CLINICAL STUDY

Familial growth hormone deficiency with mutated GHRH receptor gene: clinical and hormonal findings in homozygous and heterozygous individuals from Itabaianinha

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Abstract

Objective: To characterize clinically and hormonally the syndrome of autosomal recessive familial growth hormone deficiency (FGHD) recently identified in Itabaianinha, Sergipe, Brazil, caused by a novel mutation (mt) that inactivates the growth hormone-releasing hormone receptor (GHRH-R) gene.

Design: Clinical and hormonal evaluations were performed in 21 FGHD individuals (mt/mt group) aged 8 to 63 years, 13 heterozygotes for the GHRH-R mutation (wt/mt group) and 5 homozygotes for the wild type (wt) allele (wt/wt group), identified by genotyping of peripheral blood leukocyte DNA.

Methods: Clinical and hormonal characterization included physical examination and measurement of GH, IGF-I, IGF binding protein-3 (IGFBP-3), cortisol, prolactin, LH, FSH, and free thyroxine (FT4).

Results: Clinical features were consistent with isolated growth hormone deficiency. Height was significantly reduced in the mt/mt group compared with the wt/mt group (mean height standard deviation score (SDS) $\pm$ S.D.: -7.35 ± 1.37 vs -1.84 ± 1.44 respectively, $P < 0.0001$), and the wt/wt group (-1.85 ± 0.81 , $P = 0.0007$). The height of the 13 wt/mt subjects did not differ from the 5 wt/wt individuals. Serum GH, IGF-I, IGF-I SDS, IGFBP-3 and IGFBP-3 SDS were all significantly lower in the mt/mt group than in the wt/mt and wt/wt groups. Two affected children treated with GH for 1 year showed a normal growth response. Serum IGF-I and IGF-I SDS were lower in wt/mt compared with wt/wt group, but did not reach statistical significance. IGF-I and IGF-I SDS correlated inversely with age in wt/mt group.

Conclusions: FGHD due to an autosomal recessive GHRH-R gene mutation leads to marked dwarfism, phenotypically and hormonally indistinguishable from other forms of isolated GH deficiency. Heterozygotes for the GHRH-R mutation appear to have a partial defect in the GH/IGF axis, with no apparent height impairment.

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Introduction

In 1994, newspaper reports first described the existence of a large number of very short individuals ('dwarves') in Itabaianinha. Itabaianinha is a rural area located in the northeastern Brazilian state of Sergipe, 118 km southwest from the state capital, Aracaju. Approximately 32 000 people live in Itabaianinha, of whom 13 000 reside in the urban area, the city of Itabaianinha, with the remaining population spread over a very poorly developed rural area. In 1995, a clinical and genetic study of some families with affected individuals was started that led to the recent identification of a novel homozygous donor splice site mutation (mt) (G to

A at position +1) in intron 1 of the growth hormone releasing hormone (GHRH) receptor (GHRH-R) gene as the cause of familial growth hormone deficiency (FGHD) in this population (1). This report details clinical and biochemical characteristics of subjects who are homozygous (mt/mt) or heterozygous (wt/mt) for the GHRH-R gene mutation (wt, wild-type GHRH-R allele).

Subjects and methods

Subjects

Affected subjects and normal controls from the same families were selected on the basis of height. Written

informed consent was obtained from all subjects. A limited physical examination, including height, was performed, and peripheral venous blood was collected for hormonal measurements and DNA extraction. All affected individuals and the respective families appear caucasian, and some had blond hair. Twenty-one short individuals from 12 families, and 18 unaffected family members were evaluated (Table 1). Standard deviation scores (SDS) of height were calculated based on growth charts published by the World Health Organisation (2). Body mass index (BMI) of 10 affected (mt/mt) and 3 heterozygous (wt/mt) subjects was calculated as weight (kg)/height (m)². BMI percentile for height age (BMI%ile for HA) was calculated based on a published chart (3).

Among the 18 family members with normal stature who served as the control group, 13 were determined to be heterozygous (wt/mt) and 5 to be homozygous for the wild type allele (wt/wt) for GHRH-R gene, via genotyping by denaturing gradient gel electrophoresis (1). Therefore, the control group was divided into two subgroups, wt/mt and wt/wt (Table 1). All the phenotypically affected subjects were confirmed to be homozygous for the mutation.

