The role of tyrosine kinase in gonadotropin-induced ovulation in the rat ovary

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Abstract

Objectives: Our purpose was to elucidate the involvement of the tyrosine kinase pathway in gonadotropin-induced ovulation in the rat ovary.

Study design: We investigated the effect of a tyrosine kinase inhibitor, tyrphostin, on the rat ovulatory process in vivo and in vitro.

Methods: In cultured rat granulosa cells, the effect of tyrphostin on LH-, dibutyryl cyclic AMP ((Bu)2cAMP)- or forskolin-stimulated tissue type plasminogen activator (tPA) activities was examined by using a fibrin autography technique. In an in vivo system, tyrphostin was injected into the bursal cavity of the ovary in pregnant mare serum gonadotropin-treated rats, just before human chorionic gonadotropin administration. After 24 h, the number of oocytes in the oviduct was counted and the tyrphostin-treated ovaries were examined histologically.

Results: Tyrphostin inhibited LH-stimulated tPA activity but did not affect (Bu)2cAMP- or forskolin-stimulated ones. In an in vivo study, tyrphostin suppressed oocyte release dose-dependently. Histological observations revealed that tyrphostin-treated ovaries contained many large unruptured follicles and a few corpora lutea.

Conclusion: This study suggests that the suppressive effect of tyrphostin on ovulation may be partly due to tPA activity inhibition in the granulosa cells via the suppression of tyrosine kinase activity. Additionally, tyrosine kinase phosphorylation may be involved in gonadotropin-activated signaling systems in the rat ovulatory process.

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Introduction

Luteinizing hormone (LH)/chorionic gonadotropin (CG) regulates ovarian functions, including functional differentiation of granulosa cells, ovulation and corpus luteum formation (1). It is well established that cAMP-dependent protein kinase acts as an intracellular signaling transduction system of LH/human CG (hCG) actions (2). LH also increases phosphoinositide turnover and activates protein kinase C that may modulate gonadal functions (2–4). In recombinant LH receptor expressing L-cells, higher concentrations of LH could lead to stimulation of phospholipase C and result in the formation of inositol phosphate and the elevation in Ca2+ (5). We previously reported that a protein kinase C inhibitor, H-7, could suppress gonadotropin-induced ovulation (6). Recently, it was reported that a tyrosine kinase inhibitor suppressed follicle-stimulating hormone (FSH)-dependent differentiation in cultured granulosa cells (7, 8). Costrici et al. (9) have reported that FSH effects on aromatase activity involve the activation of cytosolic soluble protein tyrosine kinases in human granulosa cells. According to Morris & Richards (10), the luteinization of granulosa cells that is stimulated by the ovulatory surge of LH is dependent on both the increased intracellular cAMP and the activation of tyrosine kinases. Taking these findings together, it is possible that LH stimulates multiple intracellular signaling systems.

Tissue type plasminogen activator (tPA), which has been shown to be secreted by granulosa cells, may play an important role in the ovulatory process (6, 11–13). In the ovary, tPA activity has been modulated by gonadotropins (14), cAMP derivatives (15), gonadotropin-releasing hormone (GnRH) (16), phorbol ester (17), and epidermal growth factor (EGF) (18). These factors presumably act through different intracellular signaling systems. Since it is well known that EGF stimulates tyrosine kinases, it is suggested that the activation of tyrosine kinases may be involved in the induction of tPA activity in granulosa cells.

In the present study, to elucidate the involvement of the tyrosine kinase pathway in gonadotropin-induced ovulation, we have studied the effect of a tyrosine kinase
inhibitor, tyrphostin, on LH-stimulated tPA activity in cultured rat granulosa cells. Furthermore, we have studied the in vivo effect of tyrphostin on oocyte release in pregnant mare serum gonadotropin (PMSG)-treated rats by using the ovarian intrabursal injection technique.

