EXPERIMENTAL STUDY

Expression of RET and its ligand complexes, GDNF/GFRalpha-1 and NTN/GFRalpha-2, in medullary thyroid carcinomas

Tony Frisk1, Filip Farnebo 1, Jan Zedenius 2, Lars Grimelius 3, Anders Höög 4, Göran Wallin 5 and Catharina Larsson 1

1Department of Molecular Medicine, Endocrine Tumor Unit, Karolinska Hospital, Stockholm, Sweden, 2Center for Metabolism and Endocrinology, Department of Surgery, Karolinska Institutet at Huddinge University Hospital, Huddinge, Sweden, 3Department of Pathology, Uppsala University Hospital, Uppsala, Sweden, 4Department of Pathology, Karolinska Hospital, Stockholm, Sweden and 5Department of Surgery, Karolinska Hospital, Stockholm, Sweden

(Correspondence should be addressed to T Frisk, Department of Molecular Medicine, Endocrine Tumor Unit, CMM L8:01, Karolinska Hospital, S-171 76 Stockholm, Sweden; Email: tony.frisk@cmm.ki.se)

Abstract

Objective: Mutations in the RET proto-oncogene are found in about one third of sporadic medullary thyroid carcinomas (MTCs), mostly affecting codon 918. Glial cell line derived neurotropic factor (GDNF) and its membrane-bound GDNF family receptor alpha (GFRalpha-1), as well as neurturin (NTN) and its membrane-bound receptor GFRalpha-2 form a complex with the RET product, a receptor tyrosine kinase, resulting in downstream signaling to the nucleus.

Design: To elucidate the role of these RET ligands in MTC tumorigenesis, their expression was determined in 15 MTC samples, one papillary thyroid carcinoma (PTC) and three normal thyroid tissue specimens.

Methods: The mRNA expression of RET, GDNF, GFRalpha-1, NTN and GFRalpha-2 was investigated by mRNA in situ hybridization, and confirmed by reverse transcription-PCR analysis.

Results: None of the five genes was expressed in the normal thyroids or in the PTC. All MTCs showed expression of RET, 13 expressed GDNF, 12 expressed GFRalpha-1 and 9 expressed NTN and GFRalpha-2. In 7 of the tumors RET, GDNF and GFRalpha-1 were expressed at high levels, and in five of these seven tumors NTN and GFRalpha-2 genes were also expressed at high levels. The high level of expression was preferentially seen in tumor cells adjacent to stroma and connective tissue. All MTCs without expression of the RET ligands harbored the RET codon 918 mutation.

Conclusions: The results suggest that this signaling pathway is important for MTC development, and that it may be activated by expression of the RET ligand complexes by the tumor cells themselves.

European Journal of Endocrinology 142 643–649

Introduction

Medullary thyroid carcinoma (MTC) is a rare tumor comprising 5–10% of all carcinomas in the thyroid gland. The majority of MTC cases occur sporadically, however, approximately 25% of the patients have a familial form of the disease, such as multiple endocrine neoplasia type 2A and B (MEN 2A and B) and familial MTC (FMTC).

The RET proto-oncogene encodes for a tyrosine kinase receptor. The ligands, glial cell line derived neurotropic factor (GDNF) and neurturin (NTN), and their membrane bound receptors, GFRalpha-1 and GFRalpha-2, form a complex with RET. This results in the dimerization of RET, leading to the activation of its tyrosine kinase function, and allowing it to transduce downstream signaling to the nucleus by phosphorylation of tyrosine residues in signaling proteins (1). In vitro and in vivo studies have shown that GDNF can enhance the survival of various neurons by pre-venting apoptosis (2, 3). NTN and GFRalpha-2 are structurally highly homologous to GDNF and GFR-alpha-1 respectively (4, 5). Recently, two additional ligand complexes have been described for RET. These are artemin and persephin, which activate RET by binding to the membrane bound receptors designated GFRalpha-3 and GFRalpha-4 respectively (6, 7). Thus, at least four different ligand systems are involved in RET signaling.