Hormone measurements

Serum was separated from blood after one hour standing at room temperature, and frozen at -20°C until assay a few days later. Growth hormone (GH) was measured by IRMA (CIS, Gif-Sur-Yvette Cedex, France). Insulin-like growth factor-I (IGF-I) was measured by RIA after acid-ethanol extraction (Nichols Institute, San Juan Capistrano, CA, USA), and insulin-like growth factor-binding protein-3 (IGFBP-3) was measured by IRMA (Active IGFBP-3, DSL 6600, Diagnostic Systems Laboratory, Webster, TX, USA). Serum thyroxine (T4) (RIA) and thyrotrophin (TSH) (IRMA) were measured using commercial kits from DPC, Los Angeles, CA, USA. Prolactin (PRL), cortisol, luteinizing hormone (LH), and follicle-stimulating hormone (FSH), measured by fluoroimmunoassay (Auto Delfia, Wallac, Turku, Finland) were also determined for the same sera.

Treatment

Patients 1 and 3 were treated with daily subcutaneous applications of human recombinant GH (Genotropin, Pharmacia and Upjohn, Sao Paulo, Brazil), 0.1 U/kg body weight/day, for one year.

Statistical analysis

Serum concentrations of IGF-I and IGFBP-3 for each individual were expressed as standard deviation scores (SDS) from the mean to conform to age and sex differences in normal values provided by the manufacturers of the IGF-I and IGFBP-3 kits. The Mann-Whitney

rank sum test was used to compare height, SDS of height, serum concentrations of GH, IGF-I SDS and IGFBP-3 SDS between affected (mt/mt) individuals, heterozygous (wt/mt) and homozygous (wt/wt) normal subjects. Spearman rank order tests were used to evaluate correlations between age, height, SDS of height, serum GH, IGF-I SDS and IGFBP-3 SDS.

Results

Clinical data and serum hormone levels are presented in Table 1. One of the pedigrees indicates a high degree of consanguinity (Fig. 1). Among the 12 nuclear families studied, approximately 40% of the siblings of each proband were affected, ranging from 11% (family of patient 14) to 100% (family of patient 9). There were equal numbers of females ($n=11$) and males ($n=10$). None of the parents were affected in this sample. These observations are consistent with autosomal recessive inheritance of GH deficiency (GHD) and a founder effect (1).

Affected individuals had proportionate short stature, with excessive trunkal fat and poorly developed muscle mass. Normal secondary sex characteristics were present after puberty, which appeared to occur at the normal age. Children and adults had high pitched voices. Small developmental defects such as ear deformities were noted in some of the affected subjects.

All affected individuals were very short. The SDS from the mean height for the 21 affected individuals taken together was -7.35 ± 1.37 (mean $\pm$ s.d.) (range: -9.98 to -4.54) compared with wt/mt (-1.84 ± 1.44 , range: -4.45 to 1.29 , $Z_T=4.84$, $P<0.0001$) and wt/wt (-1.85 ± 0.81 , range: -2.61 to -0.61 , $Z_T=3.43$, $P=0.0007$), with highly significant statistical differences. Mean height SDS for those mt/mt individuals less than 18 years of age was -6.24 ± 0.84 (range: -6.82 to -4.54 , $n=7$) compared with -0.41 ± 1.54 (range: -1.70 to 1.29 , $n=3$) for combined (due to the low number of individuals) wt/mt and wt/wt individuals ($P<0.06$). Affected adult male height (18 years and older) was 123.3 ± 7.62 cm (range: 111 to 135 cm, $n=9$) compared with 163.0 ± 4.53 cm (range: 159 to 170 cm, $n=5$) for their combined adult male wt/mt and wt/wt relatives ($Z_T=3.023$, $P=0.003$). Affected adult females measured 118.8 ± 8.53 cm (range: 105 to 125 cm, $n=5$) compared with 150.8 ± 7.31 cm (range: 137 to 162 cm, $n=10$), for their combined adult female wt/mt and wt/wt relatives ($Z_T=3.003$, $P=0.003$).

BMI%ile for HA was 25 for case 1. All other 9 affected individuals in which this parameter could be determined were at the 75th or above percentile (Table 1). BMI%iles for HA in 3 wt/mt relatives were considered normal (percentiles 50 to 90).