**Materials and methods**

**Hormones and reagents**

McCoy’s 5a medium, penicillin–streptomycin solution, and trypan blue stain were obtained from Gibco (Santa Clara, CA, USA). Ovine LH (oLH) (NIH-oLH-26; 2.3 U/mg; FSH contamination <0.5% by weight) was obtained from the National Hormone and Pituitary Distribution Program (National Institute of Diabetes, Digestive and Kidney Diseases, National Institutes of Health, Bethesda, MD, USA). Halothane was purchased from Hoechst Japan Co. (Tokyo, Japan). Tyrphostin, dibutyryl cAMP ((Bu)²cAMP), forskolin (FSK), GnRH, mouse EGF, thrombin, androstenedione, PMSG, and hCG were purchased from Sigma Chemical Co. (St Louis, MO, USA). Fibrinogen was purchased from Calbiochem (La Jolla, CA, USA). Ammonium persulfate, N,N-methylene bis-acrylamide, sodium dodecyl sulfate (SDS), and Coomassie Brilliant Blue were purchased from Bio-Rad (Richmond, CA, USA). Seaplaque agarose was purchased from Sigma Chemical Co. (St Louis, MO, USA). Glu-plasminogen was obtained from Bio-Pool Co. (Umea, Sweden).

**Animals**

Intact immature female Sprague–Dawley rats (21 days old) were purchased from Kiwa Laboratory Animals Co. Ltd (Wakayama, Japan). Animals were housed three to five per cage with free access to food and water and with lights on from 0600–2000 h.

**Granulosa cell culture**

The animals (23 days old) were injected with 20 IU PMSG to initiate follicular development. After 48 h, the rats were killed by cervical dislocation. The ovaries were removed and were punctured with a 26 gauge needle to obtain granulosa cells. After washing, the cells were stained with 0.25% trypan blue, and viable cells were counted with a hemocytometer. The average viability of the cells was approximately 90%. The cells (5 × 10⁷ viable cells per well) were cultured in Falcon 24-multiwell culture plates (Falcon Plastics, Los Angeles, CA, USA) containing 0.5 ml McCoy’s 5a medium (modified, without serum) supplemented with 2 mmol/l L-glutamine, 100 U/ml penicillin, 100 mg/ml streptomycin sulfate, and 10⁻² mol/l androstenedione. The cells were cultured in a humidified 95% air–5% CO₂ incubator at 37°C. The cells were cultured for 48 h in the presence or absence of tyrphostin at concentrations of 0.3–100 µmol/l with oLH (30 ng/ml), mouse EGF (30 ng/ml), GnRH (30 ng/ml), (Bu)²cAMP (5 mmol/l), or FSK (10⁻⁶ mol/l). Fresh tyrphostin was added at 24 h intervals. Tyrphostin was replenished after 24 h because the cells degraded the reagent. At the end of culture, conditioned media were removed and stored at −40°C in the presence of 0.01% Tween-80 until assayed for tPA activity. Assessment of cell viability by trypan blue exclusion after the culture with tyrphostin (0.3–100 µmol/l) demonstrated no overt toxicity (data not shown).

**SDS-PAGE and fibrin autography technique**

Samples (100 µl) of conditioned medium were adjusted to 2.5% SDS and 0.0125% Coomassie Brilliant Blue and fractionated by SDS-PAGE. SDS-polyacrylamide slab gels were prepared according to the method of Laemmli (19). Electrophoresis was at 50 V for approximately 16 h. tPA activity was assayed by the fibrin autography method (13). After electrophoresis, gels were washed twice in 2.5% (v/v) Triton X-100 in distilled water for 45 min to remove SDS. The fibrin-agarose indicator gel was prepared by combining plasminogen (8 µg/ml) as the zymogen, as well as fibrinogen (2.4 mg/ml) and thrombin (0.5 U/ml) to allow the formation of fibrin as the substrate for plasmin. The electrophoresed SDS slab gel was placed onto the fibrin-agarose indicator gel and incubated at 37°C in a humidified chamber until clear lytic zones were formed, indicating the presence of tPA activity. All experiments using the fibrin autography technique were repeated four times, and a photographic record of representative experiments using darkfield illumination is presented. tPA activity was estimated semiquantitatively by measuring the areas of the lytic zones (height × width), and these results were expressed as the relative ratios between different groups by setting the enzyme level in the control group at 1.0.

