 µg/l. Plasma TSH and PRL concentrations were also determined using the Tosoh immunoenzymatic assay. For TSH assay, the sensitivity was 0.06 µU/ml and the maximal intra-assay and inter-assay coefficients of variation were 3.3 and 3.4% respectively. For PRL assay, the sensitivity and maximal intra-assay and inter-assay coefficients of variation were, respectively, 1 µg/l, 6% and 4.5%. Free thyroxine was measured by commercially available immunoenzymatic assay kits (AIA-PACK FT4, Tosoh, Tokyo, Japan) that use the automated system AIA-1200. Maximal intra-assay and inter-assay coefficients of variation were 9.6 and 7.7% respectively. The sensitivity of FT4 assay was 0.1 ng/dl. The plasma IGF-I assay was performed after an ethanol–acid extraction by means of a commercially available radio-immunoassay kit (Nichols Institute, San Juan Capistrano, CA, USA). Maximal intra-assay and inter-assay coefficients of variation were 3.0 and 8.4% respectively, and the sensitivity of the assay was 13.5 µg/l. Blood hemoglobin concentrations and hematocrit values were measured in a Coulter counter, and the serum chemistry determinations were made using an automated multichannel analyser.

**Statistical analysis**

Results are expressed as means ± S.E.M. The hormonal secretory responses were studied by a time-averaged curve (area under the curves) and time-independent (peak values) analysis. Peak hormonal concentration was considered in each test as the maximum level reached by GH, PRL, or TSH regardless of the time taken to do so. The areas under the secretory curve (AUC) for GH and PRL were calculated between 0 and 120 min by a trapezoidal method. For TSH, the AUC were calculated between 0 and 120 min by the sensitivity and maximal intra-assay and inter-assay coefficients of variation were 3.0 and 8.4% respectively. The sensitivity of the GH assay was 0.1 µg/l, 6% and 4.5%. Free thyroxine was measured by commercially available immunoenzymatic assay kits (AIA-PACK FT4, Tosoh, Tokyo, Japan) that use the automated system AIA-1200. Maximal intra-assay and inter-assay coefficients of variation were 9.6 and 7.7% respectively. The sensitivity of FT4 assay was 0.1 ng/dl. The plasma IGF-I assay was performed after an ethanol–acid extraction by means of a commercially available radio-immunoassay kit (Nichols Institute, San Juan Capistrano, CA, USA). Maximal intra-assay and inter-assay coefficients of variation were 3.0 and 8.4% respectively, and the sensitivity of the assay was 13.5 µg/l. Blood hemoglobin concentrations and hematocrit values were measured in a Coulter counter, and the serum chemistry determinations were made using an automated multichannel analyser.

Results

**Growth hormone responses**

As expected, baseline GH concentrations were significantly higher in patients than in controls (10.6 ± 3.8 vs 1.3 ± 0.3 µg/l, P < 0.001; Table 2). A paradoxical response of GH in the TRH test was considered to be present when the GH concentration increased > 100% from baseline values (2). According to this criterion, all but four patients exhibited paradoxical GH reactions after TRH injection. GH concentrations after stimulation showed a marked interindividual variability, ranging from 4 to 380 µg/l (mean 41.2 ± 23.0 µg/l) (Fig. 1A). When patients were stimulated by rhEPO plus TRH there was no significant modification in the variability of GH responses (Fig. 1A) or in the peak concentrations reached (45.5 ± 25.4 µg/l). In consequence, the AUC of GH after rhEPO plus TRH did not differ from that found after stimulation with TRH alone (Table 2).

In contrast to these findings in acromegalic patients, most of the normal subjects exhibited a flat response to both TRH and rhEPO plus TRH infusion (Fig. 1A). On average, peak GH concentrations in healthy subjects were 2.0 ± 0.5 µg/l after TRH and 2.2 ± 0.8 µg/l (P > 0.05) after rhEPO plus TRH. In a similar way, rhEPO infusion did not induce any significant change in the AUC of GH secretion in response to TRH in normal subjects.

We have also studied GH responses in male and female patients and controls separately. We could not find any significant change, induced by rhEPO infusion, in GH peaks or AUC elicited by TRH stimulation when analyzing responses in male and in female subjects.