Mean $\pm$ s.d. random serum GH (Fig. 2A) was: (i) for the affected (mt/mt) group: 0.102 ± 0.063 ng/ml (range: 0.006 to 0.211 ng/ml); (ii) for the wt/mt group: 0.377 ± 0.324 ng/ml (range: 0.026 to 1.062 ng/ml)

Table 1 Short stature from Itabaianinha: data from GH deficient subjects and their relatives.

Case	Sex	Age (y)	Height (cm)	Height SDS	Weight (kg)	BMI %ile for height age	GH (ng/ml)	IGF-I (ng/ml)	IGF-I SDS	IGFBP-3 (µg/ml)	IGFBP-3 SDS	Cortisol (µg/dl)	PRL (ng/ml)	LH (IU/l)	FSH (IU/l)	FT4 (ng/dl)
GH deficient																
1	f	8	99	-4.54	14.2	25	0.079	nd	nd	0.53	-4.09	10.0	6.1	0.6	2.3	nd
2	f	10	98	-5.90	nd	nd	0.211	≤19.8	-1.79	0.25	-3.47	25.0	12.0	0.6	2.9	1.56
3	m	11	101	-6.30	nd	nd	0.190	23.7	-2.10	0.17	-3.82	nd	nd	nd	nd	1.33
4	f	14	116	-6.64	23.3	90	0.180	≤19.8	-3.08	0.44	-3.61	nd	nd	nd	nd	1.49
5	f	15	114	-7.09	nd	nd	0.140	≤19.8	-3.08	0.30	-4.38	8.6	2.2	0.6	5.7	1.64
6	f	16	117	-6.82	22.2	75	0.130	≤19.8	-2.82	0.22	-3.78	6.9	3.5	3.4	7.5	1.69
7	f	16	120	-6.37	nd	nd	0.074	≤19.8	-2.82	0.38	-3.63	11.0	3.0	1.8	6.5	1.65
8	m	21	130	-7.12	28.0	75	0.048	≤19.8	-2.82	0.35	-4.21	17.0	6.1	0.6	8.6	nd
9	m	27	125	-7.88	37.0	>95	0.073	≤19.8	-2.69	0.35	-3.44	nd	nd	nd	nd	1.39
10	f	28	124	-6.62	nd	nd	0.104	≤19.8	-2.69	0.30	-3.29	38.0	8.0	7.8	4.4	1.33
11	m	34	130	-7.12	nd	nd	0.009	21.6	-2.67	0.45	-3.52	nd	nd	nd	nd	nd
12	m	35	135	-6.36	34.0	90	0.077	≤19.8	-2.69	0.28	-3.72	7.0	3.2	4.6	8.2	nd
13	m	37	116	-9.25	nd	nd	0.144	≤19.8	-2.69	0.17	-3.85	nd	nd	nd	nd	nd
14	f	37	125	-6.45	36.1	>95	0.079	≤19.8	-2.69	0.26	-3.17	nd	nd	nd	nd	1.24
15	f	39	105	-9.79	nd	nd	0.196	≤19.8	-2.69	0.18	-3.24	nd	nd	nd	nd	1.57
16	m	44	123	-8.19	45.0	>95	0.041	21.6	-2.68	0.48	-5.38	12.0	2.9	1.6	4.2	1.17
17	m	47	111	-9.98	nd	nd	0.175	≤19.8	-2.71	0.27	-5.80	17.0	6.2	5.2	4.4	1.93
18	m	50	118	-8.95	34.6	>95	0.086	≤19.8	-2.71	0.22	-6.25	8.8	5.3	3.0	3.1	nd
19	f	56	116	-7.96	nd	nd	0.045	≤19.8	-2.72	0.22	-6.25	13.0	2.5	34.0	60.0	1.57
