Exposure to exogenous estrogens in food: possible impact on human development and health

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Abstract

There has been increasing concern about the impact of environmental compounds with hormone-like action on human development and reproductive health over the past decades. An alternative but neglected source of hormone action that may be considered in this connection is hormone residues in meat from husbandry animals treated with sex steroid hormones for growth promotion. Treatment of cattle with naturally occurring or synthetic sex hormones may enhance lean muscle growth and improve feed efficiency and is therefore a very cost effective procedure for cattle producers who have used it for decades in some Western countries, including the USA and Canada. The Joint Food and Agricultural Organisation/World Health Organisation (JECFA) and the US Food and Drug Administration (FDA) considered, in 1988, that the residues found in meat from treated animals were safe for the consumers.

We have re-evaluated the JECFA conclusions regarding the safety of estradiol residues in meat in the light of recent scientific data, with special emphasis on estradiol levels in prepubertal children. These levels are needed for estimates of the normal daily production rates of estradiol in children, who may be particularly sensitive to low levels of estradiol.

In our opinion, the conclusions by JECFA concerning the safety of hormone residues in meat seem to be based on uncertain assumptions and inadequate scientific data. Our concerns can be summarized as follows. 1) The data on residue levels in meat were based on studies performed in the 1970s and 1980s using radioimmunoassay (RIA) methods available at the time. The sensitivity of the methods was generally inadequate to measure precisely the low levels found in animal tissues, and considerable variation between different RIA methods for measuring steroids exists. Therefore the reported residue levels may be subject to considerable uncertainty. 2) Only limited information on the levels of the various metabolites of the steroids was given despite the fact that metabolites also may have biological activity. 3) Reliable data on daily production rates of steroid hormones were and are still lacking in healthy prepubertal children. This lack is crucial as previous guidelines regarding acceptable levels of steroid residues in edible animal tissues have been based on very questionable estimates of production rates in children. Thus, even today the US FDA bases its guidelines on the presumably highly overestimated production rates in prepubertal children given in the JECFA 1988 report. 4) The possible biological significance of very low levels of estradiol is neglected.

In conclusion, based on our current knowledge possible adverse effects on human health by consumption of meat from hormone-treated animals cannot be excluded.

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In this review, we have focused on the possible effects of estradiol as estrogens are the most potent of the sex steroids and the most widely used growth promoters in cattle. As some of our comments and points may also be relevant for the other sex steroids, these are also considered when appropriate.

**Residues of sex steroids and their metabolites in edible tissues from treated animals**

The data on residues in edible tissue after treatment of cattle with hormone releasing implants has been reviewed by the joint Food and Agricultural Organisation/World Health Organisation (FAO/WHO) expert committee on food additives (JECFA) in 1988. Levels found in non-treated (control) and treated cattle (including veal calves, heifers, steers, bulls and pregnant heifers) after different doses and mixtures of naturally occurring sex steroids were considered. The assays used to measure steroid residues in animal tissues were, in general, based on the radioimmunoassay (RIA) method. It should, however, be noted that the residue levels found in both untreated and treated animals were often close to or at (if not below) the detection limit of the assays used. Furthermore, as was also pointed out by JECFA, a large variation existed in the levels obtained with different assays. Thus, the reported levels of sex steroid residues most likely are subject to substantial uncertainty. The general conclusion of the JECFA report was that residue levels of estradiol, estrone, progesterone, and testosterone in animal tissue were all significantly increased (around twofold or more) in treated cattle as compared with untreated calves, heifers and steers. However, the residue levels measured in tissues from treated cattle were all substantially lower than the levels found in untreated pregnant heifers. Thus, residue levels measured in tissues from treated animals fall well within the normal physiological range if the highest levels of testosterone found in mature bulls and the highest levels of estradiol and progesterone found in pregnant heifers were used as references. The significance of increased estrogen levels in non-pregnant treated animals will, however, depend on how small a fraction of the slaughtered cattle is constituted by pregant heifers (especially heifers in late pregnancy, where hormone levels are most elevated). Hence, if hormone implants result in a significant increase in sex steroid residue levels in the edible tissues from the bulk of the slaughtered cattle, then the overall load of sex steroid intake from meat consumption will, nevertheless, be increased. In addition, the practice of treating cattle with sex steroid hormones may contribute to an increased load of these hormones not only through consumption of meat from treated animals, but also via an increased excretion of these compounds and their metabolites to the environment where they, theoretically, may end up in the drinking water.
Endogenous production of sex steroids in humans, especially in prepubertal children