**Prolactin responses**

Baseline PRL concentrations in patients ranged from 1 to 28 µg/l (mean 10.0 ± 2.1), i.e. no one had overt hyperprolactinemia. TRH injection was followed by a prompt rise in PRL concentrations that reached a peak of 27.6 ± 4.7 µg/l at 15–30 min. We could not find any significant differences between patients and control subjects in terms of baseline and TRH-stimulated PRL concentrations (Table 2). Nevertheless, when TRH was injected during the rhEPO infusion, a significant increase in the maximum PRL concentration was observed in acromegalic patients (36.3 ± 4.8 µg/l, P < 0.01) in relation to the PRL peak after TRH alone. Analysis of the concentration versus time curves showed that rhEPO infusion induced a significant increase in PRL levels 15 to 120 min (P < 0.05) after
TRH injection (Fig. 1B). Accordingly, the AUC of prolactin were significantly higher when patients were stimulated with rhEPO plus TRH (51.1 ± 6.9 µg.h/l) than when TRH was injected alone (36.5 ± 5.6 µg.h/l, P < 0.01). All but three patients exhibited increases in their TRH-induced peaks of PRL concentration when stimulated with rhEPO. These increments ranged from 5.3 to 300% (mean 65.7 ± 24.1%). When studying the correlation between baseline PRL levels and the percentage of change in peak prolactin concentrations induced by rhEPO we found a non-significant relationship (r = -0.328, N.S.).

In the group of 12 control subjects, the PRL peak was also elicited at 15–30 min of TRH administration and reached a value of 37.1 ± 4.9 µg/l when TRH was administered alone and 41.9 ± 8.9 µg/l when TRH was injected during the rhEPO infusion (P > 0.05). No significant differences were observed between PRL responses to TRH (estimated by peaks and AUC) in the presence or in absence of rhEPO in normal subjects.

When PRL responses were classified according to gender, we observed that the significant increase (P < 0.05) in PRL peaks and AUC after rhEPO infusion was present in both male and female acromegalic patients. No significant changes were found in male or female healthy volunteers (Table 3).

**Thyrotropin responses**

Baseline TSH concentrations in acromegalic patients were lower than those found in control subjects

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### Table 2 Baseline and stimulated GH, PRL and TSH concentrations in acromegalic patients and control subjects (means ± S.E.M.).

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>Patients (n = 16)</th>
<th>Controls (n = 12)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>GH responses</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline concentrations</td>
<td>µg/l</td>
<td>10.6 ± 3.8</td>
<td>1.3 ± 0.3</td>
</tr>
<tr>
<td>Peak concentrations after TRH</td>
<td>µg/l</td>
<td>41.2 ± 23.0</td>
<td>2.0 ± 0.5</td>
</tr>
<tr>
<td>Peak concentrations after rhEPO + TRH</td>
<td>µg/l</td>
<td>45.5 ± 25.4</td>
<td>2.2 ± 0.8</td>
</tr>
<tr>
<td>AUC after TRH</td>
<td>µg.h/l</td>
<td>62.8 ± 36.8</td>
<td>2.6 ± 0.4</td>
</tr>
<tr>
<td>AUC after rhEPO + TRH</td>
<td>µg.h/l</td>
<td>66.4 ± 36.7</td>
<td>2.3 ± 0.2</td>
</tr>
<tr>
<td><strong>PRL responses</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline concentrations</td>
<td>µg/l</td>
<td>10.0 ± 2.1</td>
<td>6.9 ± 0.9</td>
</tr>
<tr>
<td>Peak concentrations after TRH</td>
<td>µg/l</td>
<td>27.6 ± 4.7</td>
<td>37.1 ± 4.9</td>
</tr>
<tr>
<td>Peak concentrations after rhEPO + TRH</td>
<td>µg/l</td>
<td>36.3 ± 4.6</td>
<td>41.9 ± 8.9</td>
</tr>
<tr>
<td>AUC after TRH</td>
<td>µg.h/l</td>
<td>36.5 ± 5.6</td>
<td>44.4 ± 5.3</td>
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<tr>
<td>AUC after rhEPO + TRH</td>
<td>µg.h/l</td>
<td>51.1 ± 6.9*</td>
<td>50.3 ± 9.2</td>
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<tr>
<td><strong>TSH responses</strong></td>
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<td></td>
<td></td>
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<tr>
<td>Baseline concentrations</td>
<td>mU/l</td>
<td>0.52 ± 0.10</td>
<td>1.05 ± 0.17</td>
</tr>
<tr>
<td>Peak concentrations after TRH</td>
<td>mU/l</td>
<td>4.85 ± 0.63</td>
<td>8.83 ± 1.12</td>
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<tr>
<td>Peak concentrations after rhEPO + TRH</td>
<td>mU/l</td>
<td>5.10 ± 0.55</td>
<td>8.43 ± 1.13</td>
</tr>
<tr>
<td>AUC after TRH</td>
<td>mU.h/l</td>
<td>4.88 ± 0.69</td>
<td>9.24 ± 1.44</td>
</tr>
<tr>
<td>AUC after rhEPO + TRH</td>
<td>mU.h/l</td>
<td>5.23 ± 0.59</td>
<td>9.06 ± 1.39</td>
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</tbody>
</table>

