CLINICAL STUDY

Amino acid variants of the vitamin D-binding protein and risk of diabetes in white Americans of European origin

Tomasz Klupa1,2,3, Maciej Malecki1,2,3, Linda Hanna1, Joanna Sieradzka3, Jakub Frey3, James H Warram1, Jacek Sieradzki3 and Andrzej S Krolewski1,2

1Research Division, Joslin Diabetes Center, Boston, Massachusetts, USA, 2Department of Medicine, Harvard Medical School, Boston, Massachusetts, USA, 3Department of Metabolic Diseases, School of Medicine, Jagiellonian University, Cracow, Poland

(Correspondence should be addressed to A S Krolewski, Section on Genetics & Epidemiology, Joslin Diabetes Center, One Joslin Place, Boston, Massachusetts 02215–5397, USA; Email: Akrolewski@joslin.harvard.edu)

Abstract

Background: Genetic variants of vitamin D-binding protein (DBP) have been reported to be associated, not only with diabetes, but also with prediabetic traits, in several populations. There are two known polymorphisms in exon 11 of the DBP gene that result in amino acid variants: at codons 416 GAT → GAG (Asp → Glu) and 420 ACG → AAG (Thr → Lys).

Objective: To examine the association of these polymorphisms with diabetes in white Americans of European origin.

Methods: We studied unrelated individuals: 181 with type 1 diabetes, 215 with type 2 diabetes, and 163 healthy controls. Exon 11 was amplified using polymerase chain reaction and the two alleles were determined by digestion with specific endonucleases: HaeIII and StyI, respectively.

Results: At codon 416, Asp/Glu allele frequencies were 45%/55% in patients with type 1 diabetes, 43%/57% in patients with type 2 diabetes, and 46%/54% in controls (χ² = 0.69, 2 d.f., P < 0.71). At codon 420, corresponding Lys/Thr frequencies were 27%/73%, 30%/70%, and 30%/70% (χ² = 1.25, 2 d.f., P = 0.53). Distributions of genotypes at both loci, and the haplotypes defined by the two loci, were also very similar in all groups.

Conclusion: DNA polymorphisms in the DBP gene are not associated with diabetes in white Americans of European origin.

European Journal of Endocrinology 141 490–493

Introduction

There is circumstantial evidence that the vitamin D pathway is involved in the development of diabetes. The β-cells contain a receptor for the 1,25-dihydroxyvitamin D₃ and the concentrations of this steroid in serum may have impact on insulin secretion (1). For example, a correlation between serum concentrations of vitamin D and insulin secretion was described in an elderly Dutch population (2), and in the population of Asian immigrants living in the United Kingdom, supplementation with this vitamin improved insulin secretion (3). Vitamin D also has immunomodulatory properties: it has an important role in inflammatory reactions, and can prevent the development of insulitis in an animal model (4). Recently, associations have been described between allelic variants of the vitamin D receptor gene and type 1 diabetes and impaired insulin secretion in a population at risk of type 2 diabetes (5, 6).

In Pima Indians, linkage has been reported between prediabetic metabolic traits and a few microsatellite markers on chromosome 4q in the region containing the vitamin D-binding protein (DBP) gene (known also as the Gc locus component) (7). DBP is essential for vitamin D cellular endocytosis and metabolism (8), thus variants of the DBP protein may affect the amount of active vitamin D in β-cells and, subsequently, insulin secretion. Two frequent polymorphisms in exon 11 of this gene result in amino acid changes (9): a nucleotide substitution in codon 416, from GAT to GAG (Asp → Glu) and 420 ACG → AAG (Thr → Lys).

The aim of the present study was to examine the associations of amino acid variants at codons 416 and 420 of the DBP gene with type 1 and type 2 diabetes in white Americans of European origin.
Participants and methods

DNA for this study was available from 163 unrelated control individuals with normal glucose tolerance, 181 unrelated patients with type 1 diabetes, and 215 unrelated patients with type 2 diabetes. All individuals were white Americans of European origin, residents of Massachusetts, USA, and were examined at the Joslin Diabetes Center in connection with two current research projects. Both projects were approved by the Human Subjects Committee at the Joslin Diabetes Center.