Estradiol, progesterone and testosterone are substances that are naturally occurring in both human and animals in identical molecular forms. However, the major metabolites of these compounds may differ in different species, as the metabolism can be species specific. The naturally occurring sex steroids that are taken up by consumption of animal tissue will, therefore, have the same biological activity in the human body as the endogenously produced hormones. Whether exogenous steroids have a significant impact on the biological processes in the body presumably depends on the exogenous doses supplied in relation to the endogenous production of steroid.

In the US Food and Drug Administration’s (FDA) guidelines for compounds used in food-producing animals, the conclusion regarding the use of natural sex steroids is: ‘...that no physiological effect will occur in individuals chronically ingesting animal tissues that contain an increase of endogenous steroid equal to 1% or less of the amount in micrograms produced by daily synthesis in the segment of the population with the lowest daily production. In the case of estradiol and progesterone, prepubertal boys synthesize the least; in the case of testosterone, prepubertal girls synthesize the least.’ (5).

From this it is clear that the limit of an acceptable increased exposure to naturally occurring sex hormones by consuming treated animal tissue suggested by FDA is based on an estimate of the endogenous daily production of these steroids in prepubertal boys and girls. Estimates of daily production rates for estradiol and progesterone in prepubertal boys and for testosterone in prepubertal girls are given in the guidelines and correspond to 6 μg/day estradiol and 150 μg/day progesterone in boys and 32 μg/day testosterone in girls. Identical production rates were also given in the ‘Thirty-second report on the joint FAO/WHO expert committee on food additives: Evaluation of certain veterinary drug residues in food’ (6).

Daily production rates (PR) of hormones are calculated from estimates of the metabolic clearance rate (MCR, the volume of serum that can be cleared of a given hormone during 24 h) and of the plasma concentrations of the given hormone by the following equation:

\[
PR (\mu g/day) = \text{plasma concentration (\mu g/ml)} \times \text{MCR (ml/day)}
\]

However, to our knowledge, estimated MCRs for estradiol, testosterone or progesterone in healthy prepubertal children have never been published and most likely do not exist (due to questionable ethics in performing such studies, which would involve injection of radioactive hormones in healthy children). In the late 1960s and the 1970s, several papers were published giving MCRs for sex steroids and some of their metabolites in adult men and women (7–12). MCRs have been published for testosterone in prepubertal children with congenital adrenal hyperplasia (13) and in two prepubertal individuals with androgen resistance (11). However, in both these conditions the endogenous hormone milieu is highly abnormal due to the disease state and the MCRs obtained in these prepubertal patients cannot be considered to reflect MCRs in healthy prepubertal children.

Due to the lack of information on MCRs for estradiol, progesterone and testosterone in prepubertal children, it is obscure how the PRs stated in the Thirty-second report on the joint FAO/WHO expert committee on food additives: Evaluation of certain veterinary drug residues in food have been constructed. The papers that the report refers to regarding the PR of estradiol (14) and progesterone (15) in prepubertal boys, and the PR of testosterone in prepubertal girls (11, 13) do not give any information on the MCRs or PRs in healthy prepubertal children. They do, however, give information on the plasma concentrations of these hormones in children. PRs for estradiol, progesterone, and testosterone in prepubertal children are, nevertheless, stated in the JECFA report and seem to be calculated on the basis of prepubertal plasma levels multiplied with MCRs obtained in adults. Thus, multiplying for example the serum concentration of estradiol in prepubertal boys given in (14) with the MCR for estradiol in adult women given in (9) gives exactly the PR value for estradiol in prepubertal boys, 6.5 μg/day, which is stated in the JECFA report (Table 1).