**Intrabursal injection technique and oocyte retrieval**

At 27 days after birth, the female rats were injected intraperitoneally with 20 IU PMSG at 0900 h to stimulate follicle development. After 48 h (0900 h on day 29), the rats were anesthetized with halothane. Fur was shaved from the right dorsolateral flank, and the shaved area was rinsed with ethanol. An approximately 1 cm incision was made through the skin and muscle layers, followed by a 5 mm cut into the peritoneum. The fat pad containing the right ovary and oviduct was gently pulled from the abdominal cavity. Forty microliters of tyrphostin dissolved in PBS or vehicle were injected into the bursal cavity of the ovary with a 30 gauge needle attached to a Hamilton syringe (Hamilton Co., Reno, NV, USA). The ovary was gently replaced in the peritoneal cavity. A 5 mm incision was then made over the ovary, and 2.5 ml of PBS was injected into the bursal cavity, and the ovary was pushed back into the peritoneal cavity. The rats were then killed by cervical dislocation. The ovaries were retrieved and were stored at −80°C until assayed for tPA activity.
The peritoneal cavity, and the abdominal wall was closed (20). The left ovary received no treatment and served as a control. After the completion of these procedures, the animals were immediately injected intraperitoneally with 10 IU hCG to induce the ovulation. After 24 h, the animals were killed by cervical dislocation. The ovaries and oviducts were removed from each animal and examined with a dissecting microscope. The oocyte–cumulus mass was removed by making a small incision into the tubal wall through which the mass was extruded. Further flushing of the oviduct and the uterus did not reveal the presence of any additional oocytes. The number of oocytes removed from each side was recorded. Statistical analysis was performed by Student’s t-test.

**Histological examinations**

Treated and untreated ovaries were fixed in Bouin’s solution. The tissue was then dehydrated, embedded in paraffin, and serially sectioned at 4 µm. The sections were mounted on glass slides and stained with hematoxylin and eosin and photographed under a microscope.

**Results**

**Specificity of effect of tyrphostin on tPA activity in rat granulosa cells**

Granulosa cells were obtained from PMSG-treated rats and cultured for 48 h in medium alone (control) or in media with mouse EGF (30 ng/ml), or GnRH (30 ng/ml) without or with tyrphostin (30 µmol/l). EGF, which activates tyrosine kinases as an intracellular signaling system, enlarged the lytic area compared with control and tyrphostin reduced the EGF-stimulated lytic area. GnRH, whose intracellular signaling system is the protein kinase C pathway, also enlarged the lytic area, whereas tyrphostin could not affect the GnRH-enlarged lytic area (Fig. 1).

We also studied the effect of tyrphostin on (Bu)2cAMP- or FSK-stimulated tPA activity in cultured rat granulosa cells in order to examine whether tyrphostin affected the protein kinase A pathway. Although (Bu)2cAMP (5 mmol/l) or FSK (10⁻⁶ mol/l) increased tPA activity, 30 µmol/l tyrphostin did not inhibit the (Bu)2cAMP- or FSK-stimulated tPA activity (Fig. 2). These data indicated that tyrphostin specifically inhibited the tyrosine kinase pathway.

**Effect of tyrphostin on LH-induced tPA activity**

We examined the effect of tyrphostin on oLH-induced tPA activity. Figure 3A shows one representative photographic record of the four separate examinations. oLH markedly enlarged the lytic area compared with control, and tyrphostin (0.3–100 µmol/l) dose-dependently reduced oLH-stimulated tPA activity. Urokinase type plasminogen activator activity was not affected in any treatment groups (data not shown). Figure 3B shows the results of semiquantitative analysis of the four different examinations on tPA activity. Setting the tPA activity in the control group at 1.0, oLH stimulated tPA activity approximately 2.5-fold as much as control, whereas tyrphostin inhibited the LH-stimulated tPA activity in a dose-dependent manner. Tyrphostin alone did not affect the tPA activity (data not shown).
We examined the time course of the effect of tyrphostin on oLH-stimulated tPA activity. oLH stimulated tPA activity by 6 h of culture. The inhibitory effect of tyrphostin on the LH-stimulated tPA activity was found by 6 h of culture and the effect was maintained at 12 h of culture (Fig. 4).