RET is expressed in both the familial and sporadic forms of MTC. Activating germline RET mutations have been shown to give rise to MEN 2A, MEN 2B, and FMTC and somatic RET mutations have been found in approximately one third of sporadic MTCs (8, 9).
somatic mutation in the RET gene commonly found in sporadic MTC is identical to the constitutional mutation found in MEN 2B at codon 918 in exon 16, where a methionine is substituted with a threonine. This is a highly conserved substrate-recognition site in the catalytic core of the tyrosine kinase domain, and the presence of this mutation in sporadic MTC has been suggested to correlate with a poor prognosis (10). In addition to the classical Met918Thr mutation, somatic RET mutations in MTC have occasionally been described in other locations including codons 611, 634, 768, and 883 (11). In MTCs with RET mutations, ligand independent receptor dimerization and/or substrate specificity shift may occur (11). However, the ligands have also been shown to increase the transforming activity in cells with the codon 918 mutation, indicating that the RET ligands are important in the oncogenetic pathway (12, 13). Hence, in MTCs without detectable RET mutation, a logical probability is that GDNF/GFRalpha-1 or NTN/GFRalpha-2 are somehow involved in the tumor development process. This involvement could theoretically comprise over-expression of the protein, stimulation of different pathways or constitutively activating point mutations.

To further elucidate the role of RET and its ligand complexes in MTC tumorigenesis, the expression of these five genes was analyzed in a panel of such tumors by mRNA in situ hybridization.

Table 1 Data on patients, tumor specimens and results from in situ hybridization.

<table>
<thead>
<tr>
<th>Case no.</th>
<th>I.D. no.</th>
<th>Sex / Age at diagnosis</th>
<th>Diagnosis</th>
<th>Sample</th>
<th>RET mutation</th>
<th>Survival</th>
<th>Expression</th>
<th>RET</th>
<th>GDNF</th>
<th>GFRalpha-1</th>
<th>NTN</th>
<th>GFRalpha-2</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>46</td>
<td>M / 40</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>DfD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>2</td>
<td>378</td>
<td>M / 65</td>
<td>MTC</td>
<td>Primary</td>
<td>No</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>3</td>
<td>378N</td>
<td>Normal</td>
<td></td>
<td>f</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>847</td>
<td>M / 58</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>5</td>
<td>1229</td>
<td>F / 73</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>6</td>
<td>1229N</td>
<td>Normal</td>
<td></td>
<td>No</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>1606</td>
<td>F / 73</td>
<td>MTC</td>
<td>Primary</td>
<td>No</td>
<td>AwoD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>8</td>
<td>1825</td>
<td>F / 69</td>
<td>MTC</td>
<td>Primary</td>
<td>No</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>9</td>
<td>1848</td>
<td>M / 44</td>
<td>MTC</td>
<td>Metastasis</td>
<td>No</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>10</td>
<td>135</td>
<td>F / 65</td>
<td>MTC</td>
<td>Primary</td>
<td>DfD</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>595</td>
<td>M / 35</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>12</td>
<td>1072</td>
<td>M / 61</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>13</td>
<td>1245</td>
<td>M / 39</td>
<td>MTC</td>
<td>Metastasis</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>14</td>
<td>1500</td>
<td>M / 76</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>15</td>
<td>1407</td>
<td>F / 76</td>
<td>MTC</td>
<td>Primary</td>
<td>918</td>
<td>AwD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>16</td>
<td>1500</td>
<td>M / 43</td>
<td>PTC</td>
<td>Primary</td>
<td>f</td>
<td>AwoD</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
</tbody>
</table>

**Materials and methods**

**Patients**

Fifteen thyroid tumor samples from 14 patients operated on for sporadic MTC were snap frozen in liquid nitrogen after removal and stored at −70°C until analysis. The patients (7 women and 7 men) had an average age of 55 years at diagnosis (Table 1). Twelve primary tumors and three metastases (nos 3, 7 and 12 in Table 1) were analyzed. From one of the patients two specimens were used, one from the primary MTC and the other from a metastasis (no. 3 in Table 1).