20	m	61	122	-8.34	52.0	>95	0.006	≤19.8	-2.72	0.44	-5.75	nd	nd	nd	nd	0.55
21	f	63	124	-6.60	nd	nd	0.045	≤19.8	-2.72	0.43	-5.78	nd	nd	nd	nd	nd
mean		31.86	117.52	-7.35			0.102	20.2	-2.7	0.32	-4.31	14.53	5.08	5.32	9.82	1.44
S.D.		17.28	10.24	1.37			0.063	1.0	0.28	0.11	1.07	9.03	2.86	9.31	15.94	0.32
wt/mt relatives																
22	f	6	121	1.29	nd	nd	0.296	49.4	-1.92	3.57	1.02	18.0	nd	nd	nd	nd
23	f	11	139	-0.83	nd	nd	0.270	279.8	-0.32	4.74	-0.02	nd	nd	nd	nd	1.04
24	f	18	162	-0.28	nd	nd	1.062	163.1	-1.76	0.95	-2.27	nd	nd	nd	nd	1.48
25	f	33	154	-1.61	48.7	75	0.253	127.5	-1.41	3.57	-0.20	11.0	3.2	4.2	3.2	1.43
26	f	35	152	-1.95	52.3	90	0.308	209.1	-0.44	3.94	0.13	8.7	6.7	2.9	5.2	1.93
27	f	35	145	-3.12	nd	nd	0.253	45.9	-2.38	0.90	-2.59	8.0	4.6	1.3	2.9	1.55
28	m	42	165	-1.80	nd	nd	0.171	33.4	-2.49	4.66	3.03	nd	nd	nd	nd	nd
29	m	44	161	-2.41	52.0	50	0.026	33.9	-2.48	2.47	-1.37	2.5	5.9	5.1	4.6	1.39
30	f	44	137	-4.45	nd	nd	0.540	105.8	-1.32	0.83	-4.67	nd	8.7	3.3	4.8	1.73
31	m	48	159	-2.71	nd	nd	0.160	48.8	-2.24	0.92	-4.49	nd	nd	nd	nd	1.29
32	f	61	146	-2.95	nd	nd	1.040	27.8	-2.55	2.48	-1.11	13.0	2.8	7.0	26.0	1.30
33	f	64	151	-2.11	nd	nd	0.140	≤19.8	-2.72	2.73	-0.54	nd	nd	nd	nd	nd
34	m	86	170	-1.04	nd	nd	0.380	21.0	-2.69	0.80	-4.93	11.0	2.5	5.5	10.0	1.10
mean		40.54	150.86	-1.84			0.377	89.6	-1.90	2.50	-1.39	10.31	4.91	4.19	8.10	1.42
S.D.		22.00	13.48	1.44			0.324	82.9	0.82	1.51	2.36	4.77	2.30	1.88	8.23	0.27
wt/wt relatives																
35	f	14	149	-1.70	nd	nd	0.235	382.9	-1.03	4.33	-0.04	nd	nd	nd	nd	1.08
36	f	29	160	-0.61	nd	nd	0.084	226.8	-0.23	3.62	-0.50	16.0	nd	nd	nd	nd
37	f	34	148	-2.61	nd	nd	0.250	115.8	-1.55	2.83	-0.86	nd	nd	nd	nd	1.11
38	f	52	153	-1.78	nd	nd	0.240	182.6	-0.09	4.51	3.52	nd	nd	nd	nd	nd
39	m	53	160	-2.56	nd	nd	0.191	33.2	-2.50	2.10	-1.97	nd	nd	nd	nd	0.93
mean		36.40	154.00	-1.85			0.200	188.3	-1.08	3.48	0.03	16.00	—	—	—	1.04
S.D.		16.44	5.79	0.81			0.069	131.0	0.99	1.02	2.08	—	—	—	—	0.10