It is highly questionable whether MCRs for sex steroids defined in adults can be used to calculate PRs in children. MCR of sex steroids are influenced by several factors (for reviews see 16, 17). The MCR is related to body size and should be expressed as a function of surface area to facilitate comparison (7). Using a MCR estimated in adults to calculate a PR in children without correcting for the difference in body size may introduce an approximately 2- to 3-fold overestimation of the PR as the body surface area of a 3- to 4-year-old child is approximately 30–50% of an adult woman. Two other factors which are known to influence the MCR of sex steroids are the binding capacity of serum transport protein and the enzymatic activity in the metabolizing tissues (mainly liver). Both these factors are partly affected by the endogenous hormonal status, including the levels of the sex steroids themselves as well as other hormones such as thyroid hormones. The bulk of both testosterone and estradiol in plasma are bound with high affinity to sex hormone-binding globulin (SHBG, previously also called testosterone-binding globulin or TeBG). Since, in general, only the free fraction of steroid is available for metabolic clearing, the MCR is inversely related to the fraction of the hormone bound to SHBG (18) (for review see also 16). In other words, the higher
the levels of SHBG, the higher the binding capacity in plasma and the lower the MCR. After steroids are extracted from plasma by the liver, they are metabolized in the microsomes. The effectiveness by which they are metabolized is determined by the activity of the enzymes involved, including reductase activity. Both thyroid hormone and androgens have been shown to increase reductase activity. The MCR of sex steroids is generally significantly higher in adult men compared with women; presumably due both to the lower plasma levels of SHBG and the higher reductase activity in the hepatic microsomes in men. Prepubertal children have substantially higher SHBG plasma levels than adults (Table 2), and therefore can be expected to have significantly lower MCRs for sex steroids than adults, even when MCRs are corrected for differences in body size. Less is known about the activity of the microsomal enzymes involved in steroid metabolism in children as compared with adults. However, there are no data suggesting that the enzyme activity should be higher in prepubertal children than in adults.

All these data strongly indicate that the MCRs of sex steroids in healthy prepubertal children can be expected to be substantially lower than in adults. A qualified guess would be that, for example, the MCR of estradiol in prepubertal children is at least 2 to 4 times lower than the MCR estimated in adult women (due to correction for differences in body size and SHBG levels).

The other factor in the calculation of the daily production rate of a given hormone is the plasma concentration of the hormone in question. The plasma levels of testosterone and estradiol in prepubertal children are most often below or close to the detection limit of the assays that have been used during the past decades. Problems in measuring low levels of steroids precisely have been recognized by studies showing high variation in the concentrations obtained between different assay methods and different laboratories (19–21). Thus, levels of sex steroids measured in prepubertal children can be regarded as relatively uncertain estimates. It should also be noted that serum levels of both testosterone and estradiol seem to increase slightly with age, even before the onset of puberty. Thus, in a cross-sectional study of more than 200 prepubertal children (all in Tanner stage I), we found that with increasing age an increasing fraction of the children had levels of estradiol and testosterone above the detection limits of the respective assays (Table 3) (22 and A-M Andersson unpublished data). The lowest serum sex steroid levels can therefore

### Table 1

Examples of estimates of daily production rates (PR) of estradiol based on various published data on estradiol serum concentrations in prepubertal boys and a metabolic clearance rate (MCR) estimated in women.

