Sex hormone-binding globulin as a marker for hyperinsulinemia and/or insulin resistance in obese children

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Abstract

Objective: A relationship between hyperinsulinemia and decreased serum sex hormone-binding globulin (SHBG) has been described in adults. We evaluated the usefulness of SHBG as an index of hyperinsulinemia and/or insulin resistance in obese children (aged 6–9 years) of both sexes and its possible influence on the androgenic status.

Design: We carried out a cross-sectional study of cases and controls. We studied 61 obese children (22 males, 39 females) with body mass index (BMI) superior to the 90th percentile and a control group of age- and sex-matched non-obese children. We measured serum glucose, insulin, TSH, free thyroxine, 17β-estradiol, testosterone and SHBG. Also, we correlated these parameters with anthropometric measures.

Results: The obese group presented significantly elevated levels of insulin (P<0.001) and insulin/glucose ratio (P<0.0012) compared with the control group. SHBG (P<0.0001) and testosterone (P<0.0169) levels were significantly lower than those in the non-obese group. We did not find any difference in the free androgen index (FAI). Fasting insulin (r=-0.4512; P<0.001), BMI (r=-0.3185; P<0.05) and testosterone (r=-0.3705; P<0.01) were inversely correlated with SHBG concentration. According to multivariate analyses, insulin was the only independent predictor factor for serum SHBG concentration in the obese group (r partial=0.1280; P=0.0171).

Conclusions: In summary, at this age there is a strong relationship between insulin and SHBG. The changes in SHBG levels of the obese group did not affect FAI and, therefore, they did not cause changes in the androgenic status. Our data support the role of insulin in the regulation of serum SHBG levels.

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Introduction

Sex hormone-binding globulin (SHBG) is a specific steroid-binding plasma glycoprotein, which is mainly synthesized in the liver (1–3). It binds testosterone with high affinity and estrogens with lower affinity. Assuming that only the unbound fraction of testosterone is bioavailable, SHBG functions as a modulator of androgen delivery to tissues.

In the classical paper of Anderson (4), the estrogen/testosterone balance was the main factor in the regulation of SHBG synthesis. Nevertheless, at present, SHBG regulation is considered as multifactorial and other non-steroidal factors seem to have an important role in the circulating levels of this binding protein (5, 6).

Now, there is much evidence that insulin may be an important modulator of SHBG concentrations. In vitro, insulin is a potent inhibitor of SHBG production by HepG2 cells (7), and reduces the stimulatory effect of 17β-estradiol and thyroxine (8). In vivo, reduced serum SHBG levels have been described in different insulin resistance states with hyperinsulinemia, such as polycystic ovary syndrome (9) or obesity (10). Also, low SHBG concentration is an independent risk factor for non-insulin-dependent diabetes mellitus (NIDDM) (11). Therefore, the relationship between decreased SHBG levels and insulin resistance is present in clinical situations with an increased risk of cardiovascular disease.

Most of the previous studies have been accomplished in adults and little is known of this relationship in children. However, recent studies indicate that features typical of the insulin resistance syndrome with an adverse cardiovascular risk are already present in adolescents (12, 13). The current study was designed to investigate whether the SHBG level is a useful index of hyperinsulinemia and/or insulin resistance in obese children (six to nine years old) of both sexes and, if so, whether this association influences the androgen status.
of these children. Also, we seek to correlate these parameters with anthropometric measures.

**Subjects and methods**

**Subjects**
We carried out a cross-sectional study of cases and controls. We studied a total of 122 children aged between six and nine years. Sixty-one obese children (22 males, 39 females) with a body mass index (BMI) above the 90th percentile, and the same number of age- and sex-matched non-obese children as a control group (BMI below the 90th percentile) were studied. We used curves of growth for our population (14). All children were in the Tanner stage 1.

The study groups were formed of children from several schools. First, their corresponding pediatricians in the different schools were informed about the realization of the study. Simultaneously, they requested parental authorisation for the participation of their children. All had to give written consent.