Specimens of normal thyroid glands from two patients operated on for MTC and one patient operated on for follicular thyroid adenoma (nos 2, 4 and 16 in Table 1), and tumor tissue from one patient operated on for papillary thyroid carcinoma (PTC; no. 15 in Table 1) were obtained and were used as control tissue. The tumors were classified as suggested by the World Health Organisation committee (14). Informed consent was obtained from all patients, and the study was approved by the Ethical Committee of the Karolinska Hospital.

**Preparation of probes**

The following synthetic oligonucleotides (Geneset, Paris, France) were used as probes for the in situ hybridization: 5’-GCCGCACGA AGCTGCGTG AGGCTGGACC GAGGTCTCGT TGCCC-3’ complementary to nt 977–933.
Expression of RET and its ligands in MTCs

of RET (15); 5′-AACCAAGTTTA TCATCTAAA AAA-GCA-3’ for nt 437–389 of GDNF (16); 5′-GGGACTGACG GTATTTTCT CTTTACGACG GCCCTG-3’ for nt 1102–1057 of GFRalpha-1 (17); 5′-GAGAAGCAGGC CCTTTCGACA CATCCAGATG GACAGCACGG-3’ for nt 86–47 of NTN (4); and 5′-AACTTTAACAATCATAAAT TGCAGCCCCCT CTTTTTGGAAA-3’ for nt 2277–2238 of GFRalpha-2 (5). As a positive control 5′-CTACATGGCA ACTGTGAGGA GGGGAGATTC AGTGTGGTGG GGGAC-3’ for nt 1193–1149 of glycerol aldehyde phosphate dehydrogenase (GAPDH) was used. The oligonucleotides were labeled at the 3′ end with α32P-dATP (NEN, Life Science Products, Boston, MA, USA) using terminal deoxynucleotidyl transferase (Amersham Life Science, Tokyo, Japan). The labeled probes were purified using Nensorb-20 columns (Du Pont, Wilmington, DE, USA).

mRNA in situ hybridization

Cryostat sections 6 μm thick were cut and thaw-mounted on SuperFrostPlus slides (Menzel-Glaser, Braunschweig, Germany). mRNA in situ hybridization was performed as follows. Hybridization solution (50% formamide, 4× SSC (1× SSC: 0.15 mol/l NaCl, 0.015 mol/l sodium citrate), 1× Denhardt’s solution (0.02% polyvinylpyrrolidone, 0.02% bovine serum albumin and 0.02% Ficoll), 1% N-lauroyl-sarcosine, 0.02 mol/l phosphate buffer (pH 7.0), 10% dextran sulfate, 500 μg/ml heat-denatured salmon sperm DNA, 200 nmol/l dithiothreitol, and 37.5 μl/ml of the labeled probes) were mixed and placed in a hybridization oven for 45 min. The solution was spread out on the sections, covered with paraffin, placed in a humidified box and incubated for 16–18 h at 42°C. After hybridization, the sections were sequentially rinsed five times (15 min in 1× SSC at 60°C), left at room temperature for 45 min, and then dehydrated with ethanol in increasing concentrations. After air drying, the sections were exposed to Hyperfilm beta-max X-ray film (Amersham, CEA AB, Uppsala, Sweden) for 4–6 days. The sections were then dipped in Kodak NTB-2 emulsion, exposed for 1–4 weeks, developed and finally counterstained with hematoxylin-eosin. The entire procedure was carried out at least twice on each gene (f, forward; r, reverse): RETf (5′-TGCCAG CCC ATACAATTGTG-3’), RETr (5′-TGCCAG CCC ATACAATTGTG-3’), GDNF (5′-GTTCGGGCT GTTGAGCT-3’), GDNFr (5′-CATCGCAAG GCGCTGAGCAG-3’), GFRalpha-II (5′-ATGTTCCCTG CGA CCTGTG-3’), GFRalpha-1r (5′-AGTGGCTCTCG CTGG CCGC-3’), NTNf (5′-AGGGCCGCTG TCTAGGCCCC-3’), NTNr (5′-AGTGGCGGAG CAGACCAGTCG-3’), GFRalpha-2f (5′-ACGAGACCT CCGCTGTTTG-3’) and GFRalpha-2r (5′-GGGAGCTTGC TGAACACTCCTC-3’). PCR amplification was performed for each gene in a total volume of 50 μl containing 1 μl of the synthesized cDNA, 10 nmol/l Tris–HCl (pH 8.3), 50 mmol/l KCl, 0.75–1.5 mmol/l MgCl2, 200 μmol/l dNTP, 0.01% gelatine, 1–2 U AmpliTaq DNA polymerase and 1–2 U of each specific primer. The PCR products were subsequently electrophoresed on 1.4% agarose gels.