<table>
<thead>
<tr>
<th>Source</th>
<th>Assay</th>
<th>Boys in pubertal stage I</th>
<th>PR</th>
<th>MCRa</th>
<th>Age in years (mean ± s.d.)</th>
<th>n</th>
<th>pg/ml (mean ± s.d.)</th>
<th>PRb</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bidlingmaier et al.</td>
<td>RIA</td>
<td>7.15</td>
<td>27</td>
<td>7.5</td>
<td>1350d</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(40)</td>
<td></td>
<td></td>
<td></td>
<td>± 2.8</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Angsusingha et al.</td>
<td>RIA</td>
<td>11.2 ± 0.4</td>
<td>6</td>
<td>4.8</td>
<td>1350d</td>
<td></td>
<td></td>
<td>6.5</td>
</tr>
<tr>
<td>(14)</td>
<td></td>
<td></td>
<td></td>
<td>± 1.5</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ducharme et al.</td>
<td>RIA</td>
<td>6–8</td>
<td>9</td>
<td>23</td>
<td>1350d</td>
<td></td>
<td></td>
<td>2.6</td>
</tr>
<tr>
<td>(41)</td>
<td></td>
<td></td>
<td></td>
<td>± 2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baker et al.</td>
<td>RIA</td>
<td>8.5 ± 3.4</td>
<td>9</td>
<td>5.3</td>
<td>1350d</td>
<td></td>
<td></td>
<td>7.2</td>
</tr>
<tr>
<td>(42)</td>
<td></td>
<td></td>
<td></td>
<td>± 3.2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Klein et al.</td>
<td>RCBA</td>
<td>9.5 ± 2.1</td>
<td>23</td>
<td>0.08</td>
<td>1350d</td>
<td></td>
<td></td>
<td>2.9</td>
</tr>
<tr>
<td>(23)</td>
<td></td>
<td></td>
<td></td>
<td>± 0.2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Note:**
- MCR in adult women, from Longcope et al. (9).
- PR calculated as serum concentration (pg/ml) · MCR (ml/day).
- Only the mean is given in the paper.
- MCR in adult women not corrected for the difference between body surface area in adult women and children.
- MCR in adult women corrected by a factor of 0.4 for an approximate difference between body surface area in adult women and children.
- The production rate for estradiol in prepubertal boys given in the JECFA report (4).

### Table 2

Mean SHBG concentrations in healthy prepubertal children and adults.

<table>
<thead>
<tr>
<th>Prepubertal boys (Tanner 1, 6–13 years)</th>
<th>Prepubertal girls (Tanner 1, 6–12 years)</th>
<th>Adult men (18–40 years)</th>
<th>Adult women (18–40 years)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (±S.D.) (nmol/l)</td>
<td>Mean (±S.D.) (nmol/l)</td>
<td>Mean (±S.D.) (µg/l)</td>
<td>Mean (±S.D.) (µg/l)</td>
</tr>
<tr>
<td>116 (36.0)</td>
<td>113 (38.4)</td>
<td>35 (15.0)</td>
<td>80 (47.9)</td>
</tr>
<tr>
<td>Number</td>
<td>Number</td>
<td>Number</td>
<td>Number</td>
</tr>
<tr>
<td>139</td>
<td>105</td>
<td>589</td>
<td>452</td>
</tr>
</tbody>
</table>

SHBG concentrations measured in hormone laboratory at Dept. of Growth and Reproduction, Copenhagen University Hospital.
presumably be found in the younger (1- to 6-year-old) non-pubertal children.

Recently, Klein and coworkers (23) developed an ultrasensitive and highly specific assay for measuring estradiol in prepubertal children. Results obtained with this new assay indicated that levels of estradiol in prepubertal boys may be more than 100 times lower than previously published (23). Although the low levels obtained by Klein and coworkers need to be confirmed by other methods, they strongly point to the fact that a substantial uncertainty exists regarding the true serum levels of estradiol in prepubertal children. Thus, not only the MCRs, but most likely also the plasma concentrations used to calculate the PRs of estradiol in prepubertal children stated in the Thirty-second report on the joint FAO/WHO expert committee on food additives: Evaluation of certain veterinary drug residues and reproductive function has long been recognized, although the role of sex steroids for growth, maturation or breast tissue were observed. In boys, significant effect on growth, no effects on vaginal maturation or breast tissue were observed. In girls with Turner syndrome, in which it has been suggested that estrogen has a biphasic dose-response relationship for epiphyseal growth, comes from clinical studies in girls with Turner syndrome, in which it has been suggested that estrogen has a biphasic dose–response relationship for epiphyseal growth. Although this low dose had a significant effect on growth, no effects on vaginal maturation or breast tissue were observed. In boys, short term treatment with a similar dose of infused estradiol has been shown to increase growth velocity.