Effect of tyrphostin on ovulation
PMSG-treated immature rats were injected with tyrphostin into the ovarian bursal space. Table 1 shows the number of released oocytes from the treated and untreated ovaries in the different dose groups. Injection of 300 or 3000 μmol/l tyrphostin significantly decreased the number of released oocytes from the treated ovaries compared with that from the contralateral untreated ovaries. PBS alone did not affect the number of released oocytes. The percent inhibition was expressed as the difference in the number of released oocytes between the treated and untreated ovaries divided by the total number of released oocytes from the untreated ovary.

Table 1 Effects of tyrphostin on PMSG–hCG induced ovulation. Results are means ± S.E.M.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>No. of oocytes/ovary (± S.E.M.)</th>
<th>Inhibition (% ± S.E.M.)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Untreated</td>
<td>Treated</td>
</tr>
<tr>
<td>PBS</td>
<td>25.3 ± 3.7</td>
<td>23.8 ± 3.6</td>
</tr>
<tr>
<td>Tyrphostin (μmol/l)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>30</td>
<td>24.2 ± 3.2</td>
<td>22.3 ± 3.7</td>
</tr>
<tr>
<td>300</td>
<td>22.7 ± 3.5</td>
<td>15.5 ± 2.6*</td>
</tr>
<tr>
<td>3000</td>
<td>28.5 ± 3.1</td>
<td>15.5 ± 1.6*</td>
</tr>
</tbody>
</table>

Forty microliters of PBS or tyrphostin (30–3000 μmol/l) were injected unilaterally into the ovarian bursa (n = 6 rats) at the time of hCG administration. The number of released oocytes from treated and untreated control ovaries are shown separately. Percent inhibitions are also shown. Percent inhibition is expressed as the difference in the number of oocytes released between the treated and untreated ovaries divided by the total number of oocytes on the untreated side. *P < 0.01, when compared with untreated ovary. **P < 0.01, when compared with PBS-injected rats.
Administration of 300 or 3000 μmol/l tyrphostin significantly inhibited ovulation compared with the PBS-treated group.

**Histological observations**

Intrabursal injections did not cause any obvious pathological changes in the ovaries, such as vascular thrombosis, infarcts, or hemorrhage. The ovaries treated with PBS contained many corpora lutea and a few large preovulatory follicles (Fig. 5A). In contrast, the ovaries treated with 3000 μmol/l tyrphostin contained a few corpora lutea and many large unruptured follicles (Fig. 5B). In these large unruptured follicles, the oocytes with normal meiotic maturation were entrapped within the follicular antrum. Figure 5C shows one representative luteinized unruptured follicle in a tyrphostin (3000 μmol/l)-treated ovary. The oocytes were entrapped in the antrum, the basal lamina separating the granulosa cells and the theca interna was disintegrated and there were no signs of disintegration of the tunica albuginea.

**Discussion**

In the present study, we investigated a possible involvement of tyrosine kinase phosphorylation in the ovulatory process *in vitro* and *in vivo*. Although quercetin and genistein block not only a variety of tyrosine kinases but also other protein kinases, including cAMP-dependent protein kinases and protein kinase C, tyrphostin seems to be a selective and nontoxic protein tyrosine kinase inhibitor which competes for the substrate site of protein tyrosine kinases and not with ATP (21, 22). Therefore, we used tyrphostin as a tyrosine kinase inhibitor. In cultured rat granulosa cells, tyrphostin could inhibit EGF-stimulated tPA activity, whereas tyrphostin did not have any effects on cAMP-, FSK-, or GnRH-stimulated tPA activity. Taking these findings together, tyrphostin can selectively inhibit tyrosine kinases in rat granulosa cells.

In this study, tyrphostin inhibited LH-stimulated tPA activity in cultured rat granulosa cells in a dose-dependent manner. It is indicated that tyrosine kinase phosphorylation might be involved in LH action.