Individuals with type 1 diabetes were examined as part of a current study of the natural history of microalbuminuria (14). For this study, we selected individuals with insulin-dependent diabetes diagnosed before age 30 years who, at the time of examination, had diabetes for 15 years or more. Individuals with type 2 diabetes were selected from among patients who had diabetes diagnosed between ages 35 and 69 years and participated in a study of the genetics of type 2 diabetes. Diabetes was diagnosed according to WHO criteria (15), and was considered type 2 if the individual was treated with diet or oral agents for at least 2 years after the diagnosis. Non-diabetic controls were recruited from among spouses of patients with type 2 diabetes participating in the study of the genetics of type 2 diabetes. All controls had no clinical history of diabetes and had normal fasting blood glucose evaluated according to current American Diabetic Association (ADA) criteria (16).

DNA was extracted from blood using standard procedures. Specific primers were designed to amplify exon 11 and parts of the flanking introns (forward primer: CAAGTCTTATCACCATCTGT; reverse primer: GCCAAGTTACATACACCCAG) on the basis of the published DBP gene sequence (GenBank accession numbers: L10641, L10642). The total length of the PCR product was 809 base pairs (bp). If the GAG (glutamic acid) variant at codon 416 is present, the expected lengths of HaeIII digestion products are 571 bp and 238 bp. If the AAG (lysine) variant at codon 416 is present, the expected lengths of StyI digestion products are 578 bp and 231 bp.

The PCR reaction was performed in a 96-well plate thermal cycler (PTC-100, MJ Research Inc., Las Vegas, NV, USA). The volume of PCR reaction was 50 μl. Each PCR reaction contained 100 ng genomic DNA, 35 pmol each primer, 200 μmol/l dNTP, 1.0 mmol/l MgCl₂ and 1.0 units TaqGold polymerase (Perkin Elmer, Foster City, CA, USA) with 5.0 μl 10× PCR buffer. We used the following PCR conditions: denaturation at 94°C for 10 min, followed by 39 cycles of denaturation for 1 min at 94°C, annealing at 54°C for 1 min and extension at 72°C for 1 min, with a final extension at 72°C for 10 min. PCR products were digested separately with HaeIII and StyI endonucleases according to the supplier’s recommendation. Restriction fragment length polymorphisms were separated on 2% agarose gel, stained by ethidium bromide and photographed.

Allele frequencies were determined from the genotype frequencies. The Asp/Glu and Thr/Lys variants jointly determine four possible haplotypes (Asp–Thr, Asp–Lys, Glu–Thr, and Glu–Lys). Haplotype frequencies and the linkage disequilibrium coefficient between the two polymorphisms was computed using the EH program (ftp://linkage.rockefeller.edu/software/eh). Under the assumptions of the Hardy–Weinberg equilibrium and random mating, this iterative procedure yields maximum likelihood estimates of the haplotype frequencies in the study population, on the basis of the joint distribution of the two polymorphic markers. The phase is unknown for individuals with doubly heterozygous genotypes, but the conditional probabilities of the alternative haplotype configurations can be calculated from the estimated haplotype frequencies. For markers in significant linkage disequilibrium, the difference between the conditional probabilities may be so large that haplotypes can be assigned to phase-unknown individuals with a high degree of confidence (17). For double heterozygotes in this study (54 controls, 49 type 1 patients, and 76 type 2 patients), the conditional probability of the haplotype pair ‘Asp–Lys/Glu–Thr’ was greater than 0.99, regardless of whether it was estimated in each group separately or in the total group. Therefore, all double heterozygotes were assumed to have this pair of haplotypes. For individuals homozygous at both loci (67 controls, 74 type 1 patients, and 87 type 2 patients) or heterozygous at only one locus (42 controls, 58 type 1 patients, and 52 type 2 patients), haplotypes can be determined directly.