Comparisons between patients and controls (Mann–Whitney U test): *P<0.05; **P<0.01; ***P<0.001.

Comparisons within each group (responses after TRH and after rhEPO plus TRH; Wilcoxon test): *P<0.001.

§ Patients, n = 15.

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### Table 3 PRL responses after TRH and rhEPO plus TRH in acromegalic patients and control subjects classified according to gender (means ± S.E.M.).

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>Patients (n = 8)</th>
<th>Controls (n = 7)</th>
<th>Patients (n = 8)</th>
<th>Controls (n = 5)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline concentration</td>
<td>µg/l</td>
<td>9.5 ± 3.0</td>
<td>5.6 ± 0.7</td>
<td>10.5 ± 3.0</td>
<td>8.8 ± 1.5</td>
</tr>
<tr>
<td>Peak concentration after TRH</td>
<td>µg/l</td>
<td>19.4 ± 3.7</td>
<td>32.6 ± 5.2</td>
<td>35.8 ± 7.8</td>
<td>43.4 ± 9.0</td>
</tr>
<tr>
<td>Peak concentration after rhEPO + TRH</td>
<td>µg/l</td>
<td>26.6 ± 5.3*</td>
<td>41.0 ± 14.3</td>
<td>46.0 ± 6.6*</td>
<td>43.2 ± 9.7</td>
</tr>
<tr>
<td>AUC after TRH</td>
<td>µg.h/l</td>
<td>27.8 ± 6.0</td>
<td>37.7 ± 4.8</td>
<td>45.3 ± 8.7</td>
<td>53.8 ± 9.7</td>
</tr>
<tr>
<td>AUC after rhEPO + TRH</td>
<td>µg.h/l</td>
<td>39.8 ± 8.8*</td>
<td>48.5 ± 15.1</td>
<td>62.3 ± 8.8*</td>
<td>52.9 ± 9.1</td>
</tr>
</tbody>
</table>

Comparisons between patients and controls (Mann–Whitney U test): N.S.

Comparisons within each group (responses after TRH and after rhEPO plus TRH; Wilcoxon test): *P<0.05.
(0.52 ± 0.10 vs 1.05 ± 0.17 mU/l, \( P < 0.01 \)). In patients, these levels reached a peak of 4.85 ± 0.63 mU/l 30 min after TRH stimulation. When this stimulus was injected during rhEPO infusion there were no significant modifications in the time or the magnitude (5.10 ± 0.55 mU/l) of the TSH peak (Fig. 1C, Table 2). TSH AUC obtained after rhEPO plus TRH stimulation were also very similar to values found after the injection of TRH alone (Table 2).

In the group of 12 normal volunteers, TRH injection was followed by an increase in TSH concentrations that reached a maximum of 8.83 ± 1.12 mU/l after 30 min. TRH administration during rhEPO infusion gave rise to a peak TSH concentration (8.43 ± 1.13 mU/l) that did not differ from that found after TRH alone. The AUC for TSH secretion obtained in control subjects after rhEPO plus TRH stimulation did not differ from values obtained after injection of TRH alone (Table 2).