After this, we selected the children consecutively among those that accepted participation in the project. The selection of the children was made in the pediatric clinic of health care and each child was included in one or another group according to his BMI, age and sex. Children with primary hyperlipidemia, hypertension, diabetes or hydrocarbonated intolerance were excluded from the study. Any child undergoing pharmacological treatment was also excluded.

**Methods**

The blood samples were collected after 12 h of fast from a vein in the antecubital fossa, without venous occlusion. All collections were made between 0800 and 0900 h. Serum samples were separated from whole blood collected into tubes without anticoagulant. After clotting was complete, the tubes were then centrifuged at 2700 g for 10 min. Serum was removed and assayed or stored in aliquots at −45 °C until the determinations could be performed.

Glucose was determined with a glucose oxidase method in a random access analyzer (Axon, Bayer Diagnostics, Tarrytown, NY, USA). Insulin, thyrotropin (TSH) and free thyroxine (T4) were quantified by a microparticle immunoassay in an automatic analyzer (IMx, Abbott Laboratories, Chicago, IL, USA). We used enzyme immunoassay methods for 17β-estradiol and testosterone (BioChem ImmunoSystems, Bologna, Italy), and for SHBG (Rasim S.A., Liège, Belgium), which were carried out in a microtiter plate analyzer (Labotech, Chemila, Roma, Italy). The coefficients of variation were always below 5% (intra-assay) and 9.5% (interassay). The free androgen index (FAI), an alternative assessment of free testosterone, was calculated as 100 times the testosterone/SHBG ratio.

**Statistical analysis**

Statistical assessment was conducted with Microstat (Ecosoft, Inc.) or GraphPAD InStat (GraphPAD Software) software. The abnormal values (outliers) were excluded by Reed’s method. Results were expressed as means ± s.e.m. and with 95% confidence interval (CI). The distribution of each variable was tested for departure from gaussian distribution and the equality of variances was controlled by Snedecor’s F-test. The comparison between the mean values of the groups was carried out by Student’s unpaired t-test. Statistical significance was set at P < 0.05.

Correlation between variables was evaluated by Pearson’s correlation coefficient and with regression analysis. The multivariate regression analysis was performed by using the stepwise method. It is important that the statistical procedure includes a check for regression. We must verify that the distribution of residuals is normal and their variance is constant. In this study, the normality of residuals was checked by the χ² test, and graphic analysis of residuals was carried out to provide information about constancy of variance. Diagnostic measures were used for the detection of problematic observations. An observation was considered an outlier if its residual was >3Sy.x. (standard deviations of residuals). For each variable, potential confounding factors (0.05 < P < 0.2) were evaluated by analysis of raw and adjusted regression coefficients.

**Results**

The patient characteristics (means and s.e.m.) of the studied groups are shown in Table 1 and the measured parameters in Table 2.

The mean values (obese vs control) of insulin (95% CI 7.20–9.20 vs 5.45–6.79) and the insulin/glucose ratio (95% CI 8.35–10.55 vs 6.34–7.94) were significantly greater in obese children (2.24 vs 1.94; P < 0.0131).

**Table 1** Patient characteristics. Results are expressed as means ± s.e.m.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Obese</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>7.7 ± 0.11</td>
<td>7.7 ± 0.12</td>
<td>NS</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>27.3 ± 0.63</td>
<td>37.7 ± 0.80</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>127.2 ± 1.02</td>
<td>129.4 ± 0.84</td>
<td>NS</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>16.7 ± 0.18</td>
<td>22.3 ± 0.27</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Waist/hip ratio</td>
<td>0.84 ± 0.005</td>
<td>0.85 ± 0.006</td>
<td>NS</td>
</tr>
<tr>
<td>Waist/height ratio</td>
<td>1.46 ± 0.01</td>
<td>1.43 ± 0.01</td>
<td>NS</td>
</tr>
</tbody>
</table>

NS, not significant.
Thyroid hormones, which stimulate SHBG secretion, were measured so as to discard any changes in the circulating levels of SHBG as a consequence of the thyroid status, and we found no differences between the thyroid hormone levels of obese and control groups.