Restriction enzyme cleavage of the RET product was performed by incubating 20 μl of the RET PCR product with 20 μl buffer and 2 μl EcoRI restriction enzyme overnight. Twenty microliters of each digestion were electrophoresed on a 1.4% agarose gel.

Results

The expression of RET and its ligand complexes GDNF/GFRalpha-1 and NTN/GFRalpha-2 was studied by mRNA in situ hybridization on frozen tissue sections

Screening for RET mutations

All MTC specimens except nos 6, 12 and 14 (Table 1) had previously been screened for RET mutations in exons 10, 11 and 16 (18), while MTC specimens nos 6, 12 and 14 were screened for specific mutation in the RET codon 918 in this study. Two rounds of PCR were carried out using three different primers for the RET sequence as described previously (19). Twenty microliters of the second PCR product were then incubated overnight with 2 μl RsaI, and the resulting restriction fragments were size-separated in a 6% polyacrylamide gel. If the RET codon 918 mutation is present, an RsaI restriction site is introduced in the second PCR. Thus the mutation can be resolved as an aberrant restriction pattern on the polyacrylamide gel.

Reverse transcription (RT)-PCR

In order to verify the specificity of the mRNA in situ hybridization, RT-PCR was performed for all five genes, and the RET product was then further confirmed by EcoRI cleavage. mRNA was isolated from case nos 5 and 8 using Ultraspec-II RNA isolation system (Biotechnology Laboratories Inc., Houston, TX, USA). cDNA was synthesized from the isolated mRNA with random primers using a First-Strand cDNA Synthesis Kit (Amerhashan Pharmacia Biotech, Uppsala, Sweden). The template mixtures were incubated at 37°C for 1 h. The primers used in the amplification of RET, GDNF and GFRalpha-1 exons have been described previously (20, 21). Primers for NTN and GFRalpha-2 were selected from published sequences and chosen to span exon borders for each gene (f, forward; r, reverse): RETf (5′-TCAACATTGTC CCTGCAAGGAG TC-3’), RETr (5′-TGCCAG CCC ATACAATTGTG-3’), GDNF (5′-GTTCGGGCT GTTGAGCT-3’), GDNFr (5′-CATCGCAAG GCGCTGAGCAG-3’), GFRalpha-II (5′-ATGTTCCCTG CGA CCTGTG-3’), GFRalpha-1r (5′-AGTGGCTCTCG CTGG CCGC-3’), NTNf (5′-AGGGCCGCTG TCTAGGCCCC-3’), NTNr (5′-AGTGGCGGAG CAGACCAGTCG-3’), GFRalpha-2f (5′-ACGAGACCT CCGCTGTTTG-3’) and GFRalpha-2r (5′-GGGAGCTTGC TGAACACTCCTC-3’). PCR amplification was performed for each gene in a total volume of 50 μl containing 1 μl of the synthesized cDNA, 10 nmol/l Tris–HCl (pH 8.3), 50 mmol/l KCl, 0.75–1.5 mmol/l MgCl2, 200 μmol/l dNTP, 0.01% gelatine, 1–2 U AmpliTaq DNA polymerase and 1–2 U of each specific primer. The PCR products were subsequently electrophoresed on 1.4% agarose gels.