Differences in allele, genotype and haplotype distributions in the study groups were examined by χ² test. Differences in quantitative traits were assessed using ANOVA. Data analysis was performed using SAS Version 6.12 (SAS Institute, Cary, NC, USA).

Results

The clinical characteristics of patients with diabetes and control subjects are summarized in Table 1. Among type 1 patients there were 68 persons with normoalbuminuria and 113 with proteinuria. As their characteristics were similar, only combined data are presented. The group of patients with type 2 diabetes was significantly more obese in comparison with the other groups.

Altogether, 568 individuals were genotyped for the Asp/Glu polymorphism at codon 416 and the Thr/Lys polymorphism at codon 420. Genotypes for both loci were in Hardy–Weinberg equilibrium in all three groups separately and in the total group. The two polymorphisms were in strong linkage disequilibrium (χ² = 388.24, 1 d.f. = 1, P = 2 × 10⁻⁶⁶).

All three groups had similar allele and genotype frequencies (Table 2). When the study groups were stratified by obesity and age at examination, the
similarity remained in all strata (data not shown). To examine whether the risk of diabetes was peculiar to a joint effect of both amino acid variants of DBP, the estimated frequencies of two locus haplotypes (see Methods) were also compared and found to be similar in all study groups (Table 3).

The Asp–Thr haplotype corresponds to the Gc1f electrophoretic variant, which in the homozygous state has been reported to be associated with quantitative prediabetic traits (9, 10). The frequency of homozygotes for this haplotype was similar in all three groups: 3.0% (n = 5) in controls, 3.3% (n = 6) in type 1 patients, and 1.9% (n = 4) in type 2 patients. Thus the frequencies of this variant were similar in all our groups and similar to the published data on white populations (18).

To test the hypothesis that variants of DBP may influence some prediabetic traits, we analysed the fasting plasma glucose and body mass index according to genotypes at both loci in the control group. No difference was found between the carriers of different genotypes (data not shown).

**Discussion**

In contrast to several previous observations, this study showed no association between two amino acid variants of vitamin D-binding protein and the risk of type 1 or type 2 diabetes in a white population (9–13). The following discussion of possible explanations for the discrepant results will deal separately with type 1 and type 2 diabetes.

Type 2 diabetes has a complex etiology. Any abnormalities in insulin secretion that are associated with these polymorphisms in the DBP gene may assume an important role only in the presence of environmentally or genetically determined insulin resistance (19). It is important to note that reported associations with the DBP polymorphism were observed in non-white populations (9–12). There have been no reports of association of the DBP polymorphisms with prediabetic traits or type 2 diabetes in white Caucasian populations. Thus it is possible that the effect of these variants of the DBP gene on the development of type 2 diabetes is peculiar to non-whites. It is interesting that the prevalence of homozygotes for the Asp–Thr haplotype, previously reported to be associated with quantitative prediabetic traits, is about seven to eight times greater in Pima Indians than in white populations (9, 18).

In whites, only one study found an association between the DBP polymorphisms and the risk of type 1 diabetes (13). As the association had only nominal significance and that disappeared after adjustment for multiple comparisons, the finding may have been a false positive (type 1 error).

Finally, the shortcomings of our study design should be considered. Although our study had enough power to detect a large or moderate impact of these polymorphisms on the risk of diabetes, it was too small to