Normal values for GH: 0–7 ng/ml; IGF-I (ng/ml), 6–8 y: 88–474; 9–11 y: 117–771 (fem), 110–565 (male); 12–15 y: 261–1096 (fem), 202–957 (male); 16–24 y: 182–780; 25–39 y: 114–492; 40–54 y: 90–360; 55+ y: 71–290; IGFBP-3 (µg/ml), 0–6 y: 1.03–4.25; 7–39 y: 1.25–7.33 (male), 1.8–8.41 (fem); 39+ y: 2.02–7.26; FT4: 0.8–2 µU/ml; nd: not determined. mt: mutated GHRH-R allele; wt: wild type GHRH-R allele.

Figure 1 Genealogy from families of patients 5 and 7 (IV-4 and IV-5 respectively), and 8 (IV-8), showing multiple consanguineous marriages and autosomal recessive pattern of inheritance. Squares, male; circles, female.

($Z_T = 3.58$, $P = 0.0004$); and (iii) for the wt/wt group: 0.200 ± 0.069 ng/ml (range: 0.084 to 0.250 ng/ml) ($Z_T = 2.65$, $P = 0.009$, relative to mt/mt).

Serum GH was less than 0.1 ng/ml in 12/21 affected individuals, and in only one wt/mt (no. 29) and one wt/wt (no. 36) individual. Four wt/mt (nos 28, 29, 31, and 33) and two wt/wt (nos 36 and 39) individuals had serum GH levels that overlapped with the affected group (Table 1). There was an inverse correlation between serum GH and age in the affected group ($r_s = -0.47$, $P = 0.03$), but not in the wt/mt group ($r_s = -0.16$, $P = 0.61$) or in the wt/wt group ($r_s = 0.10$, $P = 0.95$).

Serum IGF-I was undetectable (≤ 19.8 ng/ml) in 17 of 20 affected individuals (serum IGF-I was not measured in individual no. 1), and very low in the remaining 3 (21.6 to 23.7 ng/ml), with a mean value (assuming 19.8 ng/ml for those with ≤ 19.8 ng/ml) of 20.2 ± 1.0 ng/ml (range: ≤ 19.8 to 23.7 ng/ml). By contrast, values in normal stature wt/mt subjects were 89.6 ± 82.9 ng/ml (range: ≤ 19.8 to 279.8 ng/ml) ($Z_T = 4.52$, $P < 0.0001$), and in wt/wt subjects they were 188.3 ± 131.0 ng/ml (range: 33.2 to 382.9 ng/ml) ($Z_T = 3.76$, $P = 0.0008$ relative to mt/mt). Serum IGF-I of the three oldest control individuals, all wt/mt (61, 64, and 86 years of age: 27.8 ng/ml, ≤ 19.8 ng/ml, and 21 ng/ml respectively) overlapped with values observed in the GHD group. Serum IGF-I, expressed as SDS, was -2.68 ± 0.28 , range: -3.08 to -1.79 for the mt/mt group, -1.90 ± 0.81 , range: -2.72 to -0.32 for the wt/mt group ($Z_T = 3.63$, $P = 0.0003$), and -1.08 ± 0.99 , range: -2.50 to -0.09 for the wt/wt group ($Z_T = 3.25$, $P = 0.001$, relative to mt/mt group) (Fig. 2B).

Serum IGFBP-3 was significantly lower in the affected group: 0.32 ± 0.11 μ g/ml (range: 0.17 to 0.53 μ g/ml), compared with the wt/mt group: 2.50 ± 1.51 μ g/ml (range: 0.80 to 4.74 μ g/ml), ($Z_T = 4.85$, $P < 0.0001$), and the wt/wt group: 3.48 ± 1.02 μ g/ml (range: 2.10 to 4.51 μ g/ml), ($Z_T = 3.43$, $P = 0.0007$). IGFBP-3 in the wt/mt group tended to decrease with increasing age, without reaching statistical significance ($r_s = -0.51$, $P = 0.07$), and was below the normal range in five individuals (nos 24, 27, 30, 31, and 34). Serum IGFBP-3, expressed as SDS, was -4.31 ± 1.07 , range: -6.25 to -3.17 for the mt/mt group, -1.39 ± 2.36 , range: -4.93 to 3.03 for the wt/mt group ($Z_T = 3.25$, $P = 0.001$), and 0.03 ± 2.08 , range: -1.97 to 3.52 for the wt/wt group ($Z_T = 3.43$, $P = 0.0007$, relative to mt/mt group) (Fig. 2C). There was an inverse correlation between age of affected individuals and IGFBP-3 SDS ($r_s = -0.45$, $P = 0.04$). IGFBP-3 SDS tended to decrease with age in wt/mt individuals, without reaching statistical significance ($r_s = -0.53$, $P = 0.06$).

Serum random cortisol, PRL, LH, FSH, and free T4 (FT4) were normal, except in subject no. 20, who had a low FT4 due to primary hypothyroidism (Table 1). No statistically significant differences in these parameters were found between mt/mt, wt/mt and wt/wt groups.

After 1 year of treatment with human recombinant GH, 0.1 U/kg/day s.c., subjects nos 1 and 3 (prepubertal during this period) grew 14 cm and 12 cm respectively, without showing a significant decrease of growth velocity during the treatment period.