Univariate correlation analysis is summarized in Table 3 (obese) and Table 4 (control). In the obese group, SHBG correlated negatively with insulin, insulin/glucose ratio, BMI, testosterone and FAI. The strongest correlation was between SHBG and the FAI/testosterone ratio. Insulin correlated with the same parameters as SHBG but in a positive manner, and also with the waist/hip ratio. The correlation coefficients of the insulin/glucose ratio with these variables were higher than those of insulin. The relationship between SHBG and insulin improved when we used the natural logarithm of insulin (r = -0.4725 vs r = -0.4512) (not shown).

In the control group, nearly 30% of the SHBG concentration was explained by the levels of insulin, BMI and testosterone (R = 0.5457; P = 0.0018). However, only insulin was an independent factor of prediction (r partial=0.1280; P = 0.01711); BMI and testosterone lost statistical significance when corrected for insulin.

In the control group, using stepwise multiple regression analysis, insulin and BMI were selected as predictor variables for SHBG (R = 0.4511; P = 0.00136); however, only BMI remained as an independent factor (r partial = 0.0976; P = 0.01508).

Discussion

The control of plasma SHBG concentration, considered initially to be under the direct regulation of sex hormone levels (4), is considered today to be under multifactorial regulation. Indeed, the basal production of SHBG seems to be steroid independent and more related to general metabolic factors and nutritional status (5, 6, 15). In this way, some hormones such as growth hormone, insulin-like growth factor (IGF-I) and insulin may play a more active role in the regulation of SHBG levels (16–18). Recently, there has been increased interest in the role played by insulin in this multiregulation. It is known that insulin has an inhibitory effect on the synthesis of SHBG. This action may be by a direct mechanism: by using cultured HepG2 cells it has been found that insulin is a potent inhibitor of the basal production of SHBG (7, 8); or by an indirect mechanism: by modifying the IGF-I/IGF binding protein-1 ratio (19) the free fraction of IGF-I is increased, and free IGF-I is also an inhibitor of SHBG synthesis in vitro (20). Anyway, the relationship between insulin and SHBG is a widespread phenomenon in adults (21–24) and is present even in neonates (25).

Therefore, evidence suggests that it is a more physiological process than was originally thought (26).

Because of the intercorrelations between insulin and anthropometric measures, as well as with testosterone, these observations were subjected to multivariate regression analysis to determine which anthropometric and hormonal factors correlated independently with SHBG.

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This association is of special interest because the SHBG level could be a general marker for hyperinsulinemic insulin resistance. This would explain the observation that a low serum SHBG level is a predictor for the development of NIDDM (11, 27, 28). Hyperinsulinemic insulin resistance is also present in obesity, and low levels of SHBG have been described in obese adults of both sexes (10, 29).

In the present study, SHBG was independently and inversely correlated with fasting insulin. Our results confirm that this association, already found in adults, is present in obese children (6 to 9 years old) of both sexes. We studied this age range because it is one of the critical periods in childhood for the development of obesity (30).

We have found a significant increase in insulin in the obese group. Furthermore, the insulin/glucose ratio was also greater than in the control group. These insulin levels, indicative of a hyperinsulinemic state, were correlated with an inverse change in the concentration of SHBG. This relationship is present in both groups, obese and controls, and was sufficiently robust to survive regression analyses. According to the multivariate analyses, insulin is the only independent prediction factor for SHBG levels in obese children. In controls, using a stepwise regression analysis method, insulin and BMI were selected as prediction factors. Therefore, as the relation between SHBG and insulin is present in obese and control groups, we consider that these results support the existence of a physiological relationship.

With respect to the relationship between androgens and anthropometric measures, an association with fat distribution has been described in adults (31). In our study group, we did not find any correlation of testosterone or of SHBG with waist/hip ratio or waist/thigh ratio. These results support the low usefulness of these indices in this age group (32).