As shown in Table 2, control subjects exhibited higher values for TSH peaks and TSH AUC in comparison with patients after stimulation with TRH alone and also after rhEPO plus TRH infusion. On the other hand, no significant differences were observed in baseline TSH concentrations and its response to TRH or rhEPO plus TRH in patients or controls divided according to gender. No subjective side-effects were reported during the tests, either in control subjects or in patients with acromegaly.
Discussion

The results of this work clearly show that acutely administered rhEPO has no influence on GH responses to TRH stimulation in a group of acromegalic patients, with 12 out of 16 subjects exhibiting a GH increase of at least twice the basal level when tested with TRH alone (2). As in other studies, the variability in the GH responses to TRH may be accounted for by the intrinsic secretory characteristics of each adenoma (14). Previous investigations by Ramirez et al. (12) have demonstrated that chronic therapy with rhEPO is accompanied by a disappearance of the paradoxical response of GH to TRH in hemodialyzed patients (12). Our group has reported that acute infusion of rhEPO also abolishes this paradoxical GH reaction in patients with chronic renal failure (13).

In uremic patients, this abolition of GH release after TRH has been attributed to a possible enhancement of hypothalamic somatostatin release or a reduction of the release of GHRRH (13). However, the mechanisms underlying the paradoxical GH responses to TRH seem quite different in patients with acromegaly in relation to uremic patients. Firstly, the role of somatostatin in the regulation of TRH-stimulated GH release in acromegaly is less clear. Somatostatin may either inhibit (15) or fail to affect (16) the GH release after TRH in human GH tumor cells. In vivo, a blunting of the paradoxical GH response to TRH during high-dose (1000 μg/h) somatostatin infusion has been reported (17), although low-dose (100 μg/h) somatostatin infusion (18) did not modify the paradoxical GH response to TRH in acromegalic patients. These responses have also been shown to disappear in the majority of patients with acromegaly treated long-term with the long-acting somatostatin analogue octreotide (19–21). Interestingly, the acute increase in hypothalamic somatostatin tone caused by glucocorticoids did not modify the paradoxical GH response to TRH in acromegalic patients (14), although the glucocorticoid-induced increase in somatostatin tone is only speculative. Other studies have shown that cholinergic blockade with atropine is unable to suppress the GH reaction to TRH in patients with acromegaly (22). Hence, cholinergic pathways seem not to be involved in the paradoxical GH reaction to TRH in acromegaly (22).

Lastly, it has been reported that endogenous opiate receptors do not play a major role in modulating GH secretion in acromegaly (23, 24), and that opioid peptides failed to affect TRH-induced stimulation of GH secretion in adenomatous human pituitary cells (25).

A direct pituitary stimulatory action of TRH has been postulated (26–28). In rats, it has been shown that TRH can elicit GH secretion from normal or tumoral somatotropic cells (29) and mRNA for the TRH receptor has been localized in somatotropes (30). Paradoxical GH responses to TRH have been observed in dispersed human GH-secreting adenomatous cells that probably express TRH receptors (31), and the disappearance of these responses after adenomectomy has been reported in acromegalic patients (27). The mechanism of action of TRH has been investigated in human GH-secreting adenomas in several in vitro studies (26, 32). The human TRH receptor, whose gene has been cloned, is a member of the G protein-coupled receptor family (33). Mutations in this receptor might be involved in the etiology of pituitary adenomas, although a recent study found that the TRH-receptor structure was normal in TSH-, PRL- and GH-secreting pituitary adenomas (34). A dedifferentiation of the receptors on the somatotrophs, with a subsequent GH response to TRH, has also been proposed as a possible explanation for the TRH-induced GH release in acromegaly (25). These findings support the idea that the paradoxical GH reaction to TRH in acromegaly may be due to the presence of adenomatous tissue and that TRH may directly stimulate GH secretion at the adenoma level through interaction with specific receptors. In this setting, rhEPO seems to be unable to interfere with this hormone–receptor interaction.