The height of subjects who were heterozygous (wt/mt) for the GHRH-R gene mutation did not differ

Figure 2 Serum GH (A), IGF-I SDS (B), and IGFBP-3 SDS (C) in subjects with growth hormone deficiency (mt/mt) ($n = 18$, except for IGF-I SDS with $n = 17$), subjects heterozygous for GHRH-R mutation (wt/mt) ($n = 13$) and subjects homozygous for the wild type allele (wt/wt) ($n = 5$). P values (all relative to mt/mt) are: * $P = 0.0004$, ** $P = 0.009$, in (A), * $P = 0.0003$, ** $P = 0.001$ in (B), and * $P = 0.0001$, ** $P = 0.007$ in (C).

significantly from those who were homozygous for the normal type allele (wt/wt), as assessed by SDS for height (-1.84 ± 1.44 SDS, range: -4.45 to 1.29 SDS, vs -1.85 ± 0.81 SDS, range: -2.61 to -1.61 SDS, $Z_T = 0.25$, $P = 0.84$) (Table 1). However, serum levels of both IGF-I and IGFBP-3 were lower in the wt/mt group compared with the wt/wt group, but these differences did not reach statistical significance ($Z_T = 1.54$, $P = 0.14$ respectively $Z_T = 1.24$, $P = 0.24$ respectively), and this was also seen for both IGF-I and IGFBP-3 expressed as SDS $Z_T = 1.63$, $P = 0.11$, and $Z_T = 0.94$, $P = 0.37$ respectively) (Fig. 2B and C).

A statistically significant inverse correlation between serum IGF-I and age was found for the heterozygous (wt/mt) group ($r_s = -0.79$, $P = 0.001$ for IGF-I expressed as ng/ml, and $r_s = -0.73$, $P = 0.005$ for

Figure 3 Correlation (Spearman rank order test) between serum IGF-I expressed as ng/ml (A) and as SDS (B) and age of hetero-zygote cases for GHRH-R mutation (wt/mt).

IGF-I SDS) (Fig. 3), but not for the small wt/wt group ($r_s = -0.90$, $P = 0.08$ for IGF-I expressed as ng/ml, and $r_s = 0.00$, $P = 1.05$ for IGF-I SDS). The IGF-I/IGFBP-3 ratio did not differ between the wt/mt (48.15 ± 49.48 , range: 7 to 171) and the wt/wt (49.00 ± 27.42 , range: 15 to 88) groups ($P > 0.1$). No statistically significant correlation was found between the IGF-I/IGFBP-3 ratio and height SDS or age.

Discussion

GHD is an uncommon condition, occurring with a frequency of 1:4000 to 1:10 000 (7). Familial GHD accounts for only 3 to 30% of GHD (8). Two types of autosomal recessive FGHD involving mutations in the gene for GH have been described (8): (i) FGHD IA is due to deletions in the GH gene leading to absence of GH, with severe dwarfism and frequent development of anti-GH antibodies when exogenous GH therapy is administered; (ii) FGHD IB is due to GH deficiency caused by mutations in the GH gene that lead to defective GH. A clinically indistinguishable picture is caused by a mutation in the GHRH receptor, as recently described in 3 kindreds from the Indian subcontinent (11) and in

the patients we studied (1). FGHD II is transmitted via an autosomal dominant mode of inheritance, due to dominant-negative mutations, and FGHD III is a rare syndrome with X-linked inheritance.

Random serum GH is of limited diagnostic value for GHD. However, GH deficiency was so severe in our cases that basal serum levels of GH, IGF-I and IGFBP-3 were sufficiently low to diagnose GHD. Furthermore, GH stimulation tests (clonidine, insulin induced hypoglycaemia and GHRH) confirmed GHD in other affected individuals from the same genealogy (1). IGF-I in the mt/mt GHD group was distinctly lower than wt/mt and wt/wt non-GHD groups, except for the subjects older than 60 years of age (all three wt/mt), who have low serum IGF-I concentrations similar to the mt/mt subjects (Table 1). IGFBP-3 concentrations also declined with age but less dramatically than IGF-I. Lower serum IGF-I levels with advancing age is not unexpected (12). It is interesting that very low serum IGF-I and IGFBP-3 levels were found in the taller (1.70 m) and older (86 years of age) wt/mt subject here studied (no. 34). Random serum GH did not clearly separate mt/mt, wt/mt, and wt/wt subjects, but it helped to exclude GH resistance (13, 14).