Restriction enzyme cleavage of the RET product was performed by incubating 20 μl of the RET PCR product with 20 μl buffer and 2 μl EcoRI restriction enzyme overnight. Twenty microliters of each digestion were electrophoresed on a 1.4% agarose gel.
from 15 MTCs, three normal thyroids and one papillary thyroid carcinoma. The results are illustrated in Fig. 1 and detailed for each case in Table 1 together with the clinical information.

No expression above background level was seen for any of the five genes in the normal thyroid tissues or in the papillary thyroid carcinoma tissue (Table 1). mRNA expression from the positive control gene GAPDH was seen in all normal and tumor specimens (data not shown). RET expression was detected in all MTCs, and for seven of the cases the expression was moderate (+) whilst in eight tumors it was strong (+++) (Table 1). In addition, thirteen of the MTCs showed expression of GDNF, twelve of GFRalpha-1, and nine of NTN and GFRalpha-2 (Table 1). In seven of the eight tumors with a strong RET expression a similarly strong expression of GDNF and GFRalpha-1 was evident (Table 1). Moreover, in five of these, strong expression of NTN and GFRalpha-2 was also detected (Table 1; note that one of these MTCs, no. 7, was not analyzed for NTN and GFRalpha-2).

In all cases where strong expression (+++) of RET, GDNF, GFRalpha-1, NTN or GFRalpha-2 was observed, the signals were found in tumor cells typically located adjacent to the stroma, usually peripheral in the tumor.

**Figure 1** mRNA *in situ* expression analysis of RET, GDNF, GFRalpha-1, NTN and GFRalpha-2 in medullary thyroid carcinoma. (a and b) Expression of RET in case no. 5 is seen in tumor cells adjacent to the stroma (outlined area in (a) is enlarged in (b)). (c) Expression of RET in case no. 13. (d) No expression of RET in normal thyroid tissue. (e-i) Two sets of consecutive sections of case no. 4 showing similar expression distribution for (e) RET, (f) GDNF, and (g) GFRalpha-1, and similar expression of (h) NTN and (i) GFRalpha-2. s, stroma; t, tumor. The original magnifications were (a) ×100, (b and d) ×200, (c, h, i) ×400, (e-g) ×40.
In addition, the strong expression could also be observed within the tumor in areas where stroma and connective tissue were present. There was no difference in this expression pattern depending on whether the in situ hybridization was performed on specimens from primary tumors or metastases. In the three metastases analyzed, all five genes were found to be expressed, and in two of them, RET, GDNF, and GFRalpha-1 were expressed at high levels (+++) (Table 1). In addition, in case 3, RET and its ligands showed a stronger expression in the metastasis compared with the primary tumor (Table 1).

The fifteen MTC tumor specimens were screened for RET mutations (Table 1). All MTCs without expression of any of the RET ligands harbored a somatic codon 918 mutation, as did three of the seven tumors with strong expression, whilst four did not. No significant difference in the level of RET expression was detected between MTCs with and without the codon 918 mutation. Out of the nine mutation-positive MTCs, three showed a strong and six a moderate expression of RET, and in the six mutation-negative cases the RET expression was strong in five and moderate in one (Table 1).

The expression of mRNA from the five genes was verified by RT-PCR (Fig. 2). The samples tested confirmed the mRNA expression for all five genes. In addition, the positive RET expression was confirmed by restriction enzyme cleavage (data not shown), and as a negative control for the RET gene a series of PTC samples were used (data not shown). Products of the predicted size were seen for all genes in the tumors with positive mRNA in situ hybridization expression, but no product of RET was seen in any of the PTC samples.