![Figure 5](https://example.com/figure5.png)  
*Figure 5* Representative photomicrographs of rat ovaries after intrabursal injections. (A) Low-power photomicrograph after injection of PBS. The corpora lutea are observed (magnification × 40). (B) Low-power photomicrograph after injection of tyrphostin (3000 μmol/l). Some large unruptured follicles and no corpus luteum are observed (magnification × 40). (C) High-power photomicrograph after injection of tyrphostin (3000 μmol/l). A luteinized unruptured follicle is present (magnification × 100).
for tPA activity induction in the granulosa cells. On the other hand, it is suggested that in the granulosa cells, the tyrosine kinase activation by LH may be an indirect effect via some factors such as EGF and insulin-like growth factor-I (IGF-I) whose intracellular signaling system is tyrosine kinase phosphorylation. According to LaPolt et al. (23), EGF stimulates tPA activity in rat granulosa cells but IGF-I has no effect on tPA activity. Feng et al. (24) reported that LH/hCG reduced EGF-binding sites in rat granulosa cells. In the present time-course study, both the stimulatory effect of LH on tPA activity and the inhibitory effect of tyrphostin on LH-stimulated tPA activity occurred at the same time and there was no time lag with these effects. Therefore, the stimulation of tPA activity by LH does not seem to be an indirect effect mediated via EGF and IGF-I. Davis (25) has demonstrated that the intracellular C-terminal domain and the three putative intracellular loops of the transmembrane domain of the LH/CG receptor contain numerous motifs possessing serine, threonine, and tyrosine residues, suggesting the potential role for modulation of receptor function by phosphorylation, serine–threonine protein kinases, and tyrosine kinases. Therefore, it is suggested that the effect of tyrphostin on LH-stimulated tPA activity might be partly due to the inhibition of LH-involved tyrosine kinase phosphorylation.

According to the previous studies on ovulation, tumor-promoting phorbol ester (26) and GnRH (27) can induce ovulation without gonadotropins. Some authors have reported that oocyte maturation and gonadotropin-induced follicular rupture may be mediated via a cAMP-dependent pathway (28, 29). On the other hand, Hosoi et al. (30) have reported that cAMP fails to induce ovulation in the absence of gonadotropins. Moreover, a tyrosine kinase inhibitor blocks LH induction of luteinization and of prostaglandin endoperoxide synthase-2 expression in granulosa cells, which is one of the markers of ovulation (10, 31). These data suggest a primary role for cAMP, a supportive but essential role for protein kinase C, and an obligatory role for tyrosine kinases acting in the cascade of luteinization and ovulation. Together with these findings, multiple signal transduction systems seem to be involved in the ovulatory process. To examine the involvement of a tyrosine kinase activation in hCG-induced ovulation in vivo, tyrphostin was intrabursally injected into PMSG-treated rats. Tyrphostin suppressed hCG-induced ovulation in a dose-related manner, suggesting that tyrosine kinase phosphorylation may be involved in the ovulatory process.

Since the actual ovarian concentration of tyrphostin is unknown, it is difficult to relate concentration of tyrphostin in the in vitro studies to those in vivo. In the histological examination, ovaries treated with tyrphostin (3000 μmol/l) did not have any pathological changes such as vascular thrombosis, infarcts, or hemorrhage. Therefore, the effect of tyrphostin does not seem to be pharmacological within the doses used in this study. Although tyrphostin-treated ovaries had healthy follicles and a few normally ruptured follicles and corpora lutea, they also had many large unruptured follicles with normally maturing oocytes histologically. In some of these unruptured follicles, the basal lamina separating the granulosa cells and the theca interna were disintegrated, while the tunica albuginea was intact. These findings are consistent with the reports of Tsafiriri et al. (32) that disintegration of the apical theca externa and tunica albuginea was dependent on tPA. It is suggested that tyrphostin may not affect folliculogenesis and oocyte maturation and may inhibit the dissolution of theca externa and tunica albuginea. On the other hand, Morris & Richards (10) have shown that in the presence of an ovulatory dose of LH, the luteinization of the granulosa cells was blocked by a tyrosine kinase inhibitor, AG18. In our in vivo study, even the ovaries treated with high dose of tyrphostin had luteinized granulosa cells morphologically. Although the reason for this discrepancy is unknown, some factors other than tyrosine kinases may also be involved in luteinization mechanism.

In summary, the results of our in vitro and in vivo studies suggest that the suppressive effect of tyrphostin on hCG-induced ovulation might be partly due to the inhibition of tPA activity in the granulosa cells via the block of tyrosine kinase phosphorylations. In the ovulatory process, gonadotropin action might be mediated not only by a cAMP-dependent pathway and protein kinase C activation but also by tyrosine kinase activation. The mechanism of gonadotropin-related tyrosine kinase activation in the ovulatory process is not yet clear and further studies will be required.

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