We have also evaluated the effect that the decrease in SHBG found in obese children could have on their androgen levels. In our obese group, the total testosterone levels were below those of the control group. However, we cannot consider this result as real hypogonadism since no alteration in FAI levels was found and, therefore, no change in bioavailable testosterone (33). Thus, at this age, the evaluation of androgenic state in obese children is of little use if evaluated only with total testosterone.

Within the physiological regulations, the relationship between insulin, SHBG and testosterone is complex to explain and as yet, not well established. Insulin has stimulatory effects on androgen secretion (34) and, at the same time, testosterone influences tissue sensitivity to insulin (35). In the present study, considering the regression analysis, we believe that at this age (6 to 9 years old), the relationship between insulin, SHBG and testosterone seems defined, fundamentally, by the action of insulin on the SHBG levels. The decrease in SHBG levels produced by the hyperinsulinism in the obese group underlies the decrease in total testosterone, since SHBG is its main transport. Nevertheless, FAI remained unchanged and, therefore, there is no hypogonadism.

On the other hand, we could think about a relative androgenism if we take into account the FAI/testosterone ratio, which was significantly increased in the obese group. But it is important to indicate that this ratio seems to depend almost 100% on the SHBG concentration, as shown in our correlation studies. Therefore, the modifications of this index would be secondary to the changes in SHBG levels. Furthermore, in the obese group, the correlation between testosterone and SHBG did not resist the multivariate analyses. These results do not suggest that, in our study group, androgens have a relevant role in the circulating levels of SHBG.

In conclusion, we believe that the relationship between insulin and SHBG could be considered to have a key role in the regulation of the plasma SHBG concentration. In prepubertals, the SHBG decrease in the obese group is consistently related to the increase in insulin with no change in free androgen index. These results suggest that a hyperinsulinemic insulin resistant state may be present in obese children and that SHBG is a good marker for this hyperinsulinism.

Acknowledgements

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Table 4 Correlation matrix of SHBG with insulin (I), insulin/glucose ratio (I/G), BMI, waist/hip ratio (WHR), waist/thigh ratio (WTR), testosterone (T), FAI and FAI/T ratio in the control group.

<table>
<thead>
<tr>
<th></th>
<th>I</th>
<th>I/G</th>
<th>BMI</th>
<th>WHR</th>
<th>WTR</th>
<th>T</th>
</tr>
</thead>
<tbody>
<tr>
<td>SHBG</td>
<td>-0.3426&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.9905&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.3318&lt;sup&gt;b&lt;/sup&gt;</td>
<td>-0.1247</td>
<td>0.0650</td>
<td>0.0440&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>I</td>
<td>0.3522&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-0.3906&lt;sup&gt;ab&lt;/sup&gt;</td>
<td>0.0860</td>
<td>0.0583</td>
<td>-0.0440</td>
<td>0.1513</td>
</tr>
<tr>
<td>I/G</td>
<td>0.0118</td>
<td>-0.1058</td>
<td>-0.0486</td>
<td>0.2004</td>
<td>0.3391&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.0444</td>
</tr>
<tr>
<td>BMI</td>
<td>-0.0177</td>
<td>0.2205</td>
<td>0.3735&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.3450&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.3473&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.3685&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>WHR</td>
<td>0.0440&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.7149&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.3351&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.3450&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.3473&lt;sup&gt;c&lt;/sup&gt;</td>
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<td>0.3450&lt;sup&gt;c&lt;/sup&gt;</td>
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<tr>
<td>T</td>
<td>0.0440&lt;sup&gt;a&lt;/sup&gt;</td>
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<td>0.3351&lt;sup&gt;c&lt;/sup&gt;</td>
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<td>0.3473&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.3685&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
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</table>

<sup>a</sup>P < 0.0001; <sup>b</sup>P < 0.001; <sup>c</sup>P < 0.01; <sup>d</sup>P < 0.05.
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