Undernutrition, a condition not infrequent in the Itabaianinha region, could contribute, together with GH deficiency, to the extreme short stature of many of the GH deficient individuals. In support of this hypothesis, mean heights for both wt/mt and wt/wt subjects were also low (mean $\pm$ s.d. of height SDS: -1.84 ± 1.44 and -1.85 ± 0.81 respectively), with one very short wt/mt female individual, no. 30, with -4.45 SDS for height, in this group. Low serum IGF-I in 9 wt/mt subjects (nos 22, 24, 27, 28, 29, 31, 32, 33, and 34) and in 1 wt/wt subject (no. 39) control relatives, as well as low serum IGFBP-3 in 5 of the wt/mt group, could also be related to their nutritional status. However, the normal or even increased weight for height, as indicated by BMI percentiles for height age in all 10 affected individuals in which this parameter was determined, indicates that severe undernutrition is unlikely in this group.

Heterozygous (wt/mt) carriers of the IVSI+1G→A allele were physically similar to their non-carrier (wt/wt) relatives. However, their serum IGF-I and IGFBP-3 levels were lower (the small number of subjects, particularly in the wt/wt group, is likely to have precluded this achieving statistical significance). It is possible that normal free IGF-I, as indicated by similar IGF-I/IGFBP-3 ratio between these two groups, might have been sufficient to compensate for this lower total IGF-I. IGF-I decreased with age in both wt/mt and wt/wt groups, the latter being too small to reach statistical significance. The age-related decrease in IGF-I levels was not greater in the heterozygotes as compared with the small group of subjects homozygous for the wild type allele. However, IGF-I was low in 9 of the 13 heterozygotes (87.5%), mainly in the older individuals (7 out of 9 heterozygotes 35 years or older had low

IGF-I, in contrast to 2 in the 5 younger heterozygous individuals), with very low IGF-I, indistinguishable from the mt/mt group. Furthermore, this inverse correlation with age was maintained when IGF-I was expressed as standard deviation scores from the normal mean values in the wt/mt group but not in the wt/wt group. These data suggest that there might be a relationship between heterozygous state for GHRH-R mutation and the declining IGF-I with age. Further studies with more wt/mt and wt/wt subjects are clearly necessary to test for this hypothesis.

Subjects from the Pakistani kindred heterozygous for a GHRH-R mutation were reported to have 'only slight or perhaps no growth retardation' (10). Their height SDS was very similar to the Itabaianinha individuals (-1.89 ± 1.1 , compared with -1.84 ± 1.44 respectively). Furthermore, as in our case, their serum IGF-I and IGFBP-3 levels seemed to be lower than normal mean values, as well as their GH response to GHRH, L-dopa and clonidine.

Affected GHD individuals or heterozygous GHD gene carriers from elsewhere could have arrived in the region and their numbers could have increased due to geographical and socio-economical features favouring consanguineous marriages, especially given the genetic evidence of a founder effect (1). Considering their caucasian features and light-coloured hair and eyes, there is the possibility that their GHD gene had been introduced during the brief (1630 to 1654) Dutch domination of the Brazilian northeast region, particularly during the attack on Sergipe in 1637, when the territory had only one small town and 8 sugar cane plantations and mills. Therefore, if that gene mutation had occurred prior to the establishment of Itabaianinha, it could be present in other areas of the world where the same population has emigrated.

The observation of a possible partial phenotype (reduced serum IGF-I) in heterozygous subjects, without evidence of an effect on stature, may indicate that optimal somatic growth can be achieved with less than maximal activation of the GH-IGF-I axis. It is possible, however, that lower IGF-I serum levels in adult age may cause changes in organs influenced by GH action (such as bone, adipose tissue and lipid metabolism). A closer evaluation of these parameters in subjects heterozygous for the mutation is currently underway in the Itabaianinha kindred.

Only three other genealogies with familial growth hormone deficiency due to a mutation in the GHRH receptor gene have been described (9–11). All three families, from the Indian subcontinent (from Bombain, India (9), Sindh, Pakistan (10), and Delf, Sri Lanka (11)), had the same G-to-T transversion (GAG → TAG) in codon 72 (exon 3), corresponding to amino acid residue 50 in the mature GHRH receptor protein, which creates a stop codon and predicts a truncated protein at its extracellular domain. Growth hormone deficiency in the Itabaianinha genealogy is due to a different GHRH

receptor mutation, a G-to-A transition of the first base of the 5' splice site at the beginning of intron 1 (IVSI+1G → A), which predicts a retention of intron sequences in the mature mRNA and utilization of a downstream cryptic splice donor site (1). Since both mutations predict severely truncated GHRH receptor proteins, it is not surprising that clinical findings were also similar in respect to the degree of growth impairment and apparent absence of phenotypic changes attributable to some eventual extra-pituitary action of GHRH.

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