---

**Table 1** Clinical characteristics according to study group.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Non-diabetic controls</th>
<th>Type 1 diabetes patients</th>
<th>Type 2 diabetes patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>163</td>
<td>181</td>
<td>215</td>
</tr>
<tr>
<td>Females (%)</td>
<td>57.4</td>
<td>53.0</td>
<td>47.9</td>
</tr>
<tr>
<td>Age at examination (years)</td>
<td>52.55 ± 17.97</td>
<td>36.2 ± 7.1</td>
<td>62.3 ± 5.4</td>
</tr>
<tr>
<td>Age at diagnosis (years)</td>
<td>10.9 ± 6.2</td>
<td>9.7 ± 6.2</td>
<td>49.7 ± 6.2</td>
</tr>
<tr>
<td>Duration of diabetes (years)</td>
<td>25.4 ± 7.2</td>
<td>24.9 ± 4.29</td>
<td>31.74 ± 5.1</td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
<td>25.97 ± 4.96</td>
<td>24.96 ± 4.29</td>
<td>31.74 ± 5.1</td>
</tr>
<tr>
<td>Receiving insulin (%)</td>
<td>100.0</td>
<td>61.7</td>
<td>61.7</td>
</tr>
</tbody>
</table>

All data are expressed as means ± s.d. n, number of individuals. The high proportion of insulin-treated type 2 diabetes patients was related to the relatively long mean duration of their diabetes.

---

**Table 2** DBP allele and genotype distributions in non-diabetic controls, and patients with type 1 or type 2 diabetes.

<table>
<thead>
<tr>
<th>Codon 416</th>
<th>Allele</th>
<th>Genotype</th>
<th>( \chi^2 )</th>
<th>( P )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls</td>
<td>149 (45.7)</td>
<td>177 (54.3)</td>
<td>2 d.f.</td>
<td>32 (19.6)</td>
</tr>
<tr>
<td>Type 1 patients</td>
<td>162 (44.8)</td>
<td>200 (55.2)</td>
<td>4 d.f.</td>
<td>41 (22.7)</td>
</tr>
<tr>
<td>Type 2 patients</td>
<td>184 (42.8)</td>
<td>246 (57.2)</td>
<td>4 d.f.</td>
<td>37 (17.2)</td>
</tr>
<tr>
<td>Codon 420</td>
<td>Thr</td>
<td>Lys</td>
<td>( \chi^2 )</td>
<td>( P )</td>
</tr>
<tr>
<td>Controls</td>
<td>229 (70.3)</td>
<td>97 (29.7)</td>
<td>2 d.f.</td>
<td>82 (50.3)</td>
</tr>
<tr>
<td>Type 1 patients</td>
<td>266 (73.5)</td>
<td>96 (26.5)</td>
<td>4 d.f.</td>
<td>95 (52.5)</td>
</tr>
<tr>
<td>Type 2 patients</td>
<td>302 (70.2)</td>
<td>128 (29.8)</td>
<td>4 d.f.</td>
<td>104 (48.4)</td>
</tr>
</tbody>
</table>

Values are number (%).
Table 3 Distribution of haplotypes defined by variants at codons 416 and 420 of DBP as estimated by gene counting (see Methods), according to study group.

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>Controls</th>
<th>Type 1 diabetes patients</th>
<th>Type 2 diabetes patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asp–Thr</td>
<td>52 (16.0)</td>
<td>68 (18.8)</td>
<td>59 (13.7)</td>
</tr>
<tr>
<td>Asp–Lys</td>
<td>97 (29.7)</td>
<td>94 (26.0)</td>
<td>125 (29.1)</td>
</tr>
<tr>
<td>Glu–Thr</td>
<td>177 (54.3)</td>
<td>198 (54.7)</td>
<td>243 (56.5)</td>
</tr>
<tr>
<td>Glu–Lys</td>
<td>0 (0)</td>
<td>2 (0.5)</td>
<td>3 (0.7)</td>
</tr>
</tbody>
</table>

Values are number (%). \( \chi^2 = 6.58 \) with 6 d.f.; \( P = 0.367 \).

detect weak effects. As a case–control study design may give spurious results attributable to population stratification, including negative findings such as were found in this study, additional studies may be necessary to exclude the possibility of type 2 error.

Acknowledgments
This research was supported by NIH grants DK-47475 and DK-36836.

TK and MTM contributed equally to the work.

References