### Table 3 Fraction (and percent) of prepubertal children with estradiol and testosterone levels above the detection limits in the respective radioimmunoassays at 6–8 and 10–13 years of age.

<table>
<thead>
<tr>
<th></th>
<th>Prepubertal boys</th>
<th>Prepubertal girls</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>6–8 years 10–13 years</td>
<td>6–8 years 10–12 years</td>
</tr>
<tr>
<td>Testosterone</td>
<td>1/31 (3%) 34/56 (61%)</td>
<td>6/35 (17%) 20/33 (61%)</td>
</tr>
<tr>
<td>Estradiol</td>
<td>4/26 (15%) 24/55 (44%)</td>
<td>14/35 (40%) 23/32 (72%)</td>
</tr>
</tbody>
</table>

Testosterone and estradiol levels were measured by direct immunoassays at the Dept. of Growth and Reproduction, Copenhagen University Hospital.

Detection limits: testosterone assay = 66 pg/ml; estradiol assay = 5 pg/ml.
(27). The above mentioned dose of 100 ng/kg/day estrogen (corresponding to 4 μg/day in a 40 kg child) is above our revised PR in prepubertal children and certainly above the estimated increased levels that can be expected from consumption of meat from animals treated according to good animal husbandry practice, especially as ethinylestradiol has a higher oral bioavailability than estradiol-17β (the oral bioactivity of ethinylestradiol and estradiol-17β is approximately 60% and 10% respectively). However, even lower levels of estrogen (25–50 ng/kg/day ethinylestradiol) have been suggested for growth promotion in Turner girls (28). Furthermore, the growth stimulating effect of these doses was observed after a short period of treatment (3–5 weeks).

Little is known about the dose–response relationship in the low dose-long term treatment/exposure situation. We can, however, get some clues from the gender differences in estradiol levels in normal prepubertal children. As previously mentioned, Klein and co-workers, using their ultra sensitive estradiol bioassay, recently were able to show that the levels of estradiol in prepubertal children of both sexes were substantially lower than those that had been determined previously by radioimmunoassay (23). In addition, they showed that the mean serum estradiol level (0.6 ± 0.6 pg/ml) in prepubertal girls (5.5–10.5 years) was significantly greater than the level (0.08 ± 0.2 pg/ml) in prepubertal boys (4.5–13 years). Although this difference is extremely small in absolute figures, prepubertal girls nevertheless have approximately eightfold higher estradiol levels than prepubertal boys.

This gender difference in serum estradiol levels may have biological implications and explain why girls mature faster than boys (23). Due to the biphasic action of estrogen on bone growth and maturation, higher levels of estrogen cause maturation and finally fusion of the epiphyses, which terminates linear growth (28). Thus, the higher childhood levels of estradiol in girls may explain their earlier puberty and their more rapid epiphyseal maturation. Even small changes of estradiol serum levels within the very low prepubertual range may, therefore, have significant biological implications.

Studies on low dose estrogen treatment have additionally shown that different target tissues have different sensitivity for estrogen. Thus, the studies in Turner girls showed that growth is extremely sensitive to the stimulatory effect of estrogen, whereas other effects such as vaginal maturation, breast development, onset of menses, and changes in SHBG levels were observed at higher doses of estrogen. In healthy postmenopausal women, signs of estrogenic stimulation have likewise been observed in some responsive tissues and not in others (19). These studies have shown that while the mean plasma estradiol concentration in these women was about 8 pg/ml (corresponding to 8 pg/g plasma), the concentrations found in vagina, endometrium and myometrium were 198, 655, and 149 pg/g respectively, demonstrating a very efficient accumulation of estradiol in these tissues (29). Breast tumor tissue has also been shown to have an increased concentration of estradiol compared with serum levels (30). Circulating estradiol is presumably not the only source of estradiol in these tissues. As previously mentioned, estrone sulfate, estrone, and estradiol are readily interconvertible. Data on estrogen metabolism in endometrial, vaginal, and mammary tumor tissue indicate that these tissues are very efficient at producing estradiol from precursor estrogens (estrone sulfate and estrone) (for reviews see 19, 31). Thus, circulating estrone sulfate and estrone may be at least as important as estradiol for the final concentration of estradiol in some target tissues. Consequently, evaluation of exposure to these metabolites may be just as relevant as exposure to estradiol itself.