TRH is considered to have no effect on GH secretion in healthy humans, and our results in healthy volunteers suggest that rhEPO does not exert any influence on these responses. However, abnormal GH responses to TRH have also been reported in patients with hypothyroidism (35), anorexia nervosa (36), diabetes mellitus (37), liver cirrhosis (38) and status epilepticus (39). Investigations of these conditions suggest that the paradoxical GH reaction to TRH may be a functional consequence of several pathological states characterized by altered somatostatin control of GH secretion.

Our results show that acute infusion of rhEPO does not modify PRL responses to TRH in normal subjects, but significantly enhances PRL secretion in normoprolactinemic acromegalic patients, irrespective of the gender of the subjects. Although no patient studied exhibited overt hyperprolactinemia, two had basal PRL levels higher than 20 μg/l (24 and 28 μg/l). It is possible that PRL secretion in these patients could in fact be of tumoral origin or due to hypothalamic–pituitary deafferentation, since it has been reported that mixed GH–PRL-secreting adenomas do not always cause high serum levels of PRL (40). No relationship between baseline PRL concentrations and the effect of rhEPO on TRH-induced PRL release was found. Data obtained in controls are in agreement with results reported by Bernini et al. (41), who found no change in PRL responses to TRH after acute injection of rhEPO in normal subjects. Nevertheless, a normalization of the elevated PRL has been well established in uremic patients after chronic therapy with rhEPO (12, 42, 43). Discrepancies between our findings and those reported for uremic patients may be explained by the fact that uremic patients show elevated PRL levels, whereas the group of acromegalic patients studied here exhibit normal or near-normal PRL levels. In addition,
distinct neuroregulatory pathways of PRL secretion may act in these two pathological conditions. The mechanism of rhEPO action on prolactin secretion is not known. A direct action by the peptide on the physiological mechanism regulating PRL secretion in uremic patients seems unlikely (41), and a better oxygen supply due to the improvement of the anemia has been suggested as the causative factor of the PRL-reducing effect observed in these patients. Our results in acromegals suggest that a direct trophic action on rhEPO at the hypothalamic or pituitary levels might be possible. In fact, the effects of acutely administered rhEPO on GH responses to GHRIH (11) and TRH (13) and on LH responses to GnRH (44) in uremic patients may not be explained by an elevation in hematocrit values.

A tentative explanation for our findings in acromegalic patients could be that rhEPO would interfere with the dopamine hypothalamic pathways, thus rendering lactotrophs more responsive to the stimulation with TRH. This action seems unlikely since, in acromegals, dopaminergic stimulation inhibits PRL and GH release (2, 45) and we could not find any effect of rhEPO on GH responses. Other pathways, such as those mediated by serotonin, GABA, opioids and histamine, could also be involved. Therefore, the mechanism by which rhEPO can modulate PRL release in acromegalic patients remains unclear despite the present results.

We have also tested the effect of rhEPO on the TSH response to TRH in patients and controls. Baseline and TRH-stimulated TSH levels were significantly lower in acromegals than in controls, as reported in other studies (46). Suppression of TSH levels in euthyroid acromegals has been accounted for by a chronic suppression of TRH neurons by a compensatory hypersecretion of somatostatin (47), and by a GH-stimulated increase in the conversion of T4 into T3 (48). TSH responses to its physiological releasing hormone are not influenced by acute infusion of rhEPO in acromegalic patients or in normal subjects. A similar finding was previously reported by our group in patients with chronic renal failure (13). In addition, no changes in basal (49) and TRH-stimulated (50) levels have been found in uremic patients after correction of the anemia by rhEPO therapy.

In summary, our findings show that acute rhEPO infusion cannot modify either the paradoxical GH reaction or the normal TSH release in response to TRH stimulation in patients with acromegaly. No effects of rhEPO were observed in controls for GH, TSH and PRL release after TRH administration. On the contrary, our data also show that, although the PRL-inhibiting effect of rhEPO is well known in uremic patients, acute administration of this peptide enhances TRH-stimulated prolactin release in normoprolactinemic patients with GH-secreting pituitary adenoma. A possible direct action at pituitary or hypothalamic level is suggested for such patients.

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