**Discussion**

The role of RET in the genesis of MEN 2-related tumors is undisputed, as more than 90% of affected members of MEN 2 and FMTC families harbor germline mutations in this proto-oncogene. In addition, somatic RET mutations occur in sporadic MTCs in 25–60% of the cases, mainly affecting codon 918. No mutations in the RET gene have previously been described in parathyroid tumors. However, in the second most common MEN 2-related tumor, pheochromocytoma, somatic mutations of RET have been observed in a subset of tumors. Therefore, a logical step should be to examine elements of the RET ligand complexes, to see if activation of this pathway at a different point has occurred, and as such has contributed to the development of tumors. This is especially important for those MEN 2 families which have no documented RET mutation, and also for sporadic MTCs and other MEN 2-related tumors (11). So far, GDNF is the only RET ligand that has been analyzed in this respect, but GDNF mutations could not be demonstrated in such families, nor in sporadic MTCs or pheochromocytomas (22, 23). Interestingly, pheochromocytomas express RET, but not GDNF/GFRalpha-1 or NTN/GFRalpha-2 (E Edström, unpublished observations), suggesting that for this tumor type, artemin or persephin may be the important RET ligand.

In this study, we have shown that a significant proportion of sporadic MTCs express not only RET, but also GDNF, GFRalpha-1, NTN and GFRalpha-2 mRNA. At least half of the tumors in this series showed a very strong expression of the five genes, and in these cases the expression was preferentially detected in tumor cells adjacent to the stroma.

All MTCs showed at least some expression of RET, in contrast to the papillary thyroid carcinoma and the normal thyroid tissue samples. This might be explained by a lack of C-cells in the analyzed sections, as C-cells normally mostly occur in a centrally located area of the thyroid lobe. In two MTCs no expression of GDNF was demonstrated, three cases lacked GFRalpha-1 expression and for NTN and its ligand GFRalpha-2 no expression was detected in six of the cases. These MTCs harbored a somatic codon 918 mutation, which has been shown to be sufficient to trigger tumor development without the involvement of ligand stimuli (24). However, some tumors with strong expression of all five genes also had the 918 mutation. While signaling through wildtype RET is mediated by ligand complexes, signaling through mutated RET in cells from neuroepithelial origin give rise to phosphorylation and transformation independent of ligand stimulation. Neuroepithelial cells with a typical MEN 2A mutation in RET codon 634 show a higher biological activity than cells with the MEN 2B mutation in codon 918. Nevertheless, when a ligand is added to MEN 2B cells, the activation of RET increases plentlyfold and both types of mutated cells are able to induce cell differentiation, in contrast to cells with wildtype RET signaling (12, 13). Previous studies have shown that a heterogeneous distribution of the 918 mutation may occur within MTC tumors. Thus, large parts of some MTCs in which a codon 918 mutation has been detected may consist of cells with wildtype RET (25). It would be of interest to determine whether the expressions of GDNF, GFRalpha-1.
NTN and GFRalpha-2 in the tumors with a 918 mutation in this study occur in tumor areas without a 918 mutation. This could be achieved by microdissection of areas with high expression, followed by PCR amplification and Rsal restriction analysis of the 918 mutation.

The distribution of RET expression and its ligand complexes is striking: in seven of the specimens the expression was found preferentially in tumor cells peripheral in the tumor formations, adjacent to stroma and connective tissue (Fig. 1). These results are in contrast to the findings by immunohistochemical analysis, where RET protein expression was seen in the same MTC subpopulations that also harbored a RET mutation (26). This discrepancy may have several possible explanations, of which the most likely is that the mutated RET leads to increased stability or over-expression of the protein, but not necessarily to increased RNA levels.

A similar distribution pattern has been shown in neuroblastomas, a tumor also derived from the neural rich areas (27). This peripheral location of increased RNA levels.

expression of the protein, but not necessarily to RET the mutated RET expression was seen in the same MTC subpopulations that also harbored a RET mutation (26). This discrepancy may have several possible explanations, of which the most likely is that the mutated RET leads to increased stability or over-expression of the protein, but not necessarily to increased RNA levels.

References

17. Sanciloca M, Hessien CA, Worley DS, Cennillo P, Ehrenfels C, Wallus L et al. Glial cell line-derived neurotrophic factor-dependent RET activation can be mediated by two different cell-surface...
Expression of RET and its ligands in MTCs


Received 4 November 1999
Accepted 11 February 2000