Steroid hormones are implicated in a range of cancers, including breast and cervical cancers in women, prostate and testicular neoplasia in men, and other diseases such as polycystic ovary syndrome and malformations of the external and internal genitalia.

Carcinogenic and teratogenic effects of estrogen have been demonstrated in animal exposure studies and implicated by data from women who were prenatally exposed to diethylstilbestrol (DES). However, the doses of estrogen involved in these animal experiments and the doses of DES that were given to pregnant women were very high and clearly supra-physiological, and the question is whether very low levels of estrogen are totally irrelevant in this respect. In other words – is there a threshold dose for the observed effects of estrogen? As mentioned, the findings in Turner girls demonstrated that tissues have different sensitivities towards estrogen, indicating that specific threshold doses may apply for different tissues. Breast development, cornification of vaginal epithelium and similar indications of estrogen action are all morphological changes. Although biochemical changes (e.g. changes in gene expression, in signal transduction and in cell cycle regulation) may be much more subtle than morphological changes, they are certainly equal if not more important consequences of estrogen action. After all, the morphological changes observed following estrogen action are the result of preceding biochemical changes.

As biochemical changes are more complex to study, our knowledge of the dose–response relationship of estrogens for biochemical changes is more limited. Colleagues from our department have demonstrated specific gene activation by estrogen in the MCF7 breast cancer cell line in experimental in vitro studies using estrogen concentrations as low as 10^{-14} mol/l in the cell medium (H Leffers, unpublished observations). In reality, this concentration corresponds to 10–100 molecules per cell in the culture well, indicating that as little as a few molecules of estrogen are able to exert a
significant and measurable change in a cell's gene expression. Similarly, Masamura and coworkers (32) have shown that estradiol concentrations as low as $10^{-14} - 10^{-13}\text{ mol/l}$ are sufficient to maximally stimulate replication of MCF7 cells with enhanced sensitivity to estradiol due to previous estradiol deprivation. Sheehan and coworkers (33) have recently pointed to the fact that the concept of a threshold level (which is the basis for determining a no observed effect level (NOEL)), may not always hold for exogenous administration of a hormone that has the same action as an endogenously produced hormone. Using estradiol-induced sex reversal of turtle embryos, they showed that if the threshold for estrogen action is already exceeded by endogenous estrogen, no threshold dose exists for the action of exogenous estrogens (33).

What signs then would we expect to observe in a population exposed to excessive but low levels of estrogen based on the above discussion? We may suspect a slightly younger age of pubertal onset and therefore a slight decrease in the age when final height is achieved. In addition, an increase in estrogen-related diseases and cancer forms (mainly breast cancer) would be expected. More subtle biochemical effects may not be recognized except if they lead to explicit adverse effects – and even if or when they do, the original cause (i.e. hormonal action) may not be recognized.

The incidence of breast cancer is increasing in Western countries and has been doing so for several decades. The etiology of cancer is largely unknown and is presumed to involve both genetic preposition and environmental factors. There has also been a decrease in the age of onset of pubertal development in Western countries during this century. At present, this trend is generally attributed to the improved nutritional status. Although isolated signs of estrogenic stimulation (e.g. breast and/or pubic hair development) are not that uncommon in very young girls in the USA (seen at 7 years of age in 27.2% of African-American and 6.7% of white American girls (34)), earlier reliable data are missing in order to determine whether the incidences are increasing. Thus, although great concern regarding the role of environmental compounds with estrogenic activity on the development of hormone-related diseases and wildlife and human fertility has been raised, a clear connection may be extremely difficult to prove based on population data. The relatively few examples of a connection between environmental compounds with estrogenic activity and proven adverse effects on human or wildlife health are based on accidental exposures involving relatively large exposure doses.

A phenomenon which is important to mention in this connection, is the theory of hormonal imprinting. The term 'hormonal imprinting' covers experimental observations, which suggest that influences acting on hormone receptors during a critical period of matura-

### Differences between the action of natural and synthetic steroid hormones

It is important to bear in mind that the considerations discussed in the present paper only apply to natural steroid hormones. The action of the synthetic hormones used in cattle may diverge from endogenous steroids in a number of ways, some of which should be mentioned.

1. Many synthetic sex steroids have a low affinity for SHBG (39) and are present in an unbound form and biologically active in plasma. In contrast, approximately 98–99% of the endogenous sex steroids are bound to SHBG in plasma. Therefore, even though the actual plasma concentrations of the synthetic sex hormones may be similar or even lower than the endogenous steroids, their biological action may be much stronger.

2. Synthetic sex hormones may be metabolized differently and have different half lives to endogenous steroids.

3. Although synthetic sex hormones may have certain effects in common with the endogenous hormones, their action may not be identical in all respects.

The synthetic sex hormones zeranol, melengestrol acetate and trenbolone acetate and their metabolites do not occur naturally in animals and humans. They have, to our knowledge, no legitimate use in humans and consequently all our information of their $in \text{vivo}$ activity is limited to studies in animals. Different effects and consequences of exposure to these hormones may be expected and should be considered.

### Conclusion

We have made a critical re-evaluation of available data on which current recommendations for the administration of certain sex steroids in agricultural practice are based. The data we had access to mainly derived from the thirty-second report of the joint FAO/WHO expert committee on food additives from 1988 and from monographs from the same meeting (4, 6). In this
present paper, the data from 1988 by the FAO/WHO expert committee are re-evaluated in the light of new information coming from recent progress in hormone research.

Several points may flaw the validity of the conclusions of the FAO/WHO expert committee.

1. The data on residue levels in animal tissue presented were based on studies performed in the 1970s and 1980s using the RIA methods available at the time. The sensitivity of the methods was generally inadequate to measure precisely the low levels found in animal tissues, and considerable variation between different RIA methods for measuring steroids exists. Therefore, the reported residue levels may be subject to considerable uncertainty.

2. Only limited information on the levels of the various steroid metabolites was reported despite the fact that this information is very relevant as some of these metabolites also have biological activity.

3. Reliable data on daily production rates of steroid hormones were and are still lacking in healthy prepubertal children. This lack is crucial as previous conclusions and guidelines regarding the acceptable levels of steroid residues in edible animal tissues have been based on very questionable estimates of production rates in children. Thus, even today the US FDA bases its guidelines on acceptable residue levels in tissue from hormone-treated animals on the presumably highly overestimated production rates in prepubertal children given in the JECFA report.

4. The possible biological significance of very low levels of estradiol, especially in children, is largely neglected. In summary, steroid hormones are very potent compounds that have profound biological effects in both animals and humans. Most of our previous knowledge of the biological action of these compounds is based on the effects of adult physiological or supra-physiological doses of hormone. Recent in vitro studies in prepubertal children indicate that even small differences in hormone levels and very low doses of steroid hormone may have significant biological effects. Likewise, recent in vitro research has shown significant molecular effects of estrogen in extremely low doses. Special concern should be addressed to the lasting effects that may occur during specific sensitive time points of development, mainly during the fetal, perinatal and pubertal periods. Thus, it seems more and more evident that in practice no threshold level exists regarding the action of exogenous estrogen (and perhaps other steroid hormones as well), thus emphasizing the potent role of steroid hormones in regulating developmental events and basic functions by acting at the molecular level.

In the light of recent progress in our understanding of estrogen levels in children, we conclude that possible adverse effects on human health by consumption of meat from estrogen-treated animals cannot be excluded.

Acknowledgements

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